

**UNIVERSITY OF GAZİANTEP**  
**GRADUATE SCHOOL OF**  
**NATURAL & APPLIED SCIENCES**

**PRODUCTION OF SUMAC, APPLE, CARROT AND GRAPE**  
**CONCENTRATES BY FREEZE CONCENTRATION**

**M. Sc. THESIS**  
**IN**  
**FOOD ENGINEERING**

**BY**  
**GÜLAY ÖZKANLI**  
**FEBRUARY 2013**

**Production of Sumac, Apple, Carrot and Grape Concentrates by  
Freeze Concentration**

**M.Sc. Thesis**

**in**

**Food Engineering**

**University of Gaziantep**

**Supervisor**

**Ali Rıza TEKİN**

**by**

**Gülay ÖZKANLI**

**February 2013**

©2013 [Gülay ÖZKANLI].

REPUBLIC OF TURKEY  
UNIVERSITY OF GAZIANTEP  
GRADUATE SCHOOL OF NATURAL & APPLIED SCIENCES  
NAME OF THE DEPARTMENT

Name of the thesis: Production of Sumac, Apple, Carrot and Grape Concentrates by Freeze Concentration

Name of the student: Gülay ÖZKANLI

Exam date: 18.02.2013

Approval of the Graduate School of Natural and Applied Sciences

Assoc. Prof. Dr. Metin BEDİR

Director

I certify that this thesis satisfies all the requirements as a thesis for the degree of Master of Science.

Prof. Dr. Ali Rıza TEKİN

Head of Department

This is to certify that we have read this thesis and that in our consensus/majority opinion it is fully adequate, in scope and quality, as a thesis for the degree of Master of Science.

Prof. Dr. Ali Rıza TEKİN

Supervisor

Examining Committee Members

Prof. Dr. Ali Rıza TEKİN

Assoc. Prof. Dr. Abuzer ÇELENKLİ

Assist. Prof. Dr. Bülent BELİBAĞLI

Signature



**I hereby declare that all information in this document has been obtained and presented in accordance with academic rules and ethical conduct. I also declare that, as required by these rules and conduct, I have fully cited and referenced all material and results that are not original to this work.**

**Gülay Özkanlı**

A handwritten signature in black ink, appearing to read 'Gülay', with a long horizontal stroke extending to the right.

## **ABSTRACT**

### **PRODUCTION OF SUMAC, APPLE, CARROT AND GRAPE CONCENTRATES BY FREEZE CONCENTRATION**

**ÖZKANLI, Gülay**

**M.Sc. in Food Eng.**

**Supervisor: Prof. Dr. Ali Rıza TEKİN**

**February 2013**

**71 pages**

Fruit juices contain high amounts of water; so it is beneficial to concentrate them as much as possible to reduce the microbial spoilage and cost of transportation. Various methods are used to produce fruit concentrates. Among them, freeze concentration can be considered as a non-thermal concentration technique. The quality properties of the original juice will not, therefore be altered appreciably after being concentrated.

In this study, sumac, apple, grapes and carrot juices were thawed as a function of time at different temperatures (10 to 25° C) and initial concentrations (1 to 40 °Brix) to see the effect of these parameters on the degree of separation. The present method can be named as “differential freeze concentration” due to its dependency on time.

As expected, the degree of separation decreased with increase in the initial concentration, thawing temperature and time for all juices. The size and pH values of the samples being thawed were also observed to affect the degree of separation. Thus, increase in size and decrease in the pH values of the samples, resulted in increase in degree of separation.

**Key Words:** Freeze concentration, fruit concentrate, fruit juice

## ÖZET

### **DONDURARAK KONSANTRE METODUYLA SUMAK, ELMA, HAVUÇ VE ÜZÜM KONSANTRELERİ ÜRETİMİ**

**ÖZKANLI, Gülay**

**Yüksek Lisans, Gıda Mühendisliği Bölümü**

**Tez Danışmanı: Prof. Dr. Ali Rıza TEKİN**

**Şubat 2013**

**71 sayfa**

Meyve suları yüksek miktarda su içerir; bu nedenle, mikrobiyal bozulma riskini ve nakliye masraflarını azaltmak için bunları mümkün olduğu kadar konsantre etmek gerekir. Meyve sularını konsantre etmek için değişik metotlar kullanılmaktadır. Bunlardan, dondurarak konsantre etme, ısı olmayan metotlardan biri olarak kabul edilmektedir. Orijinal meyve suyunun kalite özellikleri, bu yüzden, elde edilen meyve konsantresinde fazla değişikliğe uğramamaktadır.

Bu çalışmada sumak, elma, üzüm ve havuç suları dondurulduktan sonra, zamana bağlı olarak, farklı sıcaklıklarda ( 10 - 25° C ) ve başlangıç konsantrasyonlarında (1-40 °Brix) tavlana tabi tutulmuş, bu değişkenlerin meyve sularının konsantre edilme derecesine etkileri araştırılmıştır. Zamana bağlı olması nedeniyle, bu metot “diferansiyel dondurarak konsantre etme” metodu diye adlandırılabilir.

Beklendiği gibi, konsantre edilme derecesi, -tüm meyve suları için- başlangıç konsantrasyonu, tavlama sıcaklığı ve zaman arttıkça azalma göstermiştir. Tavlama örneklerin büyüklüğü ve pH değerlerinin de konsantre edilme derecesini etkilediği gözlenmiştir. Böylece, örnek büyüklüğünde artış ile pH değerlerinde düşüş, konsantre edilme derecesinde artışa neden olmuştur.

**Anahtar Kelimeler:** Dondurarak konsantre etmek, meyve konsantresi, meyve suyu

*To my parents...*



## **ACKNOWLEDGEMENTS**

I would like to express my deep and sincere gratitude to my supervisor, Professor Dr. Ali Rıza Tekin for his continuous support and patience throughout my thesis study. His immense knowledge and logical way of thinking have been invaluable to me. His understanding, encouraging, academic and personnel guidance have been helped me in all the time of research and writing of this thesis.

I would like to thank to my committee members, Assoc. Prof. Dr. Abuzer Çelenkli and Assist. Prof. Dr. Bülent Belibağlı for their useful discussions and suggestions throughout the study.

Most importantly, none of this would have been possible without the love and patience of my family. They are always beside me during the happy and hard moments and motivate me.

## CONTENTS

	<b>Page</b>
ABSTRACT.....	v
ÖZET.....	vi
ACKNOWLEDGEMENTS.....	viii
CONTENTS.....	ix
LIST OF FIGURES.....	xi
LIST OF TABLES.....	xiv
CHAPTER 1: INTRODUCTION.....	1
1.1. Concentration of Fruit Juice.....	1
1.2. Evaporative Concentration.....	1
1.2.1. Some Application of Evaporative Concentration in the Food Industry.....	2
1.3. Membrane Concentration (Reverse Osmosis).....	2
1.3.1. Some Application of Membrane Concentration in the Food Industry.....	2
1.4. Freeze Concentration.....	3
1.4.1. History of Freeze Concentration.....	3
1.4.2. Some Application of Freeze Concentration in the Food Industry.....	4
1.4.3. Freeze Concentration Process.....	5
1.4.3.1. Function of Crystallizer.....	6
1.4.3.2. Function of Wash Column.....	6
1.4.4. Freezing techniques.....	8
1.4.4.1. Suspension Freeze Concentration.....	8
1.4.4.2. Progressive Freeze Concentration.....	9
1.4.4.3. Block Freeze Concentration.....	12
1.4.4.4. Eutectic Freeze Concentration.....	13
1.5. Aim of the Present Study.....	13
CHAPTER 2: MATERIALS AND METHODS.....	15
2.1. Materials.....	15
2.1.1. Sumac Juice.....	15

2.1.2. Apple Juice.....	15
2.1.3. Grape Juice.....	15
2.1.4. Carrot Juice.....	15
2.1.5. Water.....	15
2.2. Preparation of Concentrated Juices.....	15
2.3 Variation of pH value of Concentrated Juices with Concentration.....	16
2.4. Titratable Acidity of Juices.....	16
2.5. Measurement of Concentration of Juices.....	16
2.6. Preparation of Juices with Varied Concentration.....	16
2.7. Analysis of Separation of Juice from Ice.....	16
2.8. Statistical Analysis.....	17
CHAPTER 3: RESULTS AND DISCUSSION.....	18
3.1. The Effect of Initial Concentration on Degree of Separation.....	18
3.2. Effect of Concentration on the pH value of Different Juices.....	20
3.3. Titratable Acidity of Different Sample.....	21
3.4. Effect of Different Temperatures on the Concentration of Sumac Concentrates.....	21
3.5. Effect of Different Temperatures on the Concentration of Apple Concentrates.....	27
3.6. Effect of Different Temperatures on the Concentration of Grape Solutions.....	33
3.7. Effect of Different Temperatures on the Concentration of Carrot Solutions.....	39
3.8. Effect of the Size of Frozen Samples on the Degree of Separation.....	45
3.9. Effect of Time on the Concentration.....	53
3.10. Effect of Temperature.....	59
CHAPTER 4: CONCLUSION.....	65
4.1 Suggestion for Future Work.....	66
REFERENCES.....	67

## LIST OF FIGURES

	<b>Page</b>
Figure 1.1. Freeze Concentration Pilot Plant.....	4
Figure 1.2. Freeze concentration system for concentrated juice production.....	7
Figure 1.3. (a) Suspension Freeze Concentration and (b) Progressive Freeze Concentration.....	10
Figure 2.1. The experimental set-up of the system.....	17
Figure 3.1. Variations of concentration with time for sumac concentrates at different temperatures: (a) initial concentration 1 °Brix, (b) initial concentration 2 °Brix, (c) initial concentration 3 °Brix, (d) initial concentration 4 °Brix.....	22
Figure 3.2. Variations of concentration with time for sumac concentrates at different temperatures: (a) initial concentration 5 °Brix, (b) initial concentration 10 °Brix, (c) initial concentration 15 °Brix, (d) initial concentration 20 °Brix.....	24
Figure 3.3. Variations of concentration with time for sumac concentrates at different temperatures: (a) initial concentration 25 °Brix, (b) initial concentration 30 °Brix, (c) initial concentration 35 °Brix, (d) initial concentration 40 °Brix.....	26
Figure 3.4. Variations of concentration with time for apple concentrates at different temperatures: (a) initial concentration 1 °Brix, (b) initial concentration 2 °Brix, (c) initial concentration 3 °Brix, (d) initial concentration 4 °Brix.....	28
Figure 3.5. Variations of concentration with time for apple concentrates at different temperatures: (a) initial concentration 5 °Brix, (b) initial concentration 10 °Brix, (c) initial concentration 15 °Brix, (d) initial concentration 20 °Brix.....	30
Figure 3.6. Variations of concentration with time for apple concentrates at different temperatures: (a) initial concentration 25 °Brix, (b) initial concentration 30 °Brix, (c) initial concentration 35 °Brix, (d) initial concentration 40 °Brix.....	32
Figure 3.7. Variations of concentration with time for grape concentrates at different temperatures: (a) initial concentration 1 °Brix, (b) initial concentration 2 °Brix, (c) initial concentration 3 °Brix, (d) initial concentration 4 °Brix.....	34
Figure 3.8. Variations of concentration with time for grape concentrates at different temperatures: (a) initial concentration 5 °Brix, (b) initial concentration 10 °Brix, (c) initial concentration 15 °Brix, (d) initial concentration 20 °Brix.....	36
Figure 3.9. Variations of concentration with time for grape concentrates at different temperatures: (a) initial concentration 25 °Brix, (b) initial concentration 30 °Brix, (c) initial concentration 35 °Brix, (d) initial concentration 40 °Brix.....	38
Figure 3.10. Variations of concentration with time for carrot concentrates at different temperatures: (a) initial concentration 1 °Brix, (b) initial concentration 2 °Brix, (c) initial concentration 3 °Brix, (d) initial concentration 4 °Brix.....	40

Figure 3.11. Variations of concentration with time for carrot concentrates at different temperatures: (a) initial concentration 5 °Brix, (b) initial concentration 10 °Brix, (c) initial concentration 15 °Brix, (d) initial concentration 20 °Brix.....	42
Figure 3.12. Variations of concentration with time for carrot concentrates at different temperatures: (a) initial concentration 25 °Brix, (b) initial concentration 30 °Brix, (c) initial concentration 35 °Brix, (d) initial concentration 40 °Brix.....	44
Figure 3.13. Variations of concentration with time for sumac concentrates at different sizes: (a) initial concentration 1 °Brix, (b) initial concentration 5 °Brix, (c) initial concentration 10 °Brix.....	46
Figure 3.14. Variations of concentration with time for apple concentrates at different sizes: (a) initial concentration 1 °Brix, (b) initial concentration 5 °Brix, (c) initial concentration 10 °Brix.....	48
Figure 3.15. Variations of concentration with time for grape concentrates at different sizes: (a) initial concentration 1 °Brix, (b) initial concentration 5 °Brix, (c) initial concentration 10 °Brix.....	50
Figure 3.16. Variations of concentration with time for carrot concentrates at different sizes: (a) initial concentration 1 °Brix, (b) initial concentration 5 °Brix, (c) initial concentration 10 °Brix.....	52
Figure 3.17. Variation of concentration versus temperature for 10 °Brix grape concentrate at 0 min.....	53
Figure 3.18. Variation of concentration versus temperature for 10 °Brix grape concentrate at 25 min.....	53
Figure 3.19. Variation of concentration versus temperature for 10 °Brix grape concentrate at 50 min.....	54
Figure 3.20. Variation of concentration versus temperature for 10 °Brix grape concentrate at 75 min.....	54
Figure 3.21. Variation of concentration versus temperature for 10 °Brix grape concentrate at 100 min.....	54
Figure 3.22. Variation of concentration versus temperature for 40 °Brix grape concentrate at 0 min.....	55
Figure 3.23. Variation of concentration versus temperature for 40 °Brix grape concentrate at 15 min.....	55
Figure 3.24. Variation of concentration versus temperature for 40 °Brix grape concentrate at 30 min.....	55
Figure 3.25. Variation of concentration versus temperature for 10 °Brix carrot concentrate at 0 min.....	56
Figure 3.26. Variation of concentration versus temperature for 10 °Brix carrot concentrate at 25 min.....	56
Figure 3.27. Variation of concentration versus temperature for 10 °Brix carrot concentrate at 50 min.....	57
Figure 3.28. Variation of concentration versus temperature for 10 °Brix carrot concentrate at 75 min.....	57

Figure 3.29. Variation of concentration versus temperature for 10 °Brix carrot concentrate at 100 min.....	57
Figure 3.30. Variation of concentration versus temperature for 10 °Brix carrot concentrate at 125 min.....	58
Figure 3.31. Variation of concentration versus temperature for 40 °Brix carrot concentrate at 0 min.....	58
Figure 3.32. Variation of concentration versus temperature for 40 °Brix carrot concentrate at 15 min.....	59
Figure 3.33. Variation of concentration versus temperature for 40 °Brix carrot concentrate at 30 min.....	59
Figure 3.34. First order reaction for 35 °Brix carrot concentrate.....	61
Figure 3.35. Arrhenius plot for 35 °Brix carrot concentrate.....	61

## LIST OF TABLES

	<b>Page</b>
Table 3.1. Concentration of the first drop of sumac juice at different initial concentrations at 10 °C.....	18
Table 3.2. Concentration of the first drop of apple juice at different initial concentrations at 10 °C.....	19
Table 3.3. Concentration of the first drop of grape juice at different initial concentrations at 10 °C .....	19
Table 3.4. Concentration of the first drop of carrot juice at different initial concentrations at 10 °C.....	20
Table 3.5. Variation of pH with concentration for different juices.....	20
Table 3.6. Titratable acidity of different fruit juices.....	21
Table 3.7. Regression analyses for sumac concentrates (initial concentration 1, 2, 3 and 4 °Brix) at different temperatures .....	23
Table 3.8. Regression analyses for sumac concentrates (initial concentration 5, 10, 15 and 20 °Brix) at different temperatures... ..	25
Table 3.9. Regression analyses for sumac concentrates (initial concentration 25, 30, 35 and 40 °Brix) at different temperatures.....	27
Table 3.10. Regression analyses for apple concentrates (initial concentration 1, 2, 3 and 4 °Brix) at different temperatures .....	29
Table 3.11. Regression analyses for apple concentrates (initial concentration 5, 10, 15 and 20 °Brix) at different temperatures.....	31
Table 3.12. Regression analyses for apple concentrates (initial concentration 25, 30, 35 and 40 °Brix) at different temperatures.....	33
Table 3.13. Regression analyses for grape concentrates (initial concentration 1, 2, 3 and 4 °Brix) at different temperatures.....	35
Table 3.14. Regression analyses for grape concentrates (initial concentration 5, 10, 15 and 20 °Brix) at different temperature.....	37
Table 3.15. Regression analyses for grape concentrates (initial concentration 25, 30, 35 and 40 °Brix) at different temperatures.....	39
Table 3.16. Regression analyses for carrot concentrates (initial concentration 1, 2, 3 and 4 °Brix) at different temperatures.....	41
Table 3.17. Regression analyses for carrot concentrates (initial concentration 5, 10, 15 and 20 °Brix) at different temperatures.....	43
Table 3.18. Regression analyses for carrot concentrates (initial concentration 25, 30, 35 and 40 °Brix) at different temperatures.....	45

Table 3.19. Regression analyses for sumac concentrates (initial concentration 1, 5 and °Brix) at different sizes.....	47
Table 3.20. Regression analyses for apple concentrates (initial concentration 1, 5 and °Brix) at different sizes.....	49
Table 3.21. Regression analyses for grape concentrates (initial concentration 1, 5 and °Brix) at different sizes.....	50
Table 3.22. Regression analyses for carrot concentrates (initial concentration 1, 5 and °Brix) at different sizes.....	52
Table 3.23. Activation energy for different concentrated juices.....	62



## **CHAPTER 1**

### **INTRODUCTION**

#### **1.1. Concentration of Fruit Juice**

Concentration can be defined as the removal of excess water to reduce volume and weight. Fruit juices contain large amounts of water. So, fruit juices are generally pasteurized or concentrated after squeezing to enhance storage life. Concentration processes reduce the water content from about 75 – 90% to 25 – 35%. That is, the soluble solid content is increased up to 65 – 75%. The concentration of fruit juices has several advantages like economical packaging, transportation and storage as well as microbiological stability (Ramteke et al., 1993).

Various methods are available to remove water from liquid food products. Different kinds of concentration methods such as evaporative concentration, membrane concentration (reverse osmosis), and freeze concentration can be applied commercially (Thijssen, 1975).

#### **1.2. Evaporative Concentration**

Evaporation is known as the oldest technique for concentration. At first, fruit juice was concentrated by vacuum evaporation in 1920s. The separation principle of evaporation is the difference in volatilities (vapor pressure) with producing a liquid and a vapor phase. Evaporation removes the solvent, generally water, from solution to increase the solute concentration. The liquid flows through a tube while heat is applied to the outside of the tube. The solvent boils and it is separated from the concentrated liquid. Main components of this system are: A feed preheater and a feed distribution system. The feed preheater is used to bring the feed nearly to the boiling point and the feed distribution system is used to distribute the feed among the tubes (Heldman and Lund, 2007).

Evaporative concentration has several advantages: It causes to decrease the water activity and increases the storage stability of foods. The cost of evaporative concentration systems is generally low so this method is still used widely (Ramteke

et al., 1993). On the other hand, evaporation causes loss of volatile substances and aromas of foods. In addition, some foods are heat sensitive so evaporation is not preferred for these type foods (Ramaswamy and Marcotte, 2006a). Moreover; evaporator consumes much energy (Ramtake et al., 1993).

### **1.2.1. Some Application of Evaporative Concentration in the Food Industry**

Evaporation is used for fruit products to concentrate fruit juices. Jams and jellies are produced from fruit juices and purees and pastes are produced from vegetable juices by evaporation, also. In addition, this technique is used for dairy products to concentrate milk, whey and lactose before drying process. It is used for refining sucrose from sugar beet and cane in sugar industry. Salt is produced by evaporation of sea water (Heldman and Lund, 2007).

### **1.3. Membrane Concentration (Reverse Osmosis)**

Reverse osmosis works on the principle of the presence of pressure gradient. The separation is provided by different combined solubilities and diffusivities of species in membrane. This method reduces loss of aroma and flavor so a good quality product is obtained and it consumes low energy (Jiano et al., 2004). In addition, the equipment is not expensive (Cassano et al., 2007). However, high concentrated juice in reverse osmosis system is limited because of high osmotic pressure (Cassano et al., 2007) and the high organic concentration and pH values are not suitable to membranes (Lemmer et al., 2000).

Reverse osmosis was used for desalination by U.S. Department of the Interior, Office of Saline Water in 1953. In 1970s, it was used in the chemical industry e.g. textiles, oil recovery, paint, pulp and paper. The food and biotechnology industries used this technology in the 1980s. This method was then modified to micro-filtration and nanofiltration, so application of membrane technology has been widened (Heldman and Lund, 2007).

#### **1.3.1. Some Applications of Membrane Concentration in the Food Industry**

Membrane concentration is used in water treatment, waste treatment, demineralized sea water and desalination. In dairy industry, lactose is produced by this method. Membranes can be applied to fruit juice process for clarification, concentration and

acidification (Heldman and Lund, 2007). Other applications of membrane system are concentrated egg white (Conrad et al., 1993; Elaya and Gunasekaran, 2002), coffee and tea extracts, and concentrated flavor compounds (Heldman and Lund, 2007).

#### **1.4. Freeze Concentration**

The other commercial method is the freeze concentration and this study is related with this method. Freeze concentration has several advantages. It does not use up much energy (Cassano et al., 2007). The latent heat of phase transition from water to steam is 540 kcal/kg while the latent heat of phase transition from water to ice is 76 kcal/kg so energy is saved by freezing concentration (Yuping and Baomin, 2001; Canovas et al., 2005). Freeze system is worked at low temperatures so corrosion is not significant, thus it requires less maintenance. The most important advantage of this method is that, the characteristics of the original product do not change due to lower temperatures. It also reduces microbiological activity (Lee and Lee, 1999). Freeze concentration is carried out in closed systems so oxidation and denaturation are prevented. This technique is selected to produce a high quality concentrated liquid food and it is suitable for new product development. In fact, it is extremely reliable; it can be both batch and continuous. The process can be used for different products from small volume to large volume. A freeze concentration pilot plant apparatus was shown in Figure 1.1. For freeze concentration, water removal capacity is between 5 and 30,000 kg/hour in industry. Various configurations with capacities ranging from 100 to 400,000 tons per year have been installed (<http://www.gea-messo-pt.com>). The freeze concentration is not to become widespread due to its the high price of the equipment (Shamsudin, 2000). The degree of concentration is limited like reverse osmosis (Cassano et al., 2007) and it is not used for cheap juices (Ramaswamy and Marcotte, 2006a). Sensory quality is nearly the same for both freeze concentration and reverse osmosis (Sanchez et al., 2009).

##### **1.4.1. History of Freeze Concentration**

Anton Maria Lorgna was issued the first article about freezing process in Italy in 1786 (Nebbia and Menozzi, 1968). He used this method to desalinate sea water. His work was important to develop some refrigerating devices. The difficulties in establishing a simple equipment to produce drinking water was pointed out in 1940s (Parker, 1942). The development of freeze concentration put forward commercial

scale. It was used to purify p-xylene in the chemical and petroleum industry in the 1950s. Then, this system was applied to different industrial areas like wastewater treatment, chemical industry, pulp and paper industry, and food industry etc. Generally, this technique has been the most successful in food industry. Warezak (1969) developed the clathrate process in which one component molecules were trapped within the crystals structure of another physically. In this way, freeze concentration was used in beet industry in 1951 and it was used to concentrate vinegar solution in 1959 in a commercial scale (Englezos, 1992).



Figure 1.1. Freeze Concentration Pilot Plant

#### **1.4.2. Some Application of Freeze Concentration in the Food Industry**

Generally freeze concentration is used in food and chemical industry. It has been applied to various food samples because it is independent of the food composition (Lemmer et al., 2000).

Some examples can be given why this method is used in food industry. Wine and beer production is aimed to prevent loss of alcohol and flavors by freezing which occurs before fermentation because sugar, not alcohol, is concentrated. Concentration of coffee and tea before freeze drying is necessary for flavor retention.

Preconcentration is essential before freeze drying so freeze concentration is selected to flavor containing liquid foods (Karel, 1975). Especially heat-sensitive fruit juices which are not suitable for the application conventional concentration processes can be deformed physically by extensive heat. This system is therefore used to prevent loss of aroma and prevent any thermal damage (Jusoh et al., 2009). Freeze concentration is also used in dairy industry for new product development, high quality ingredients, and improved final products etc. (Karel, 1975). According to Lemmer et al. (2000), the concentration of wastewater is preferred due to economic reasons. Preconcentration of wastewater is carried out by two methods which are evaporative and freeze concentration. Wastewater contains organic components and salts. High organic concentration and pH values are not suitable for membranes. By freeze concentration system, pure water stream is improved from the wastewater and this stream is used to prepare demineralized water again. 75% of the water in the wastewater can be recovered by this method. This technique has been the best preserved method of freshness of food product until now. It is an improvable method to become commercial (Lemmer et al., 2000). Freeze concentration presents new opportunities in a wide variety of applications.

#### **1.4.3. Freeze Concentration Process**

The freeze concentration is a sophisticated method because it fits very well into modern food processing plant. Freeze concentration can be carried out by several ways due to equipment and refrigerant alternatives; so a number of conduits, pumps and controls can be added to the system. Therefore, several freeze-concentration process systems can be imagined. For a single stage process is composed of minimum two components which are a crystallizer or a freezer and a separator (wash column). A multi-stage process is a more appropriate system for industry and its production rate is higher as well as consuming lower energy (Rane and Jabade, 2002).

The basic principle of freeze concentration is solid-liquid phase equilibrium. The solution has the solvent as water and soluble components. The freeze concentration separates water from soluble solid without affecting the other components at low temperatures. This condition is important for sensory properties of the original food because lower temperatures protect to preserve quality of the original food (Jusoh et

al., 2009). The freezing temperature of solution is reduced by the presence of solute so it freezes below pure water freezing temperature (Yuping and Baomin, 2001; Canovas et al., 2005). Freeze concentration removes heat from aqueous solution and one component crystallizes. It is obvious that freeze concentration removes water as ice crystals; thus water content is lowered in solution and remained part gets highly concentrated solution (Jusoh et al., 2009). The basic components of a freeze concentration unit are crystallizer and wash column.

#### **1.4.3.1. Function of Crystallizer**

The crystallizer is a container with a cooling jacket. The inner wall of crystallizer is scraped by rotating blades. The outer wall of crystallizer is cooled by a circulating refrigerant which produces small ice crystals, called nuclei. These ice crystals are removed heat from the solution. It is achieved by a scraped surface heat exchanger (SSHE) at high supercooling and low residence times. Then, these ice crystals pass the recrystallizer for growing large spherical ice crystals (Rane and Jabade, 2002; <http://www.gea-messo-pt.com>). The system is required to get large and symmetrical ice crystals because large crystals are separated easily. Large ice crystals are required to minimize removing a portion of solid within water, also (Wenzelbergel, 1951). Paddle crystallizer is accomplished rather softly stirring thick slurry of ice crystals and allowed growing the bigger crystals from smaller ones (Muller, 1967). The each crystal separation shows different degree of clumping of the crystals (Kessler, 1986). Definitely, the function of crystallizer produces ice crystals and grows these crystals.

#### **1.4.3.2. Function of Wash Column**

The function of wash column is separation of the concentrated liquid and ice crystals efficiently by unique device. The separation can be achieved by centrifugation, vacuum filtration, filter pressing as well as wash column. According to Mellor's article (1978), the result of the efficiencies of separation technique was noted as 50% for centrifuging, 71% for vacuum filtration, 89–95% for filter pressing and 99.5% for wash columns. Wash column is a vertical enclosed cylinder whose bottom is operated by feeding ice concentrate. Ice crystals are removed by gravity, hydraulic pressure, or mechanically such as piston and screw. For removing all traces of concentrated liquid, a compressed ice crystal bed is washed with melted ice.

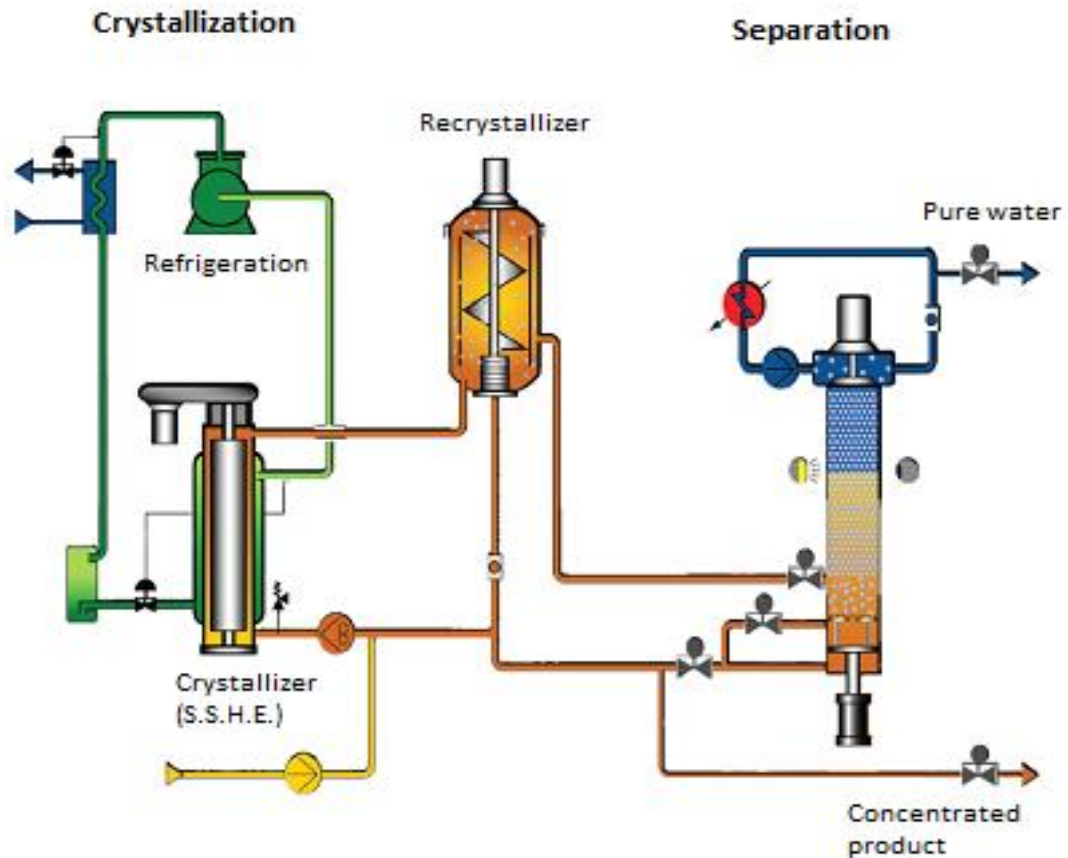


Figure 1.2. Freeze concentration system for concentrated juice production (adapted from <http://www.gea-messo-pt.com>).

A concrete example can be given for a freeze concentrator which has a scraped-wall heat exchanger attached to an adiabatic recrystallizer tank as shown in Figure 1.2. The liquid sample is pumped from feed tank into the heat exchanger. Basically, the solution is cooled to a temperature which is nearly freezing point by a feed heat exchanger. The juice flows over the refrigerated surface. Small ice crystals are formed due to high supercooling and short residence times. This crystal slurry is passed to the recrystallizer in where it is blended with larger crystals (Lemmer et al., 2000). The recrystallizer tank is used to grow these small ice crystals which form desired pure ice size. Larger crystals have a higher equilibrium temperature than smaller crystals and the big ice crystals affect smaller ones which melt and recrystallize on the surface of larger crystals (Gibbs-Thompson effect). Then, ice and concentrated juice are separated at wash column. The concentrated juice is discharged through a filter plate and ice crystals are apart from the concentrate. The wash column is rinse due to remove adhering concentrate from the ice crystals (Myerson and Ginde, 1993).

#### **1.4.4. Freezing Techniques**

##### **1.4.4.1. Suspension Freeze Concentration**

The freezing technique or cryoconcentration technology is based on suspension crystallization method. Generally, the suspension crystallization is known as first system of freezing technique (Huige and Thijssen, 1972; Hartel and Espinel, 1993). This system is also known as crystallization process which forms in two phases. Ice nuclei are formed in initial phase and the growth of ice nuclei in the solution occurs in second phase. As a result; suspended ice crystals are formed in the liquid sample. The ice growth rate of suspension freeze concentration is different from other methods, the growth rate is lower than the others but this provides higher ice purity than the others. However, a limitation presents the size of ice crystals (Aider and Halleux, 2008) and the concentration degree is limited. In addition, it is complicated system which contains surface-scraper heat exchanger, re-crystallization, washing tower for producing seed ice, growth ice crystals and separation of ice crystals, respectively (Miyawaki et al., 2005). The scraped surface heat exchanger needs to produce ice seeds to provide ice growth. The size of ice crystals are enlarged in a ripening vessel. Moreover, a filtration system is used to separate ice crystals from concentrated liquid (Widehem and Cochet, 2003).

Suspension crystallization is difficult for separation of ice crystals from the concentrated solution. Therefore, Ostwald ripening mechanism is used to grow large ice crystals (Huige and Thijssen, 1972). Ostwald ripening is defined as water vapor tends to transfer from higher vapor pressure area to lower vapor pressure area at a constant temperature. Higher vapor pressure is in the surface of small crystals and lower vapor pressure is in the surface of larger crystals. However, the most important problem is caused by temperature fluctuation. When the temperature of frozen storage is increased, small ice crystals melt, so the amount of water rises. If the temperature is decreased, the form of nucleus does not continue, so water refreezes on the surface areas of large crystals. Therefore, the number of ice crystals decreases and the crystal size increase.

Ice nucleus begins to grow in a solution, ice phase is formed and solutes are not accepted in this phase. The interfacial tension between ice and mother liquor is low, concentrate adheres to the crystal surface. As a result, a modification is observed in



solid-liquid equilibrium temperature. Equilibrium temperature increases decreasing solute concentration, thus, supercooling increases in front of the interphase which can cause instability for growth of ice crystals. Ice nucleation agents are recommended to avoid impurities during initial supercooling in the ice phase. Ice nucleation agents are known as insoluble materials, biogenic and non biogenic origin. Biogenic ice nucleation agent reduces the supercooling degree by increasing ice nucleation temperature. Non biogenic is not preferred for application of food products (Petzold and Aguilera, 2009). The suspension freeze concentration method has been used concentrated coffee extract, fruits juice like apple, berry and kiwi juices etc. (Deshpande et al., 1982; Braddock and Marcy, 1985), milk (Hartel and Espinel, 1993) and wastewater treatment (Heist, 1979).

#### **1.4.4.2. Progressive Freeze Concentration**

The second system is known as layer crystallization. Crystallization of water in the solution forms as an ice layer on a cold surface (Muller and Sekoulov, 1992; Flesland, 1995). This technique is called progressive freeze concentration. It is easy separation due to not necessary other unit operations. Thus, progressive method is very different technique in the crystallization process especially from the suspension crystallization method (Miyawaki et al., 2003). Because progressive method produces a single ice crystal while the suspension method produces many small ice crystals. The method is based on cooling surface in a crystallization vessel. Ice and mother solution is to be separated easily with this way (Miyawaki et al., 2003), but impure ice crystals can be produced.

For progressive freeze concentration, Wakisaka et al. (2001) experienced nearly 1.4 fold concentrated glucose solution to represent wastewater. As well as this, average 135 kg ice was produced in 70 minutes by batch operation. In an article by Miyawaki et al. (2005) used sucrose solution, coffee extract and tomato juice for progressive freeze concentration and their results gave high yield and productivity on account of proof tubular ice system suited to progressive freeze concentration. Raventos et al. (2006) studied sugar solutions such as glucose, fructose and sucrose in progressive freeze concentration which produced 31.2° Brix sucrose from 5° Brix sucrose in 16.6 hours in a successful conclusion. This method has been used to purify organic materials (Atwood, 1973; Gilbert, 1991), aqueous and organic solutions (Matthews

and Coggeshall, 1959; Shapiro, 1961), concentrated impurities in organic solvent (Matthews and Coggeshall, 1959), waste water treatment and low thermal energy regeneration (Miyawaki et al., 2009). However; productivity of progressive freeze concentration is lower than suspension freeze concentration (Jusoh et al., 2009). New designs have been aimed high efficiency and productivity. Suspension and progressive freeze concentration are shown in Figure 1.3.

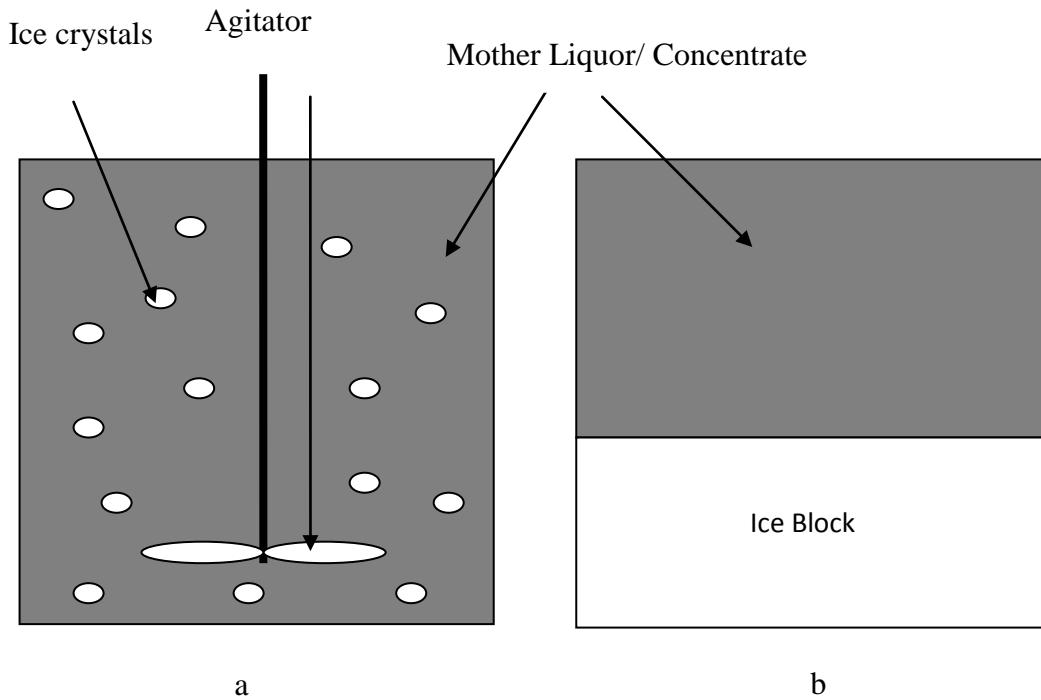


Figure 1.3. (a) Suspension Freeze Concentration and (b) Progressive Freeze Concentration (Miyawaki et al., 2003; Sanchez et al., 2009).

Concentration is purposed of reduction of volume and this phenomenon can be described as effective partition constant value,  $K$ , calculated from the concentration and volume of liquid and solid phases (Gu et al., 2008). Effective partition constant is related to initial concentration. This constant is increased by initial concentration. The higher initial concentration, the higher  $K$  value but efficiency of the system is lower (Miyawaki et al., 1992). According to Miyawaki (1998), the effective partition coefficient of solute between ice and liquid phases at the ice–liquid interface is the important factor of this process efficiency at progressive cryoconcentration technology. It is defined as:

$$K = C_S / C_L \quad (1)$$

Where  $C_S$  is solute concentration in the ice while  $C_L$  is the solute concentration in the

solution phase. The K value is calculated experimentally. Equation (1) can be obtained from a mass balance, and is defined as:

$$C_L V_L = -C_S dV_L + (C_L + dC_L)(V_L + dV_L) \quad (2)$$

where  $V_L$  is the liquid volume and  $C_L$  is the liquid concentration.  $(-dV_L)$  is defined as a small volume increase in the ice phase and  $(dC_L)$  is the solute concentration increase in the solution phase. It is assumed that complete mixing in the solution phase and no mixing in the ice phase. Combining equation 1 and 2:

$$(dC_L / C_L) / (dV_L / V_L) = K - 1 \quad (3)$$

If K is assumed constant equation (3) is rewritten as:

$$(1 - K) \log (V_L / V_0) = \log (C_0 / C_L) \quad (4)$$

Where  $V_0$  is the liquid volume at time zero and  $V_L$  is the liquid volume at an arbitrary time and  $C_0$  and  $C_L$  are the concentration of solute in the solution at the corresponding time. K value is between 0 and 1 and this means that when  $K=0$ , freeze concentration does not occur and when  $K=1$  freeze concentration occurs completely. K is calculated from this equation and a straight line is observed from graph of  $\log (C_0 / C_L)$  versus  $\log (V_L / V_0)$  (Miyawaki et al., 2005). The effective partition coefficient is the noticeable coefficient of solute between ice phase and bulk liquid phase experimentally. The concentration polarization model analyzes the effective partition coefficient on the solute concentration distribution near the ice–liquid interface, and the advance rate of the ice front ( $u$ ) and the mass transfer coefficient ( $k$ ) at the ice–liquid interface are stated as functions. The partition coefficient is defined at following equation (Miyawaki et al., 1998):

$$K = K_0 / [K_0 + (1 - K_0) \exp (-u/k)] \quad (5)$$

Where  $K_0$  is defined as the limiting partition coefficient representing the partition coefficient of solute at the ice–liquid interface.  $K_0$  is response the partition coefficient at an infinitesimal advance rate of ice front ( $u$  tends to 0) and at infinite mass transfer coefficient ( $k$  tends to  $\infty$ ) because K is equal to  $K_0$  in Eq. (5).

A mass balance equation compared with theory is used:

$$W_{pred} = (C_{FS} - C_I) / (C_{FS} - C_H) \quad (6)$$

Where  $W_{\text{pred}}$  is defined as predicted ice mass ratio (kg ice/ kg solution),  $C_I$  is the initial solutes concentration (°Brix),  $C_{FS}$  is the final solutes concentration (°Brix) and  $C_H$  is the solutes concentration of ice (°Brix) (Pradistsuwana et al., 2003).

The efficiency of each concentration stage is calculated by following equation:

$$\text{Efficiency (\%)} = [1 - (C_H / C_{FS})] * 100 \quad (7)$$

Where  $C_{FS}$  is the concentration solutes in the liquid phase (°Brix) and  $C_H$  is the concentration of solutes (°Brix) in the ice formed at the end of each concentration stage.

The impurity of ice is defined as soluble solid remaining in the ice. The relative impurity of ice is determined as the ratio of ice concentration and the concentration in the concentrated juice at the end (Sanchez et al., 2010).

$$\text{Impurity ratio (\%)} = (C_F / C_{FS}) * 100 \quad (8)$$

For freeze concentration requires large and spherical ice crystals because large ice crystals provides very easy separation and requires short time. So, optimal operation parameters should be selected. Refrigerant temperature, crystal growth rate and agitation rate should be controlled due to heat and mass rates. Moreover, the average distribution coefficient (K) was defined as:

$$K = W_{\text{ice}} / W_S \quad (9)$$

Where  $W_{\text{ice}}$  and  $W_S$  are the solute mass fraction in ice (wt %) the solute, the solute mass fraction in the bulk solution (wt %), respectively (Sanchez et al., 2011).

#### **1.4.4.3. Block Freeze Concentration**

The third procedure is defined as block crystallization. A liquid solution is frozen completely and then the temperature at the center of the product is decreased below the freezing point. After that, this frozen solution is thawed and the concentrated part is separated from ice. If the thawing temperature is controlled, it is possible to achieve process efficiency higher than 90%, so solute in the ice crystal is decreased successfully (Aider and Halleux, 2008). The phenomenon of these methods produced ice by different heat extraction. The advantage of block concentration is that it is easier than the others. That is, the equipment usage is easy due to no moving parts

(Sanchez et al., 2009). Complete block technology is influential procedure for concentrated liquid foods which have high nutritive value and organoleptic properties (Aider et al., 2007; Aider and Halleux, 2008).

Partial block freeze technology is new and effective technique. A liquid solution is firstly introduced in a crystallizer chamber and then passed a pipe, in which a cryogen is circulated, is occurred partially frozen from the center.

A block freezer is based on separation of solution at low temperatures. Refrigeration machine provides a block of ice. The block of ice is to be separated by gravitation. Further perfection of the block freezers can be carried out at the expense of intensification of the processes of heat transfer and mass transfer in the boundary layer (Burdo et al., 2007). This technology is succeed in aromatic compounds from grape seeds, fruit juices, coffee extract and concentrate milk whey (Aider and Halleux, 2008).

#### **1.4.4.4. Eutectic Freeze Concentration**

Different method is used to separate inorganic salts from aqueous solution by crystallization of both solute and ice at the same time. This technique is called as eutectic freeze concentration (Vaessen et al., 2002). The solution is cooled down below the eutectic temperature which can be defined as the melting point dependent on ratio of ingredients in solution for frozen heterogeneous liquid food. The ratio of mixture affects melting point which is as far as low. Moreover, the crystal size distribution is important because it affects performance of the downstream separation equipment. An example for eutectic cryoconcentration, it can be applied to the purification of waste water streams with high soluble salts. Aqueous electrolyte solutions can be distinguished into pure water as ice and salt. Gravitation achieves separation between salt and ice crystals which have different densities (Vaessen et al., 2002; Vaessen et al., 2003). Eutectic cryoconcentration technology has been investigated by Stepakoff et al. 1974. The system requires equipment which is composed of solid/ solid separator, filters, wash column, pumps etc. This technique has been recently modernized so it requires low energy consumption, and provides high purity of products (Van der Ham et al., 1998).

### **1.5. Aim of the Present Study**

The freeze concentration method is important because of maintaining original properties of foods. Among different freeze concentration methods, block freeze concentration is adopted in this study due its simplicity and applicability under laboratory conditions. In addition, the degree of separation was analyzed by a differential melting process which is time dependent. This method is unique, because no similar methods have not been observed in the literature.

In this study, separation of high-concentrated soluble solids from ice was accomplished for varied initial concentrations of various fruit juices at different temperatures. The effect of the frozen samples size on the degree of separation of the soluble solids from the ice was also studied at different time intervals.

## **CHAPTER 2**

### **MATERIALS AND METHODS**

#### **2.1. Materials**

##### **2.1.1. Sumac Juice**

Sumac berries were supplied from sumac trees in the campus of University of Gaziantep. After harvesting sumac berries in clusters, they were destemmed. Then, these berries were screened to separate unripe flower residues and impurities. They were soaked and leached in distilled water. The suspended insoluble materials in the sumac juice were removed by filtration.

##### **2.1.2. Apple Juice**

Apple juice was purchased from a local market in Gaziantep. 100% apple juice, not containing added sugar, was selected.

##### **2.1.3. Grape Juice**

Grape juice was purchased from a local market in Gaziantep. 100% grape juice, not containing added sugar, was selected.

##### **2.1.4. Carrot Juice**

Carrots were purchased from a local market in Gaziantep. Carrots were washed to remove impurities. Carrot juice was obtained by using hard fruit squeezer.

##### **2.1.5. Water**

Distilled water was used for adjusting concentration of juices. It was supplied from the Food Engineering Department, General Chemistry Laboratory, water distilling unit.

#### **2.2. Preparation of Concentrated Juices**

These samples were concentrated using two types evaporators: a rotary evaporator (Bibby, RE 100, Rotary Evaporator) and a laboratory type rising film evaporator

(Armfield FT 22-A, Rising Film Evaporator).

### **2.3. Variation of pH Value of Concentrated Juices with Concentration**

The pH value of various fruit juices at different concentration was measured with a digital pH meter (WTW 2A10-1012, Germany). The measurement was done at room temperature  $23 \pm 2$  °C.

### **2.4. Titratable Acidity of Juices**

Titratable acidity was measured by titration with 0.1 N NaOH. The sample was titrated with 0.1 N NaOH using a few drops of 1% phenolphthalein solution as indicator until a pink color was observed.

### **2.5. Measurement of Concentration of Juices**

The concentration of solutes was measured in terms of °Brix using Pocket PAL-1 and Pocket PAL-2, ATAGO hand refractometer. The accuracy of these refractometers was  $\pm 0.1$  with a measurement range of 0-53 °Brix for Pocket PAL-1 and 45-93 °Brix for Pocket PAL-2. Results were expressed as Brix degrees. The measurements were made at the different temperatures (10, 15, 20 and 25°C).

### **2.6. Preparation of Juices with Varied Concentration**

Various concentration of juices were prepared. Samples with various soluble solid contents (1, 2, 3, 4, 5, 10, 15, 20, 25, 30, 35 and 40 °Brix) were prepared by dilution of the original concentrated juice. They were prepared for each sample.

### **2.7. Analysis of Separation of Juice from Ice**

100 mL of each sample was placed in 150 mL bottles. These samples were frozen in a deep freezer of a refrigerator at -18 °C. Then, they were thawed at different temperatures (10, 15, 20 and 25 °C) in a temperature-controlled oven (Binder, Germany). This experiment was shown in Figure 2.1. The bottle was put in this oven. Frozen juice began to melt. Then, melting portion was collected in a small beaker. The concentration of melting portion was measured at 5-minute time intervals during melting. All experimental measurements were made in double with samples prepared from the same concentrated juice. The same procedure was used for 250, 500 and 750 mL of samples at 10°C.



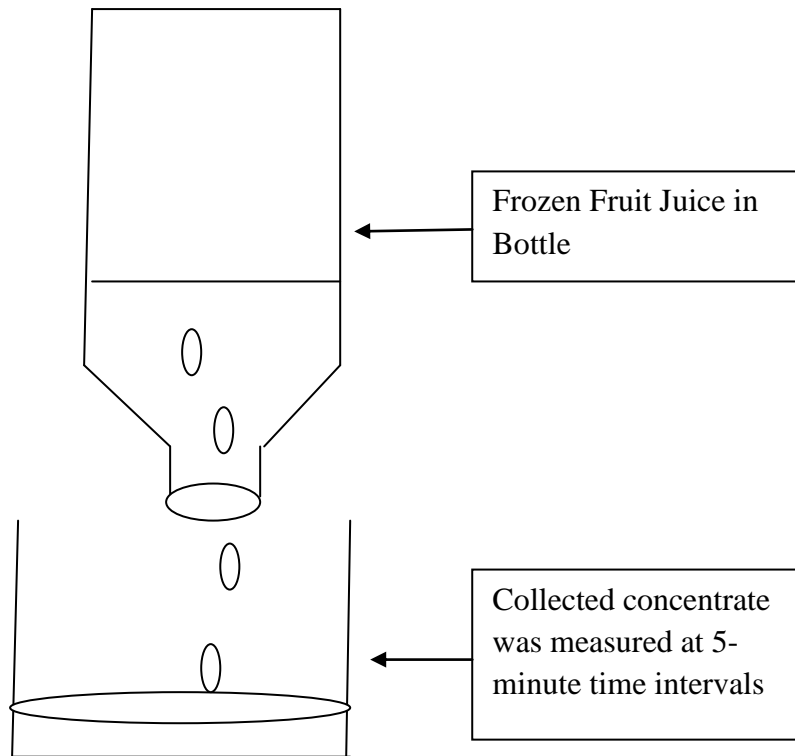


Figure 2.2. The experimental set-up of the system

## 2.8. Statistical Analysis

Exponential decay model was observed to be the best fitted model for all data. The parameters of the models were calculated by the NLIN (non-linear) and linear procedures of the Sigma Plot (Scientific Graph System, version 11.00).

## CHAPTER 3

### RESULTS AND DISCUSSION

#### 3.1. The Effect of Initial Concentration on Degree of Separation

Different samples at constant initial concentrations were thawed as a function of different temperatures. During the experiment, separation of the concentrated juice from ice was observed. The degree of separation could be dependent upon initial concentrations of the juice. This behavior is clearly shown in Tables 3.1– 3.4. Thus, the degree of separation decreased with increasing initial concentration of the juice. In other words, an inverse relationship between the initial concentration and the degree of separation was found. The concentration of the first drop of juice (having an initial concentration of 1 °Brix) was observed to be 26.1 °Brix. The concentration of the first drop of juice (having an initial concentration of 20 °Brix) was observed to be 42.9 °Brix. The concentration of the first drop of juice (having an initial concentration of 40 °Brix) was increased to 47.6 °Brix.

Table 3.1. Concentration of the first drop of sumac juice at different initial concentrations at 10 °C

<b>Initial Concentration (°Brix)</b>	<b>Concentration of the First Drop (°Brix)</b>	<b>Degree of Separation (Concentration of the First Drop/ Initial Concentration)</b>
<b>1</b>	26.1	26.10
<b>2</b>	28.2	14.10
<b>3</b>	32.0	10.67
<b>4</b>	33.1	8.28
<b>5</b>	34.4	6.88
<b>10</b>	39.1	3.91
<b>15</b>	42.5	2.83
<b>20</b>	42.9	2.15
<b>25</b>	43.7	1.75
<b>30</b>	44.8	1.49
<b>35</b>	46.9	1.34
<b>40</b>	47.6	1.19

Table 3.2. Concentration of the first drop of apple juice at different initial concentrations at 10 °C

<b>Initial Concentration (°Brix)</b>	<b>Concentration of the First Drop (°Brix)</b>	<b>Degree of Separation (Concentration of the First Drop/ Initial Concentration)</b>
1	24.3	24.30
2	28.4	14.20
3	30.2	10.07
4	30.5	7.63
5	30.7	6.14
10	40.8	4.08
15	42.6	2.84
20	44.7	2.24
25	45.8	1.83
30	47.5	1.58
35	47.7	1.36
40	48.5	1.21

Table 3.3. Concentration of the first drop of grape juice at different initial concentrations at 10 °C

<b>Initial Concentration (°Brix)</b>	<b>Concentration of the First Drop (°Brix)</b>	<b>Degree of Separation (Concentration of the First Drop/ Initial Concentration)</b>
1	26.6	26.60
2	27.3	13.65
3	29.7	9.90
4	31.1	7.78
5	31.9	6.38
10	39.7	3.97
15	41.1	2.74
20	44.9	2.25
25	45.6	1.82
30	46.5	1.55
35	47.8	1.37
40	48.8	1.22

Table 3.4. Concentration of the first drop of carrot juice at different initial concentrations at 10 °C

<b>Initial Concentration (°Brix)</b>	<b>The Concentration of the First Drop (°Brix)</b>	<b>Degree of Separation (Concentration of the First Drop/ Initial Concentration)</b>
1	23.2	23.20
2	24.5	12.25
3	25.4	8.47
4	26.2	6.55
5	28.7	5.74
10	30.7	3.07
15	33.4	2.23
20	35.5	1.78
25	37.1	1.48
30	40.2	1.34
35	41.1	1.17
40	42.4	1.06

### 3.2. Effect of Concentration on the pH Value of Different Juices

pH was measured at room temperature for different fruit juices. Table 3.5 shows

Table 3.5. Variation of pH with concentration for different juices

<b>Concentration (°Brix)</b>	<b>pH (Sumac Juice)</b>	<b>pH (Apple Juice)</b>	<b>pH (Grape Juice)</b>	<b>pH (Carrot Juice)</b>
1	2.94	3.78	3.75	6.64
2	2.93	3.76	3.74	6.61
3	2.91	3.74	3.73	6.57
4	2.86	3.73	3.72	6.53
5	2.80	3.68	3.71	6.46
10	2.76	3.66	3.70	5.99
15	2.69	3.64	3.67	5.86
20	2.61	3.63	3.66	5.63
25	2.59	3.63	3.65	5.57
30	2.55	3.58	3.64	5.47
35	2.49	3.56	3.64	5.43
40	2.38	3.55	3.63	5.35

variation in pH values by changing concentration for each sample. According to these results, pH values of the concentrates did not change with concentration significantly. Values of pH slightly changed for buffer solution. Therefore, these concentrates were observed to behave like buffers.

### 3.3. Titratable Acidity of Different Samples

Titrateable acidity is a measure of the total amount of acid present in the solution. The titrateable acidity of a solution occurs for any acid that loses protons in an acid- base reaction.

Table 3.6. Titratable acidity of different fruit juices

<b>Sample</b>	<b>°Brix</b>	<b>Titrateable Acidity (g/100 mL)</b>	<b>Corrected Titrateable Acidity (g/100 mL)</b>
<b>Sumac</b>	1.5	1.69	12.28
<b>Apple</b>	10.9	2.79	2.79
<b>Grape</b>	10.6	1.23	1.26
<b>Carrot</b>	3.5	0.32	0.99

### 3.4. Effect of Temperature on the Concentration of Sumac Concentrates

In this study, four temperatures were studied 10, 15, 20 and 25 °C. At mentioned temperatures concentrations were measured at five minute time intervals. The data of concentration versus time were plotted at each temperature.

As expected, the juice that separates from the frozen block is more concentrated at the beginning of the thawing step and decreases as thawing proceeds as in Figures 3.1– 3.12. These figures show that the degree of separation was observed to be dependent upon initial concentrations of the juice. The degree of separation decreased with increasing initial concentration of the juice. Tables 7– 18 indicate that equation coefficient decreases with increasing temperature. Temperature effect on the separation of concentrated juice is seen clearly. This is shown that lower temperatures exhibit higher degree of separation.

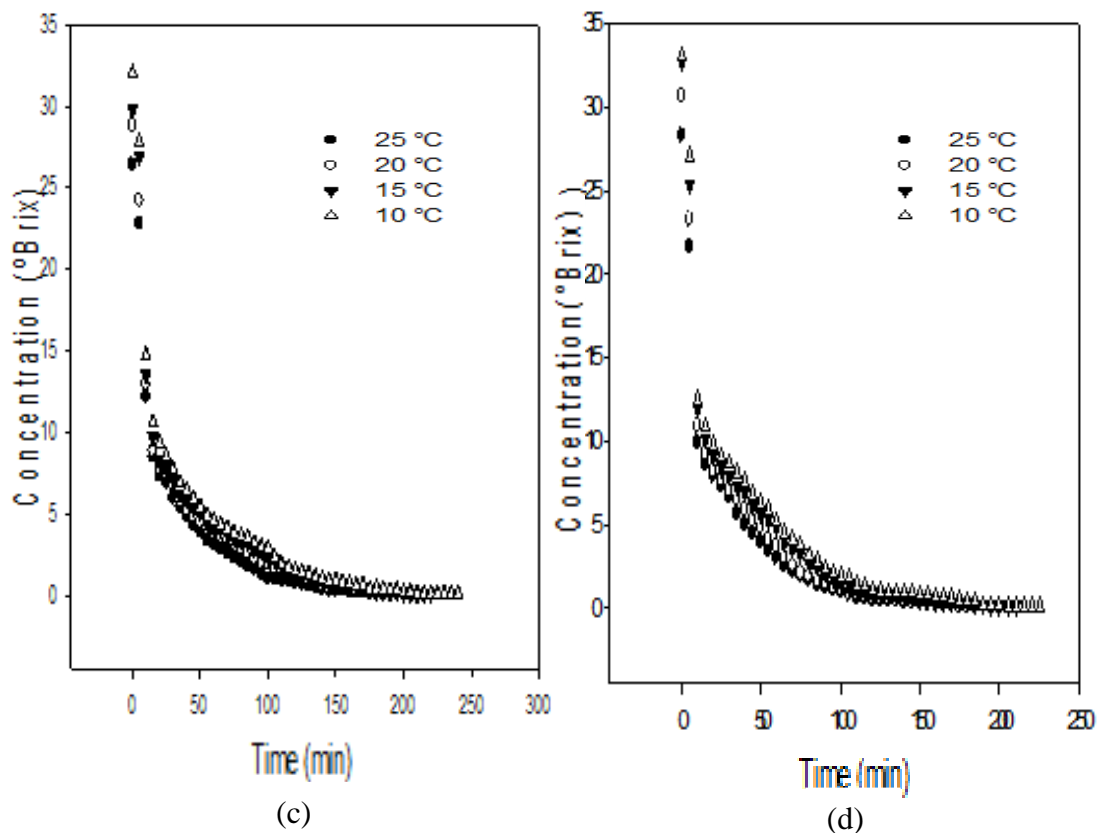
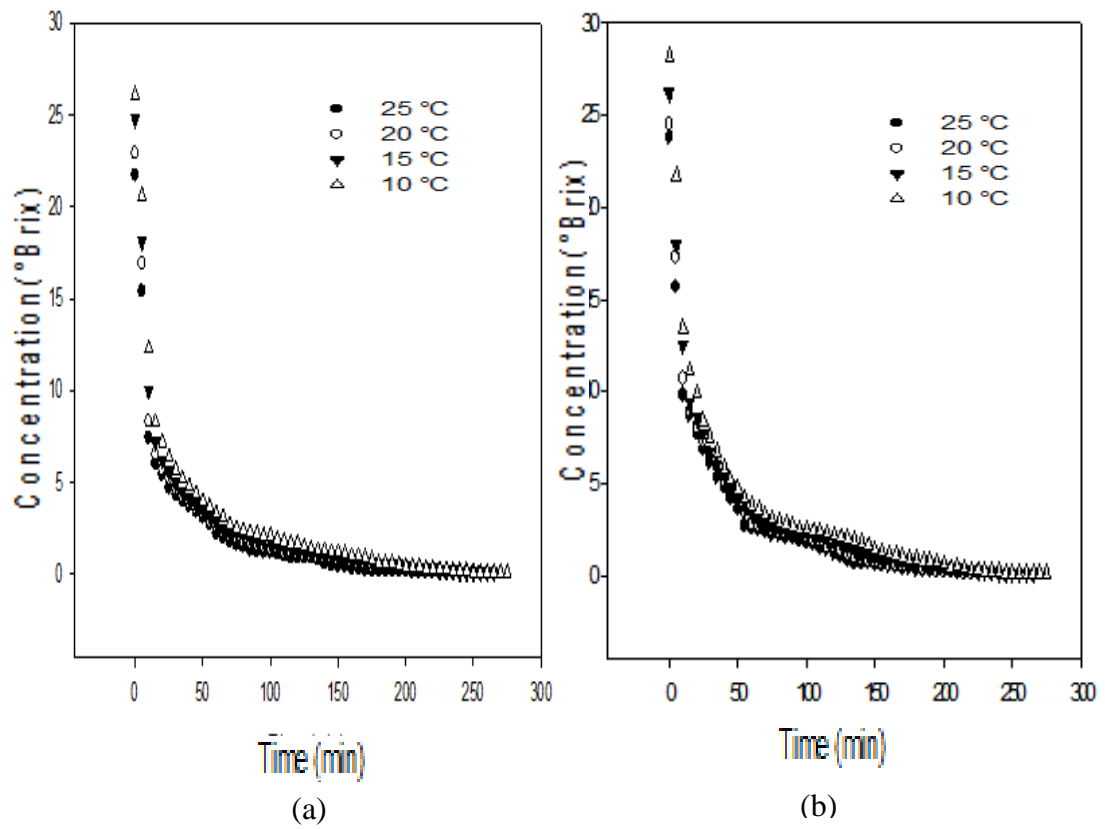


Figure 3.1. Variation of concentration with time for sumac concentrates at different temperatures: (a) initial concentration 1 °Brix, (b) initial concentration 2 °Brix, (c) initial concentration 3 °Brix, (d) initial concentration 4 °Brix

Table 3.7. Regression analyses for sumac concentrates (initial concentration 1, 2, 3 and 4 °Brix) at different temperatures

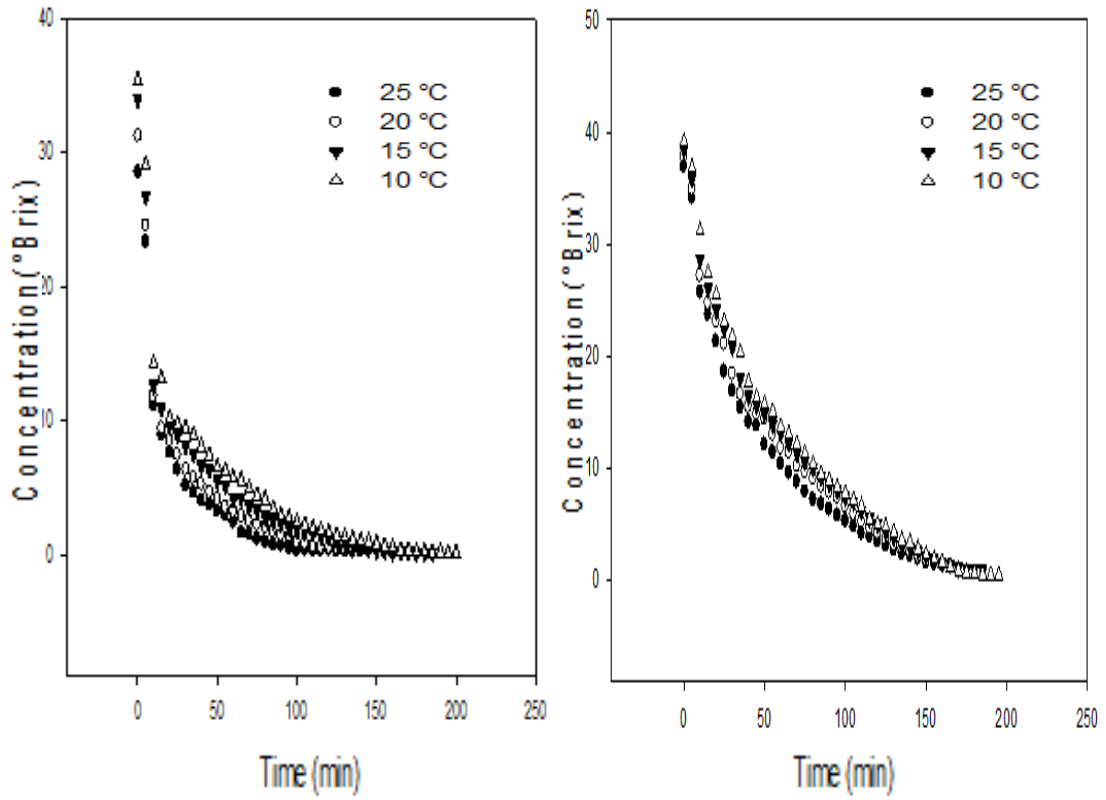
<b>Initial Concentration (°Brix)</b>	<b>Temperature (°C)</b>	<b>Equation</b>	<b>R<sup>2</sup></b>
<b>1</b>	<b>10</b>	y= 23.39*exp(-0.044*x)	0.916
	<b>15</b>	y= 22.12*exp(-0.050*x)	0.913
	<b>20</b>	y= 20.38*exp(-0.051*x)	0.908
	<b>25</b>	y= 19.23*exp(-0.053*x)	0.912
<b>2</b>	<b>10</b>	y= 24.012*exp(-0.035*x)	0.926
	<b>15</b>	y= 21.625*exp(-0.036*x)	0.926
	<b>20</b>	y= 20.29*exp(-0.036*x)	0.930
	<b>25</b>	y= 19.63*exp(-0.038*x)	0.929
<b>3</b>	<b>10</b>	y= 28.68*exp(-0.040*x)	0.912
	<b>15</b>	y= 27.25*exp(-0.042*x)	0.904
	<b>20</b>	y= 26.22*exp(-0.045*x)	0.918
	<b>25</b>	y= 24.89*exp(-0.047*x)	0.940
<b>4</b>	<b>10</b>	y= 27.98*exp(-0.037*x)	0.907
	<b>15</b>	y= 27.94*exp(-0.041*x)	0.909
	<b>20</b>	y= 27.05*exp(-0.047*x)	0.923
	<b>25</b>	y= 25.32*exp(-0.049*x)	0.929

\* x= Time (min)

\* y= Concentration (°Brix)

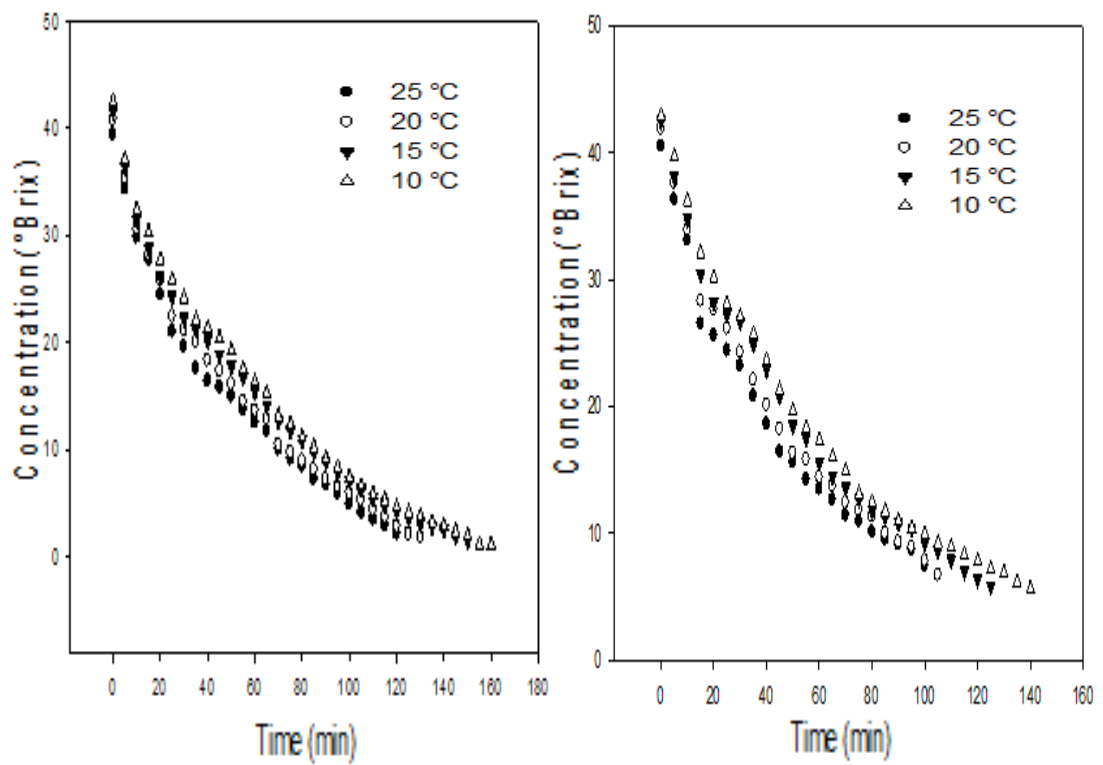
Table 3.7 shows the data fitted by non-linear regression analysis at different temperatures for lower sumac concentrates. The determination of coefficient (R<sup>2</sup>) values changed between 0.904 and 0.940. The higher R<sup>2</sup> values were obtained by using exponential decay model. This model was observed to be the best fitted model for all data. In the model equation, y-axis shows that the concentration (°Brix) decays with x-axis (time). Negative exponent indicates exponential decay.

Figure 3.2 illustrates the change in °Brix as a function of time at different temperatures for initial concentration between 5 and 20 °Brix of sumac concentrates.



(a)

(b)



(c)

(d)

Figure 3.2. Variation of concentration with time for sumac concentrates at different temperatures: (a) initial concentration 5 °Brix, (b) initial concentration 10 °Brix, (c) initial concentration 15 °Brix, (d) initial concentration 20 °Brix



Table 3.8. Regression analyses for sumac concentrates (initial concentration 5, 10, 15 and 20 °Brix) at different temperatures

<b>Initial Concentration (°Brix)</b>	<b>Temperature (°C)</b>	<b>Equation</b>	<b>R<sup>2</sup></b>
<b>5</b>	<b>10</b>	$y = 29.86 \cdot \exp(-0.0350 \cdot x)$	0.906
	<b>15</b>	$y = 29.46 \cdot \exp(-0.0424 \cdot x)$	0.912
	<b>20</b>	$y = 28.88 \cdot \exp(-0.0530 \cdot x)$	0.937
	<b>25</b>	$y = 27.66 \cdot \exp(-0.0592 \cdot x)$	0.958
<b>10</b>	<b>10</b>	$y = 37.73 \cdot \exp(-0.017 \cdot x)$	0.992
	<b>15</b>	$y = 36.07 \cdot \exp(-0.017 \cdot x)$	0.989
	<b>20</b>	$y = 35.04 \cdot \exp(-0.018 \cdot x)$	0.985
	<b>25</b>	$y = 34.27 \cdot \exp(-0.021 \cdot x)$	0.984
<b>15</b>	<b>10</b>	$y = 40.66 \cdot \exp(-0.017 \cdot x)$	0.990
	<b>15</b>	$y = 39.22 \cdot \exp(-0.017 \cdot x)$	0.989
	<b>20</b>	$y = 38.75 \cdot \exp(-0.019 \cdot x)$	0.992
	<b>25</b>	$y = 37.56 \cdot \exp(-0.020 \cdot x)$	0.991
<b>20</b>	<b>10</b>	$y = 41.77 \cdot \exp(-0.015 \cdot x)$	0.997
	<b>15</b>	$y = 40.78 \cdot \exp(-0.015 \cdot x)$	0.994
	<b>20</b>	$y = 40.02 \cdot \exp(-0.017 \cdot x)$	0.992
	<b>25</b>	$y = 38.51 \cdot \exp(-0.016 \cdot x)$	0.986

\* x= Time (min)

\* y= Concentration (°Brix)

According to regression analyses (Table 3.8), the higher determination coefficient (R<sup>2</sup>) values were obtained for sumac concentrates by exponential decay model. R<sup>2</sup> values changed between 0.906 and 0.997. In the model equation, y-axis shows that the concentration (°Brix) decays with x-axis (time).

The change in °Brix as a function of time at different temperatures for higher sumac concentrates with initial concentration between 25 and 40 °Brix are shown in Figure 3.3.

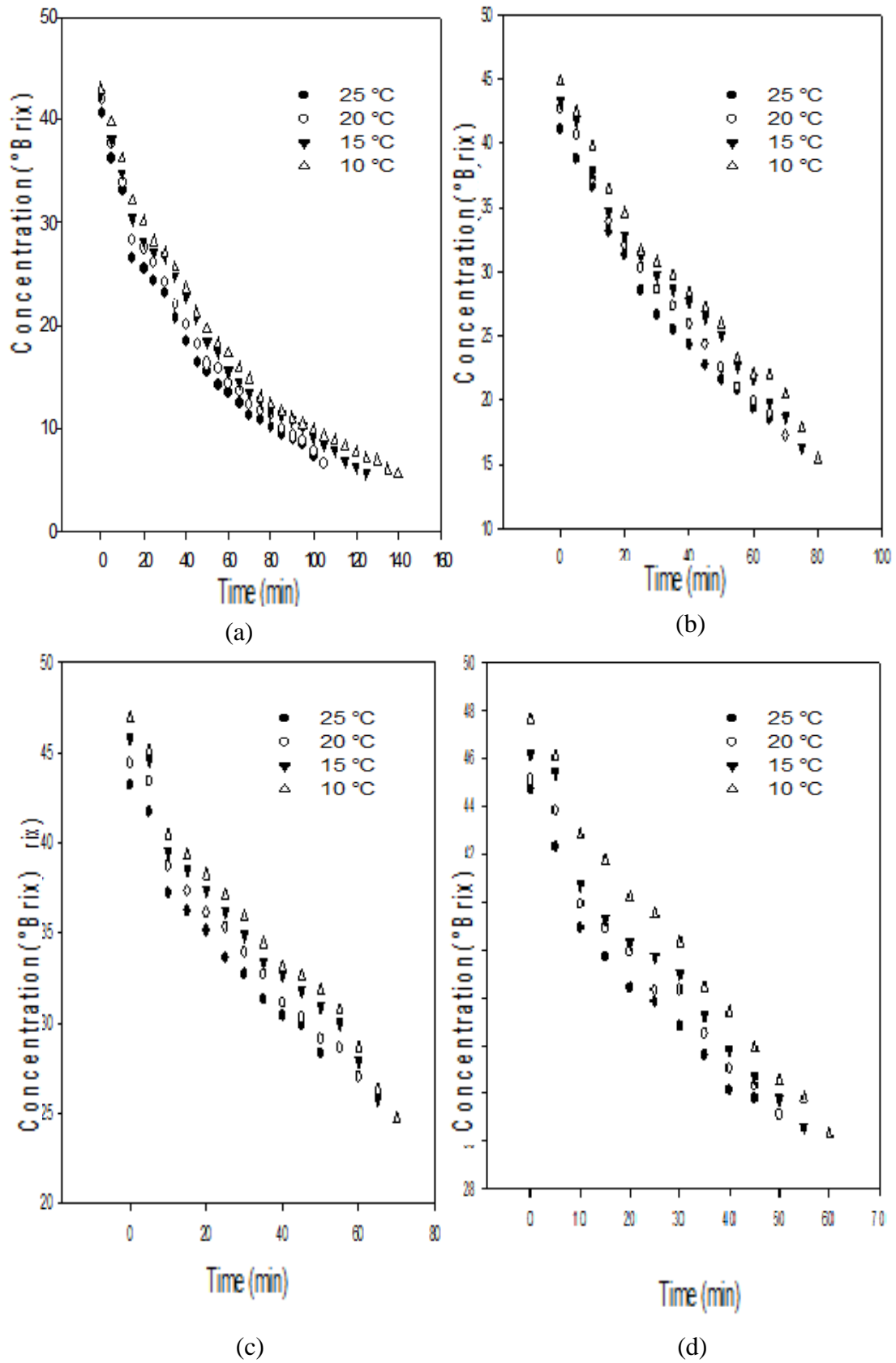


Figure 3.3. Variation of concentration with time for sumac concentrates at different temperatures: (a) initial concentration 25 °Brix, (b) initial concentration 30 °Brix, (c) initial concentration 35 °Brix, (d) initial concentration 40 °Brix

Table 3.9. Regression analyses for sumac concentrates (initial concentration 25, 30, 35 and 40 °Brix) at different temperatures

<b>Initial Concentration (°Brix)</b>	<b>Temperature (°C)</b>	<b>Equation</b>	<b>R<sup>2</sup></b>
<b>25</b>	<b>10</b>	y= 44.81*exp(-0.016*x)	0.998
	<b>15</b>	y= 43.16*exp(-0.016*x)	0.997
	<b>20</b>	y= 42.28*exp(-0.017*x)	0.994
	<b>25</b>	y= 41.40*exp(-0.018*x)	0.992
<b>30</b>	<b>10</b>	y= 44.38*exp(-0.012*x)	0.989
	<b>15</b>	y= 42.95*exp(-0.012*x)	0.988
	<b>20</b>	y= 42.29*exp(-0.013*x)	0.996
	<b>25</b>	y= 40.75*exp(-0.013*x)	0.993
<b>35</b>	<b>10</b>	y= 45.72*exp(-0.008*x)	0.975
	<b>15</b>	y= 44.65*exp(-0.008*x)	0.972
	<b>20</b>	y= 43.45*exp(-0.008*x)	0.976
	<b>25</b>	y= 42.09*exp(-0.008*x)	0.964
<b>40</b>	<b>10</b>	y= 47.12*exp(-0.007*x)	0.993
	<b>15</b>	y= 45.36*exp(-0.007*x)	0.968
	<b>20</b>	y= 44.31*exp(-0.007*x)	0.973
	<b>25</b>	y= 43.31*exp(-0.008*x)	0.959

\* x= Time (min)

\* y= Concentration (°Brix)

Table 3.9 shows the data fitted by non-linear regression analysis at different temperatures for higher sumac concentrates. The determination of coefficient (R<sup>2</sup>) values changed between 0.959 and 0.998. This model was observed to be the best fitted model for all data.

### **3.5. Effect of Temperature on the Concentration of Apple Concentrates**

The change in °Brix as a function of time at different temperatures for lower apple concentrates with a constant initial concentration illustrates Figure 3.4.

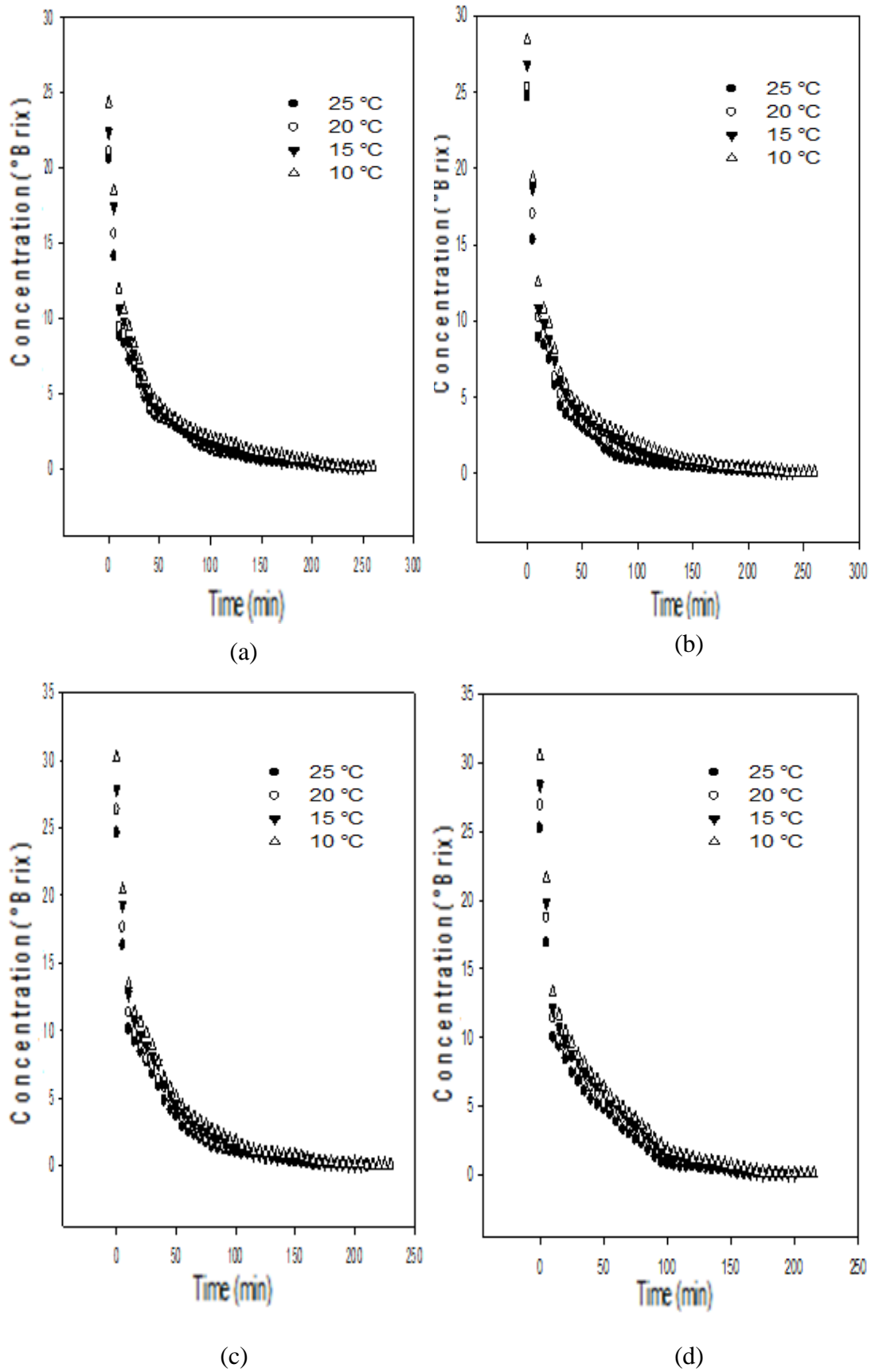


Figure 3.4. Variation of concentration with time for apple concentrates at different temperatures: (a) initial concentration 1 °Brix, (b) initial concentration 2 °Brix, (c) initial concentration 3 °Brix, (d) initial concentration 4 °Brix

Table 3.10. Regression analyses for apple concentrates (initial concentration 1, 2, 3 and 4 °Brix) at different temperatures

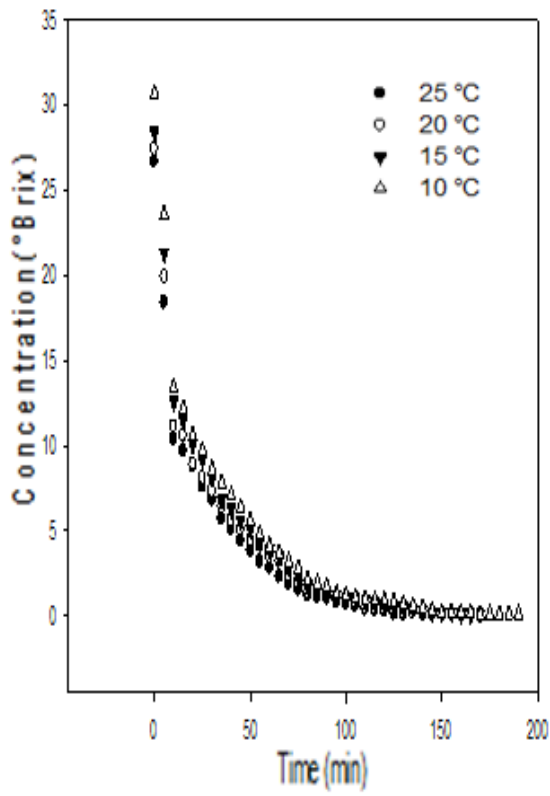
<b>Initial Concentration (°Brix)</b>	<b>Temperature (°C)</b>	<b>Equation</b>	<b>R<sup>2</sup></b>
<b>1</b>	<b>10</b>	y= 20.61*exp(-0.032*x)	0.945
	<b>15</b>	y= 19.26*exp(-0.033*x)	0.950
	<b>20</b>	y= 17.84*exp(-0.034*x)	0.947
	<b>25</b>	y= 17.03*exp(-0.035*x)	0.941
<b>2</b>	<b>10</b>	y= 23.85*exp(-0.038*x)	0.937
	<b>15</b>	y= 22.93*exp(-0.042*x)	0.935
	<b>20</b>	y= 22.02*exp(-0.046*x)	0.945
	<b>25</b>	y= 21.52*exp(-0.053*x)	0.946
<b>3</b>	<b>10</b>	y= 24.91*exp(-0.035*x)	0.949
	<b>15</b>	y= 23.42*exp(-0.036*x)	0.954
	<b>20</b>	y= 22.06*exp(-0.038*x)	0.950
	<b>25</b>	y= 20.83*exp(-0.040*x)	0.952
<b>4</b>	<b>10</b>	y= 24.31*exp(-0.030*x)	0.931
	<b>15</b>	y= 22.48*exp(-0.030*x)	0.927
	<b>20</b>	y= 21.66*exp(-0.032*x)	0.929
	<b>25</b>	y= 20.47*exp(-0.035*x)	0.928

\* x= Time (min)

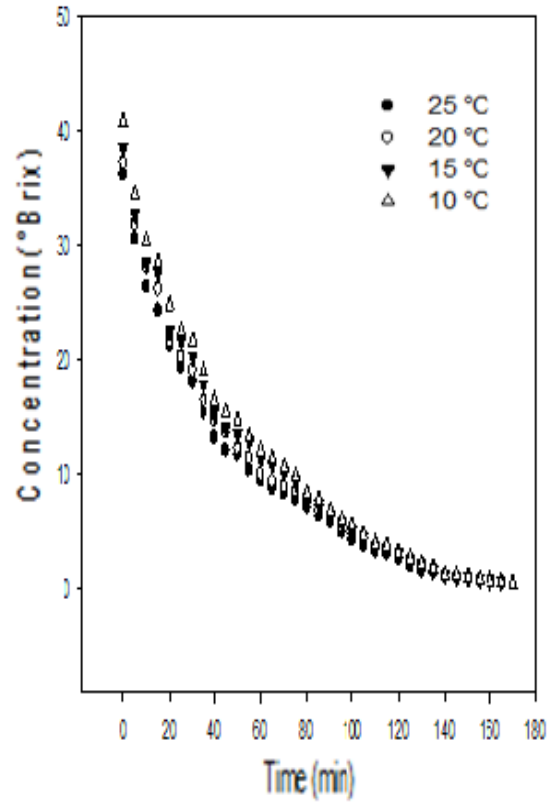
\* y= Concentration (°Brix)

By statistical analyses of the results (Table 3.10), the best fit was obtained by exponential decay model and of R<sup>2</sup> values were obtained between 0.928–0.954. Results indicated that equation coefficient decreases with increasing temperature. This is clearly shown that lower temperatures exhibit higher degree of separation.

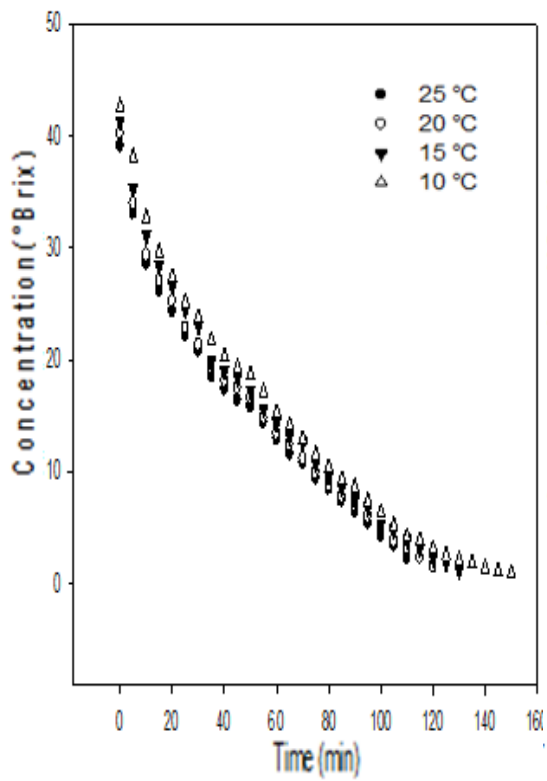
Figure 3.5 illustrates the change in °Brix as a function of time at different temperatures for apple concentrates with initial concentration between 5 and 20 °Brix.



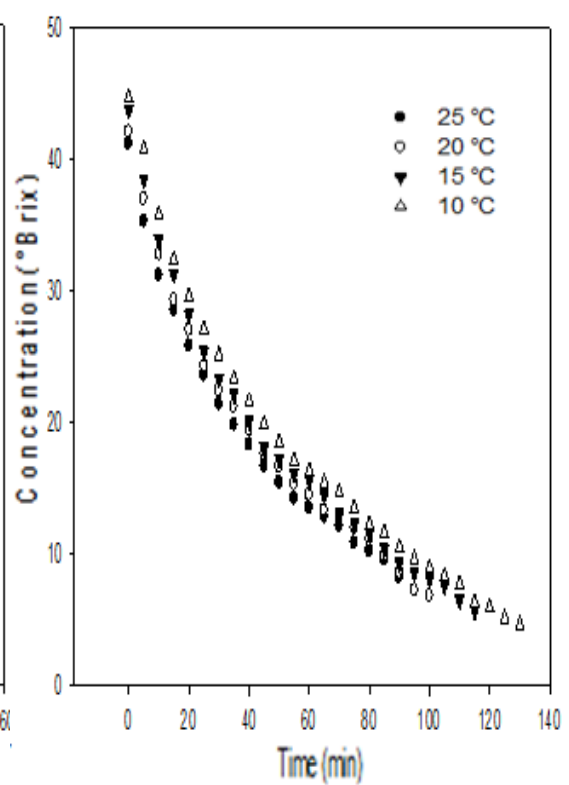
(a)



(b)



(c)



(d)

Figure 3.5. Variation of concentration with time for apple concentrates at different temperatures: (a) initial concentration 5 °Brix, (b) initial concentration 10 °Brix, (c) initial concentration 15 °Brix, (d) initial concentration 20 °Brix

Table 3.11. Regression analyses for apple concentrates (initial concentration 5, 10s, 15 and 20 °Brix) at different temperatures

<b>Initial Concentration (°Brix)</b>	<b>Temperature (°C)</b>	<b>Equation</b>	<b>R<sup>2</sup></b>
<b>5</b>	<b>10</b>	y= 26.66*exp(-0.037*x)	0.954
	<b>15</b>	y= 24.73*exp(-0.037*x)	0.959
	<b>20</b>	y= 23.71*exp(-0.040*x)	0.947
	<b>25</b>	y= 23.19*exp(-0.044*x)	0.949
<b>10</b>	<b>10</b>	y= 38.79*exp(-0.020*x)	0.994
	<b>15</b>	y= 36.57*exp(-0.020*x)	0.993
	<b>20</b>	y= 35.46*exp(-0.021*x)	0.995
	<b>25</b>	y= 34.04*exp(-0.021*x)	0.993
<b>15</b>	<b>10</b>	y= 41.34*exp(-0.018*x)	0.987
	<b>15</b>	y= 39.43*exp(-0.018*x)	0.986
	<b>20</b>	y= 37.86*exp(-0.019*x)	0.984
	<b>25</b>	y= 36.74*exp(-0.019*x)	0.986
<b>20</b>	<b>10</b>	y= 42.68*exp(-0.016*x)	0.994
	<b>15</b>	y= 41.11*exp(-0.017*x)	0.992
	<b>20</b>	y= 39.65*exp(-0.017*x)	0.990
	<b>25</b>	y= 38.38*exp(-0.018*x)	0.988

\* x= Time (min)

\* y= Concentration (°Brix)

Table 3.11 shows that the higher determination coefficient (R<sup>2</sup>) values were obtained for apple concentrates by exponential decay model. R<sup>2</sup> values changed between 0.947 and 0.995. In the model equation, y-axis shows that the concentration (°Brix) decays with x-axis (time).

Temperature effect on the separation of concentrated juice is seen clearly in Figure 3.6. Frozen concentrates thawed at lower temperatures and concentrations were observed to exhibit higher degree of separation.

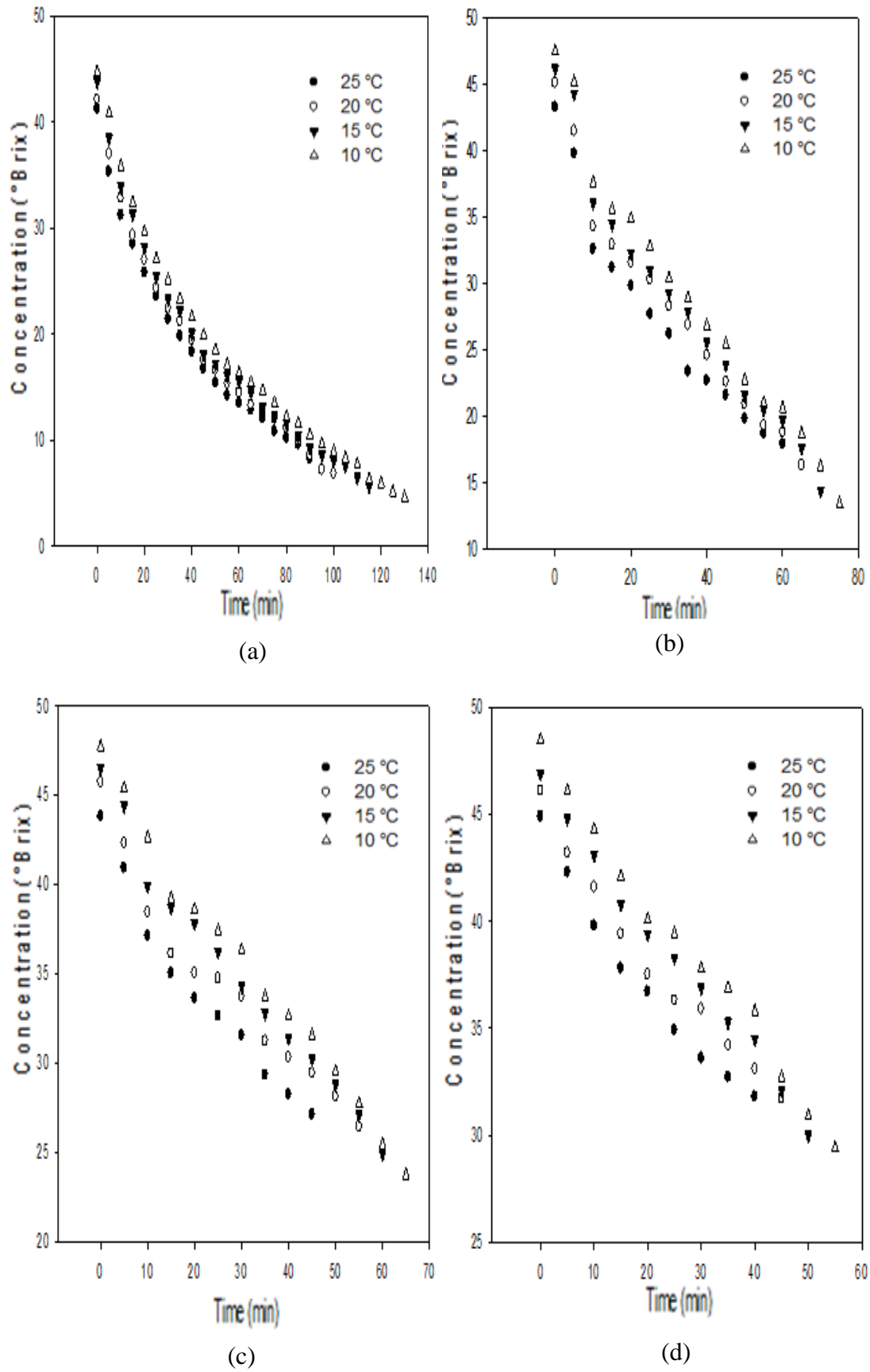


Figure 3.6. Variation of concentration with time for apple concentrates at different temperatures: (a) initial concentration 25 °Brix, (b) initial concentration 30 °Brix, (c) initial concentration 35 °Brix, (d) initial concentration 40 °Brix



Table 3.12. Regression analyses for apple concentrates (initial concentration 25, 30, 35 and 40 °Brix) at different temperatures

Initial Concentration (°Brix)	Temperature (°C)	Equation	R <sup>2</sup>
25	10	$y = 46.82 \cdot \exp(-0.017 \cdot x)$	0.994
	15	$y = 44.95 \cdot \exp(-0.017 \cdot x)$	0.996
	20	$y = 42.95 \cdot \exp(-0.018 \cdot x)$	0.995
	25	$y = 41.53 \cdot \exp(-0.018 \cdot x)$	0.997
30	10	$y = 46.72 \cdot \exp(-0.015 \cdot x)$	0.983
	15	$y = 45.11 \cdot \exp(-0.015 \cdot x)$	0.979
	20	$y = 43.20 \cdot \exp(-0.015 \cdot x)$	0.978
	25	$y = 41.13 \cdot \exp(-0.015 \cdot x)$	0.971
35	10	$y = 47.41 \cdot \exp(-0.009 \cdot x)$	0.987
	15	$y = 45.74 \cdot \exp(-0.009 \cdot x)$	0.988
	20	$y = 43.65 \cdot \exp(-0.009 \cdot x)$	0.964
	25	$y = 42.51 \cdot \exp(-0.011 \cdot x)$	0.976
40	10	$y = 48.33 \cdot \exp(-0.008 \cdot x)$	0.987
	15	$y = 46.79 \cdot \exp(-0.008 \cdot x)$	0.992
	20	$y = 45.13 \cdot \exp(-0.008 \cdot x)$	0.985
	25	$y = 44.02 \cdot \exp(-0.009 \cdot x)$	0.983

\* x= Time (min)

\* y= Concentration (°Brix)

Table 3.12 shows the data fitted by non-linear regression analysis at different temperatures for higher apple concentrates. The determination of coefficient (R<sup>2</sup>) values changed between 0.964 and 0.997.

### 3.6. Effect of Temperature on the Concentration of Grape Concentrates

Figure 3.7 illustrates the change in °Brix as a function of time at different temperatures for lower grape concentrates with a constant initial concentration. The observed effect of initial concentration was similar to other concentrates.

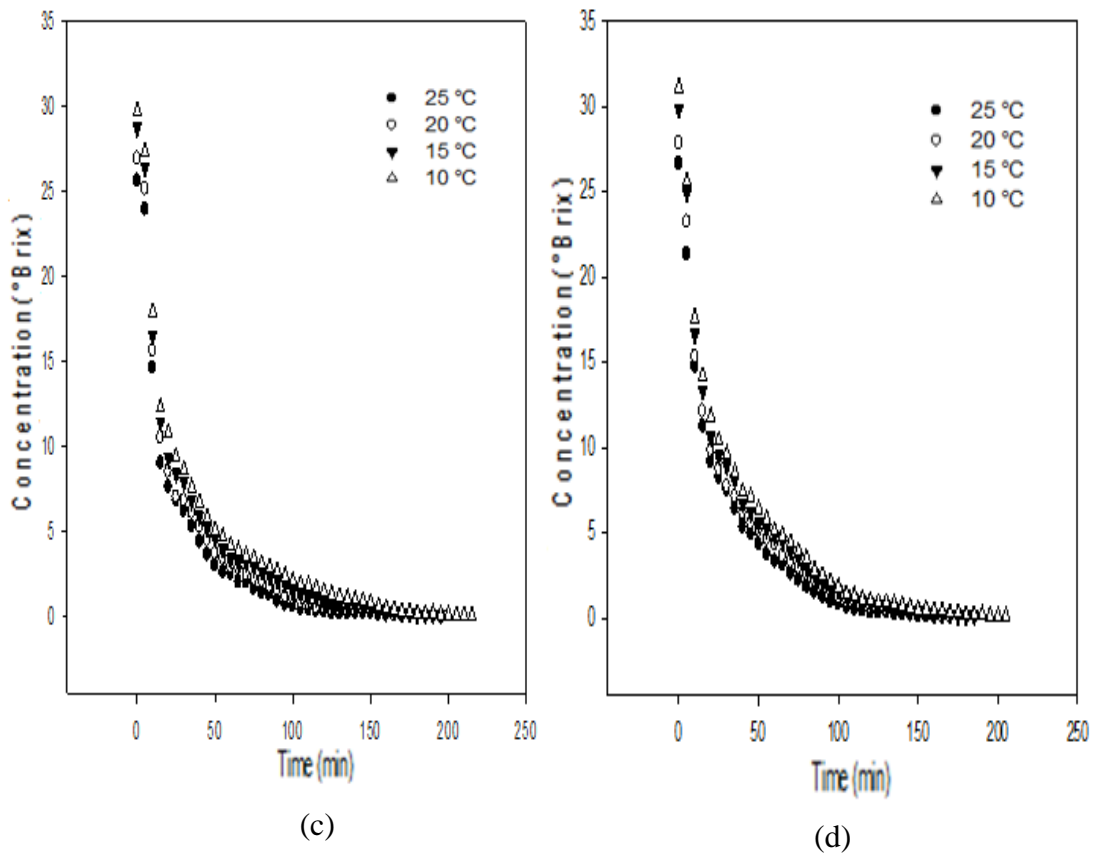
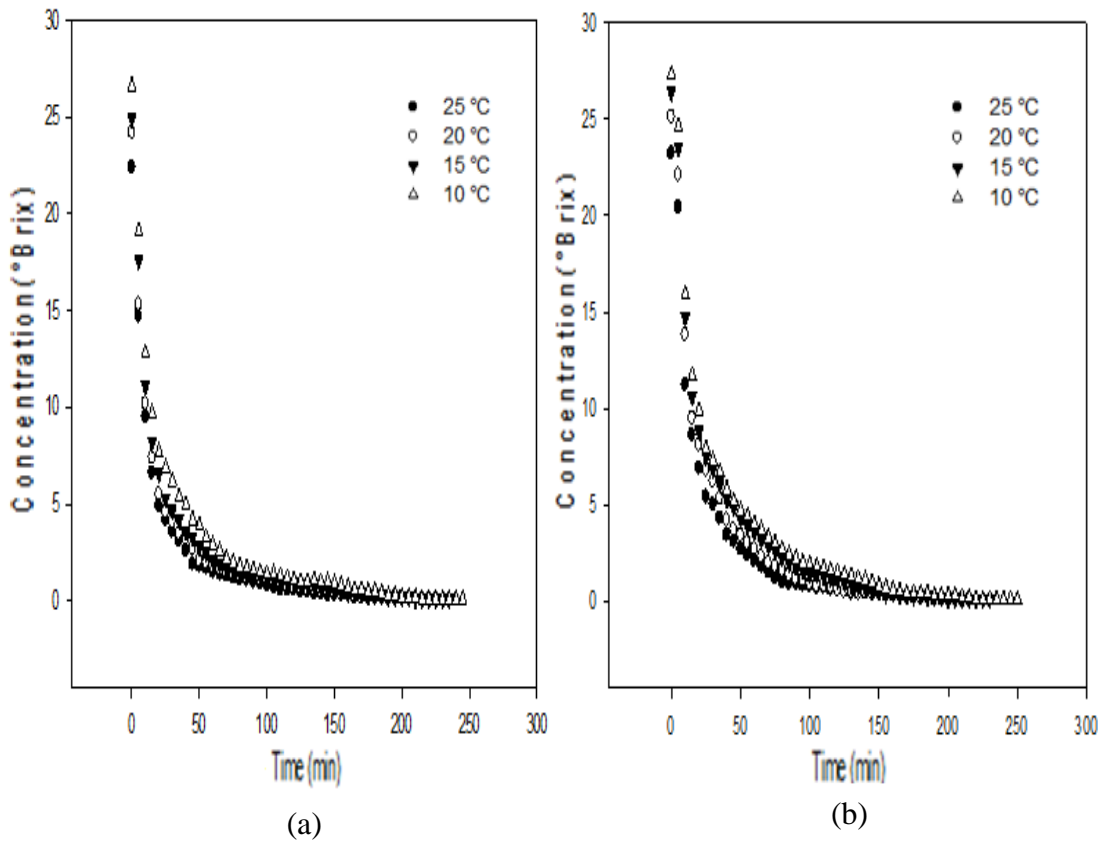


Figure 3.7. Variation of concentration with time for grape concentrates at different temperatures: (a) initial concentration 1 °Brix, (b) initial concentration 2 °Brix, (c) initial concentration 3 °Brix, (d) initial concentration 4 °Brix

Table 3.13. Regression analyses for grape concentrates (initial concentration 1, 2, 3 and 4 °Brix) at different temperatures

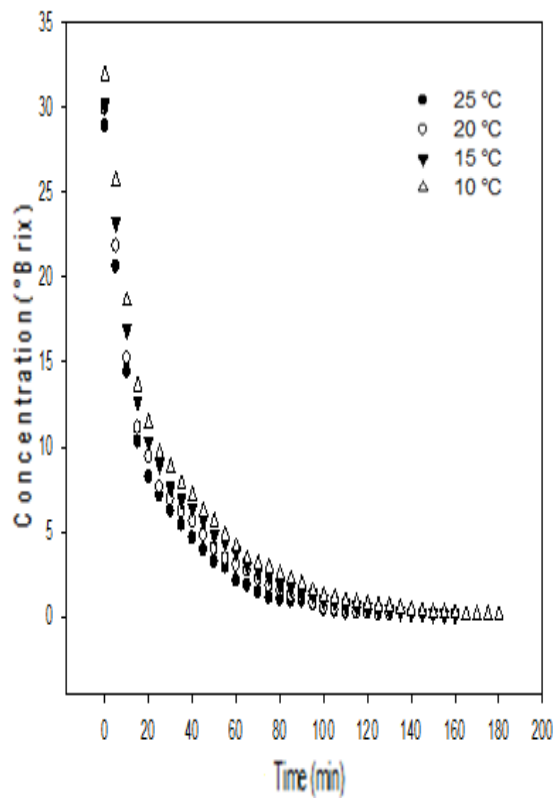
Initial Concentration (°Brix)	Temperature (°C)	Equation	R <sup>2</sup>
1	10	y= 23.68*exp(-0.044*x)	0.954
	15	y= 23.00*exp(-0.055*x)	0.958
	20	y= 22.26*exp(-0.062*x)	0.959
	25	y= 21.10*exp(-0.066*x)	0.968
2	10	y= 25.73*exp(-0.037*x)	0.957
	15	y= 25.04*exp(-0.040*x)	0.960
	20	y= 24.56*exp(-0.046*x)	0.972
	25	y= 22.97*exp(-0.052*x)	0.969
3	10	y= 28.15*exp(-0.036*x)	0.957
	15	y= 27.71*exp(-0.041*x)	0.958
	20	y= 26.74*exp(-0.050*x)	0.964
	25	y= 25.93*exp(-0.050*x)	0.967
4	10	y= 27.84*exp(-0.033*x)	0.969
	15	y= 27.02*exp(-0.035*x)	0.966
	20	y= 25.50*exp(-0.037*x)	0.967
	25	y= 24.69*exp(-0.040*x)	0.977

\* x= Time (min)

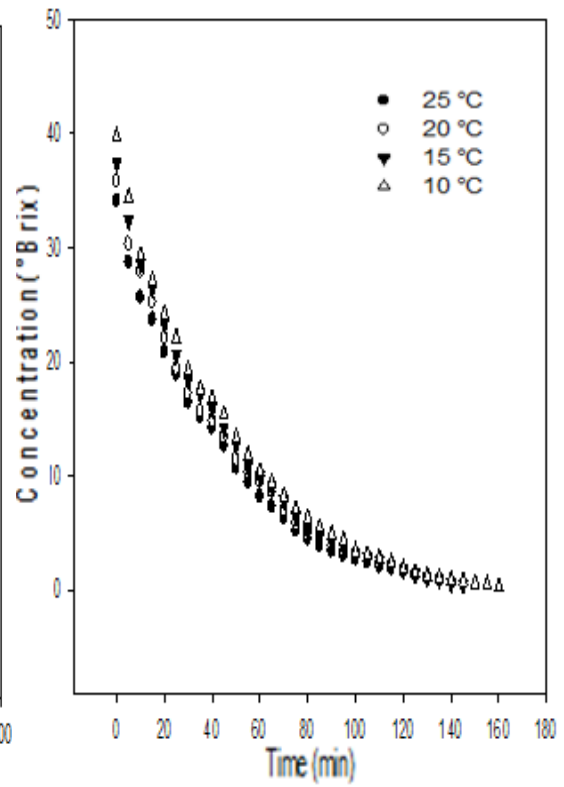
\* y= Concentration (°Brix)

Table 3.13 shows that the higher (R<sup>2</sup>) values were obtained for apple concentrates by exponential decay model. R<sup>2</sup> values changed between 0.954 and 0.977. In the model equation, y-axis shows that the concentration (°Brix) decays with x-axis (time).

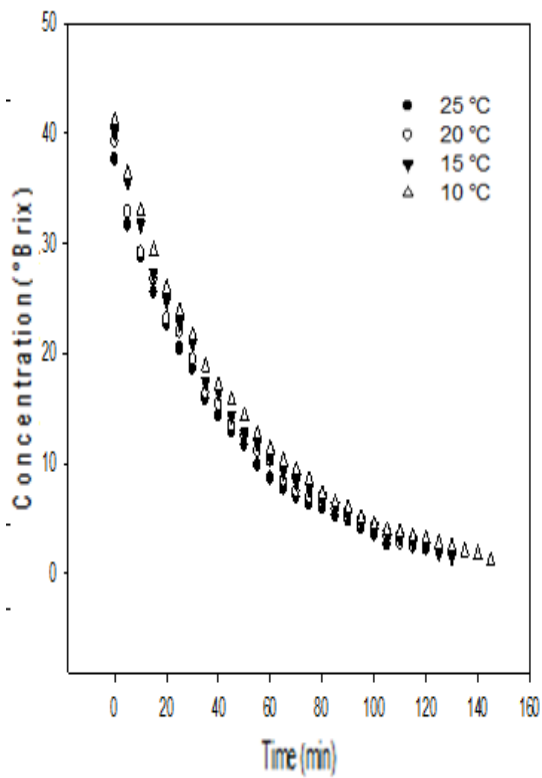
Temperature effect on the separation of concentrated juice is seen in Figure 3.8. The degree of separation was observed to be dependent upon initial concentrations of the juice, also.



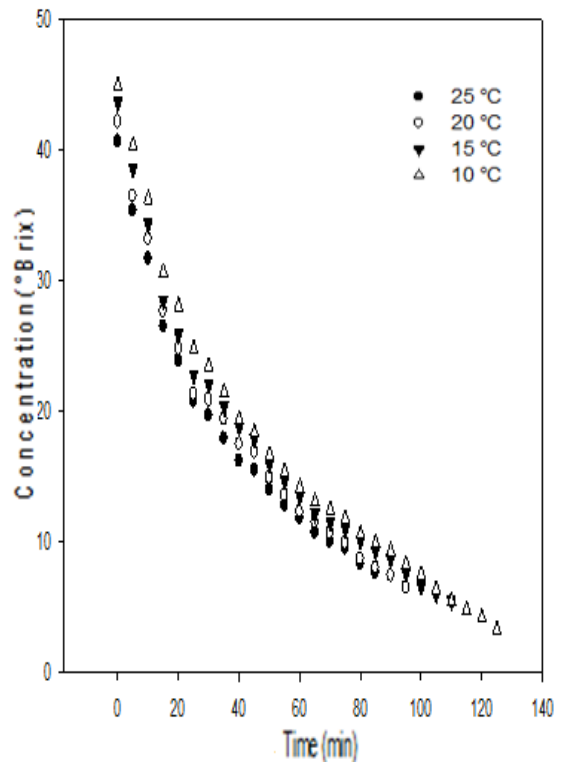
(a)



(b)



(c)



(d)

Figure 3.8. Variation of concentration with time for grape concentrates at different temperatures: (a) initial concentration 5 °Brix, (b) initial concentration 10 °Brix, (c) initial concentration 15 °Brix, (d) initial concentration 20 °Brix

Table 3.14 shows that the best fit was obtained for grape concentrates between initial concentrations 10 and 20 °Brix due to their higher R<sup>2</sup> values ranging from 0.973 to 0.999. Results indicated that, the higher determination coefficients are obtained from grape concentrates

Table 3.14. Regression analyses for grape concentrates (initial concentration 5, 10, 15 and 20 °Brix) at different temperatures

<b>Initial Concentration (°Brix)</b>	<b>Temperature (°C)</b>	<b>Equation</b>	<b>R<sup>2</sup></b>
<b>5</b>	<b>10</b>	y= 29.58*exp(-0.039*x)	0.978
	<b>15</b>	y= 27.76*exp(-0.040*x)	0.978
	<b>20</b>	y= 27.44*exp(-0.046*x)	0.973
	<b>25</b>	y= 26.97*exp(-0.052*x)	0.980
<b>10</b>	<b>10</b>	y= 38.80*exp(-0.023*x)	0.996
	<b>15</b>	y= 37.08*exp(-0.023*x)	0.996
	<b>20</b>	y= 35.32*exp(-0.023*x)	0.997
	<b>25</b>	y= 33.50*exp(-0.024*x)	0.996
<b>15</b>	<b>10</b>	y= 40.76*exp(-0.022*x)	0.999
	<b>15</b>	y= 39.64*exp(-0.022*x)	0.998
	<b>20</b>	y= 37.73*exp(-0.023*x)	0.997
	<b>25</b>	y= 36.62*exp(-0.024*x)	0.998
<b>20</b>	<b>10</b>	y= 42.88*exp(-0.018*x)	0.992
	<b>15</b>	y= 40.80*exp(-0.019*x)	0.986
	<b>20</b>	y= 39.62*exp(-0.020*x)	0.986
	<b>25</b>	y= 38.39*exp(-0.021*x)	0.986

\* x= Time (min)

\* y= Concentration (°Brix)

It was obvious in the Figure 3.9 that the higher the initial concentration of the juice, the lower degree of separation was achieved. This method is, therefore, not feasible for samples having an initial concentration higher than 20 °Brix.

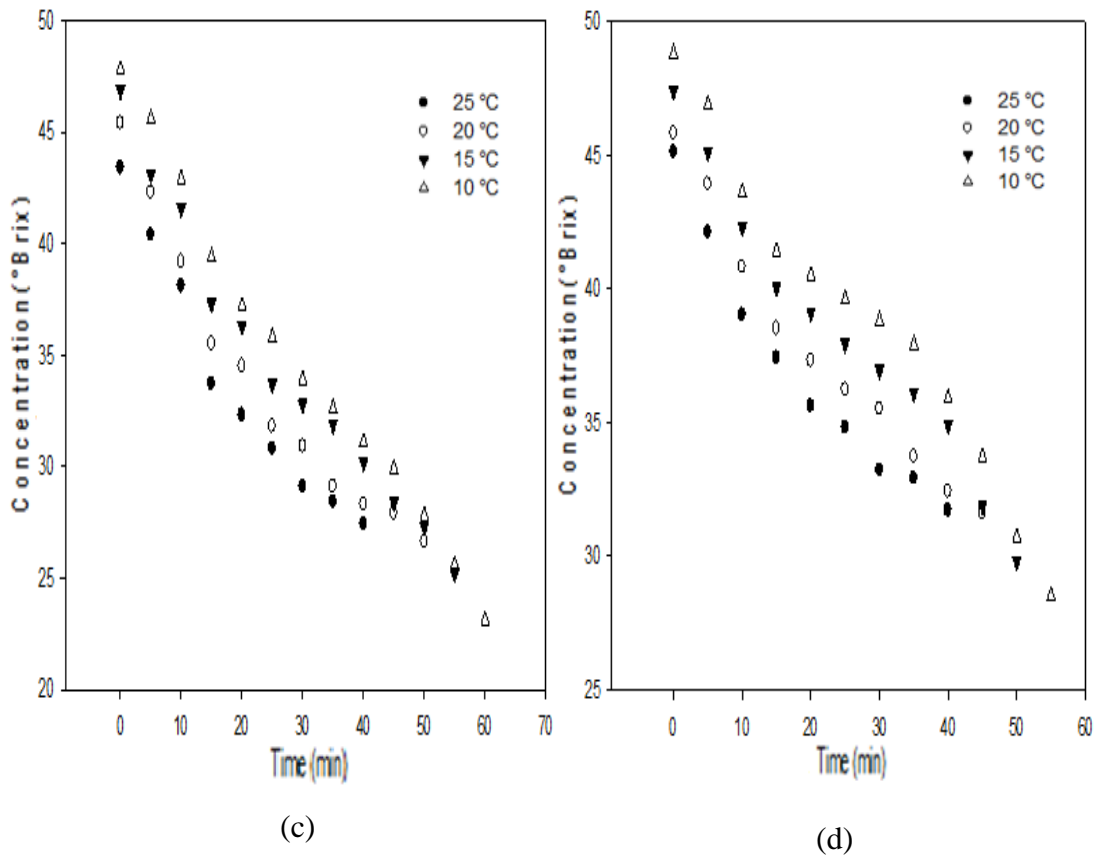
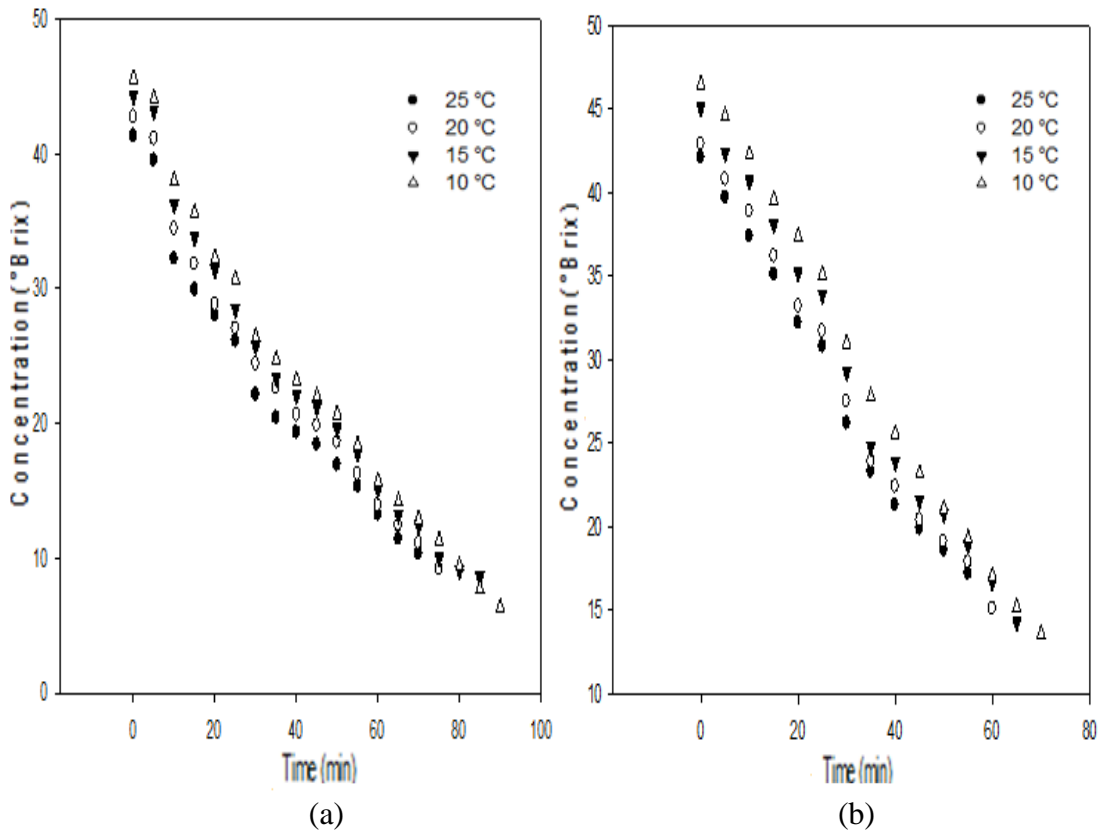


Figure 3.9. Variation of concentration with time for grape concentrates at different temperatures: (a) initial concentration 25 °Brix, (b) initial concentration 30 °Brix, (c) initial concentration 35 °Brix, (d) initial concentration 40 °Brix

Table 3.15. Regression analyses for grape concentrates (initial concentration 25, 30, 35 and 40 °Brix) at different temperatures

Initial Concentration (°Brix)	Temperature (°C)	Equation	R <sup>2</sup>
25	10	$y = 47.01 \cdot \exp(-0.018 \cdot x)$	0.989
	15	$y = 45.19 \cdot \exp(-0.018 \cdot x)$	0.991
	20	$y = 42.99 \cdot \exp(-0.019 \cdot x)$	0.991
	25	$y = 41.16 \cdot \exp(-0.019 \cdot x)$	0.989
30	10	$y = 49.34 \cdot \exp(-0.017 \cdot x)$	0.983
	15	$y = 47.15 \cdot \exp(-0.017 \cdot x)$	0.982
	20	$y = 44.92 \cdot \exp(-0.017 \cdot x)$	0.984
	25	$y = 43.59 \cdot \exp(-0.017 \cdot x)$	0.986
35	10	$y = 47.68 \cdot \exp(-0.011 \cdot x)$	0.992
	15	$y = 45.67 \cdot \exp(-0.011 \cdot x)$	0.986
	20	$y = 43.94 \cdot \exp(-0.011 \cdot x)$	0.968
	25	$y = 42.565 \cdot \exp(-0.012 \cdot x)$	0.970
40	10	$y = 48.55 \cdot \exp(-0.008 \cdot x)$	0.963
	15	$y = 46.69 \cdot \exp(-0.008 \cdot x)$	0.974
	20	$y = 44.98 \cdot \exp(-0.008 \cdot x)$	0.978
	25	$y = 43.69 \cdot \exp(-0.009 \cdot x)$	0.958

\* x= Time (min)

\* y= Concentration (°Brix)

Table 3.15 shows that the higher (R<sup>2</sup>) values were obtained for apple concentrates by exponential decay model. R<sup>2</sup> values changed between 0.958 and 0.992. In the model equation, y-axis shows that the concentration (°Brix) decays with x-axis (time).

### 3.7. Effect of Temperature on the Concentration of Carrot Concentrates

Figures (3.10–3.12) illustrate the change in °Brix as a function of time at different temperatures for carrot concentrates with a constant initial concentration.

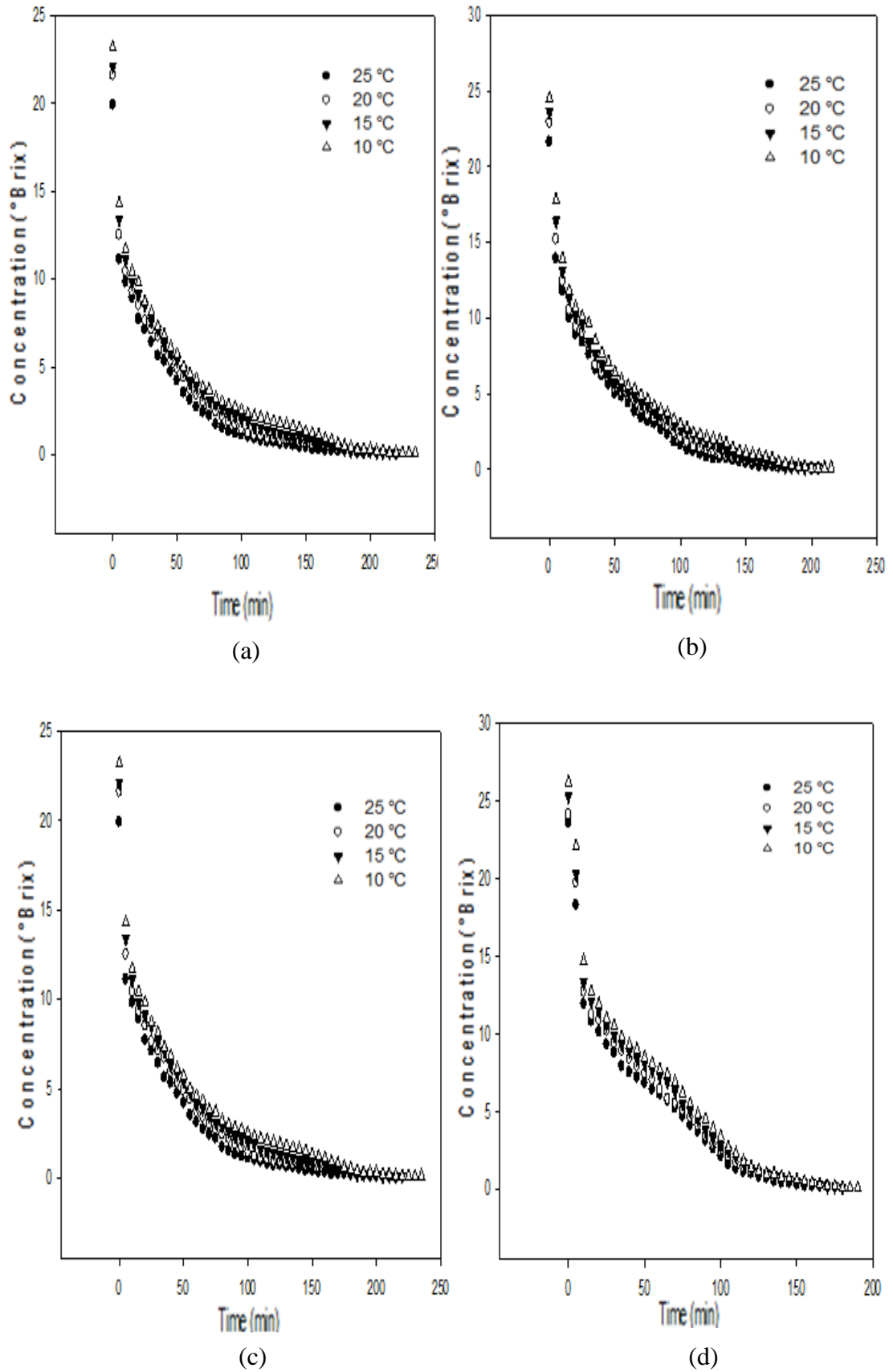


Figure 3.10. Variation of concentration with time for carrot concentrates at different temperatures: (a) initial concentration 1 °Brix, (b) initial concentration 2 °Brix, (c) initial concentration 3 °Brix, (d) initial concentration 4 °Brix



Table 3.16. Regression analyses for carrot concentrates (initial concentration 1, 2, 3 and 4 °Brix) at different temperatures

<b>Initial Concentration (°Brix)</b>	<b>Temperature (°C)</b>	<b>Equation</b>	<b>R<sup>2</sup></b>
<b>1</b>	<b>10</b>	y= 17.41*exp(-0.022*x)	0.939
	<b>15</b>	y= 16.94*exp(-0.024*x)	0.947
	<b>20</b>	y= 16.61*exp(-0.027*x)	0.943
	<b>25</b>	y= 15.67*exp(-0.029*x)	0.950
<b>2</b>	<b>10</b>	y= 19.58*exp(-0.022*x)	0.958
	<b>15</b>	y= 18.73*exp(-0.023*x)	0.954
	<b>20</b>	y= 18.20*exp(-0.025*x)	0.954
	<b>25</b>	y= 17.40*exp(-0.026*x)	0.960
<b>3</b>	<b>10</b>	y= 20.41*exp(-0.022*x)	0.938
	<b>15</b>	y= 19.27*exp(-0.022*x)	0.928
	<b>20</b>	y= 18.95*exp(-0.025*x)	0.939
	<b>25</b>	y= 18.80*exp(-0.028*x)	0.956
<b>4</b>	<b>10</b>	y= 21.81*exp(-0.020*x)	0.945
	<b>15</b>	y= 20.67*exp(-0.021*x)	0.943
	<b>20</b>	y= 19.98*exp(-0.022*x)	0.944
	<b>25</b>	y= 19.15*exp(-0.023*x)	0.936

\* x= Time (min)

\* y= Concentration (°Brix)

Table 3.16 shows the data fitted by non-linear regression analysis at different temperatures for lower carrot concentrates. The determination of coefficient (R<sup>2</sup>) values changed between 0.928 and 0.960. The higher R<sup>2</sup> values were obtained by using exponential decay model. This model was observed to be the best fitted model for all data. In the model equation, y-axis shows that the concentration (°Brix) decays with x-axis (time). Negative exponent indicates exponential decay.

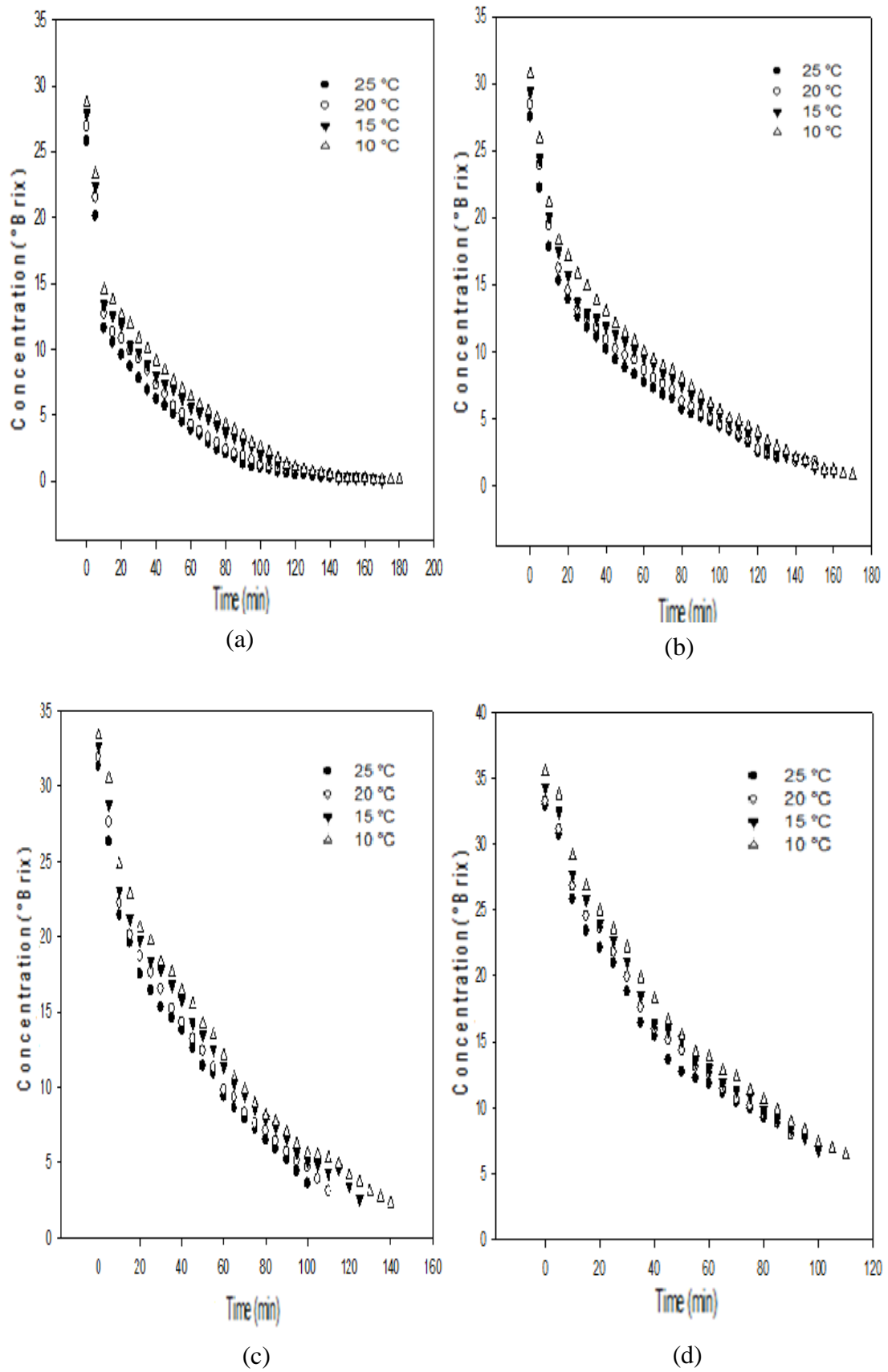


Figure 3.11. Variation of concentration with time for carrot concentrates at different temperatures: (a) initial concentration 5 °Brix, (b) initial concentration 10 °Brix, (c) initial concentration 15 °Brix, (d) initial concentration 20 °Brix

Table 3.17. Regression analyses for carrot concentrates (initial concentration 5, 10, 15 and 20 °Brix) at different temperatures

<b>Initial Concentration (°Brix)</b>	<b>Temperature (°C)</b>	<b>Equation</b>	<b>R<sup>2</sup></b>
<b>5</b>	<b>10</b>	$y = 24.27 \cdot \exp(-0.025 \cdot x)$	0.957
	<b>15</b>	$y = 23.43 \cdot \exp(-0.027 \cdot x)$	0.949
	<b>20</b>	$y = 23.29 \cdot \exp(-0.031 \cdot x)$	0.956
	<b>25</b>	$y = 22.36 \cdot \exp(-0.035 \cdot x)$	0.952
<b>10</b>	<b>10</b>	$y = 26.72 \cdot \exp(-0.017 \cdot x)$	0.975
	<b>15</b>	$y = 25.07 \cdot \exp(-0.017 \cdot x)$	0.964
	<b>20</b>	$y = 24.35 \cdot \exp(-0.018 \cdot x)$	0.964
	<b>25</b>	$y = 23.18 \cdot \exp(-0.019 \cdot x)$	0.958
<b>15</b>	<b>10</b>	$y = 31.47 \cdot \exp(-0.017 \cdot x)$	0.990
	<b>15</b>	$y = 30.03 \cdot \exp(-0.017 \cdot x)$	0.984
	<b>20</b>	$y = 29.20 \cdot \exp(-0.019 \cdot x)$	0.983
	<b>25</b>	$y = 28.28 \cdot \exp(-0.019 \cdot x)$	0.977
<b>20</b>	<b>10</b>	$y = 34.85 \cdot \exp(-0.016 \cdot x)$	0.996
	<b>15</b>	$y = 33.64 \cdot \exp(-0.016 \cdot x)$	0.994
	<b>20</b>	$y = 32.49 \cdot \exp(-0.016 \cdot x)$	0.994
	<b>25</b>	$y = 31.85 \cdot \exp(-0.017 \cdot x)$	0.985

\* x= Time (min)

\* y= Concentration (°Brix)

According to regression analyses (Table 3.17), the lower determination coefficients are obtained from carrot concentrates. The degree of separation was observed to be influenced also by the nature of the juice being concentrated.

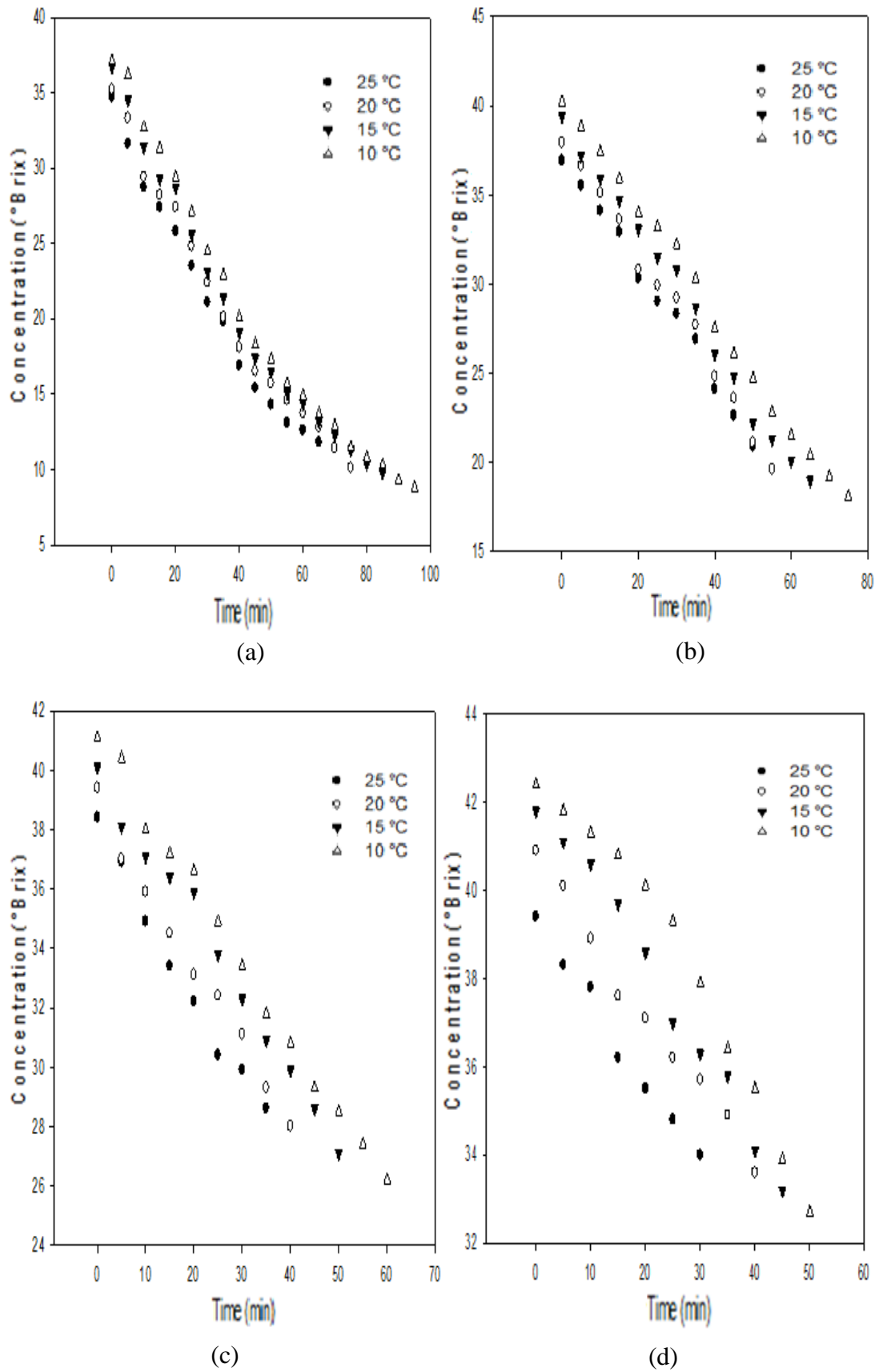


Figure 3.12. Variation of concentration with time for carrot concentrates at different temperatures: (a) initial concentration 25 °Brix, (b) initial concentration 30 °Brix, (c) initial concentration 35 °Brix, (d) initial concentration 40 °Brix

Table 3.18 shows the data fitted by non-linear regression analysis at different temperatures for higher carrot concentrates. The determination of coefficient ( $R^2$ ) values changed between 0.952 and 0.996.

Table 3.18. Regression analyses for carrot concentrates (initial concentration 25, 30, 35 and 40 °Brix) at different temperatures

<b>Initial Concentration (°Brix)</b>	<b>Temperature (°C)</b>	<b>Equation</b>	<b>R<sup>2</sup></b>
<b>25</b>	<b>10</b>	$y = 38.69 \cdot \exp(-0.016 \cdot x)$	0.995
	<b>15</b>	$y = 37.28 \cdot \exp(-0.016 \cdot x)$	0.996
	<b>20</b>	$y = 35.80 \cdot \exp(-0.016 \cdot x)$	0.993
	<b>25</b>	$y = 34.87 \cdot \exp(-0.017 \cdot x)$	0.994
<b>30</b>	<b>10</b>	$y = 41.66 \cdot \exp(-0.011 \cdot x)$	0.985
	<b>15</b>	$y = 40.35 \cdot \exp(-0.011 \cdot x)$	0.981
	<b>20</b>	$y = 38.99 \cdot \exp(-0.011 \cdot x)$	0.977
	<b>25</b>	$y = 37.74 \cdot \exp(-0.011 \cdot x)$	0.981
<b>35</b>	<b>10</b>	$y = 41.62 \cdot \exp(-0.008 \cdot x)$	0.994
	<b>15</b>	$y = 40.29 \cdot \exp(-0.008 \cdot x)$	0.984
	<b>20</b>	$y = 39.02 \cdot \exp(-0.008 \cdot x)$	0.991
	<b>25</b>	$y = 38.26 \cdot \exp(-0.009 \cdot x)$	0.993
<b>40</b>	<b>10</b>	$y = 43.42 \cdot \exp(-0.005 \cdot x)$	0.952
	<b>15</b>	$y = 42.37 \cdot \exp(-0.005 \cdot x)$	0.983
	<b>20</b>	$y = 40.83 \cdot \exp(-0.005 \cdot x)$	0.990
	<b>25</b>	$y = 39.36 \cdot \exp(-0.005 \cdot x)$	0.989

\* x= Time (min)

\* y= Concentration (°Brix)

### 3.8. Effect of the Size of Frozen Samples on the Degree of Separation

Three different sizes of various concentrations were for different juices. Concentration was plotted against time each volume separately. Figure 3.13 indicates the variation of concentration with time for sumac concentrates at different sizes. For

each experiment, concentration decreased by time so exponential decay model was selected.

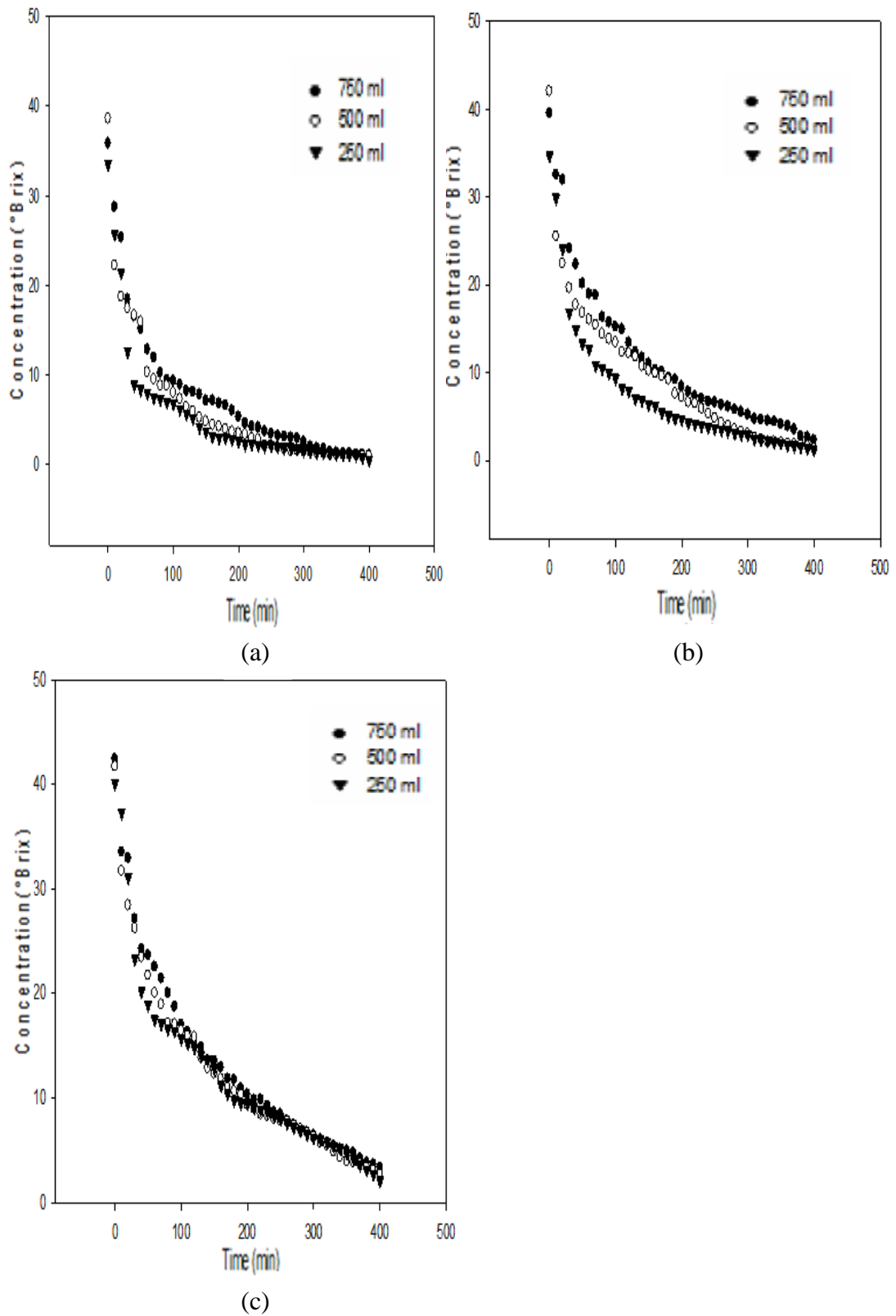


Figure 3.13. Variation of concentration with time for sumac concentrates at different sizes: (a) initial concentration 1 °Brix, (b) initial concentration 5 °Brix, (c) initial concentration 10 °Brix

Table 3.19. Regression analyses for sumac concentrates (initial concentration 1, 5 and °Brix) at different sizes

<b>Initial Concentration (°Brix)</b>	<b>mL</b>	<b>Equation</b>	<b>R<sup>2</sup></b>
<b>1</b>	<b>250</b>	$y = 29.36 * \exp(-0.018 * x)$	0.911
	<b>500</b>	$y = 29.77 * \exp(-0.014 * x)$	0.930
	<b>750</b>	$y = 29.88 * \exp(-0.011 * x)$	0.944
<b>5</b>	<b>250</b>	$y = 29.22 * \exp(-0.011 * x)$	0.928
	<b>500</b>	$y = 29.53 * \exp(-0.008 * x)$	0.912
	<b>750</b>	$y = 33.26 * \exp(-0.007 * x)$	0.961
<b>10</b>	<b>250</b>	$y = 32.61 * \exp(-0.007 * x)$	0.917
	<b>500</b>	$y = 33.11 * \exp(-0.006 * x)$	0.958
	<b>750</b>	$y = 35.16 * \exp(-0.006 * x)$	0.968

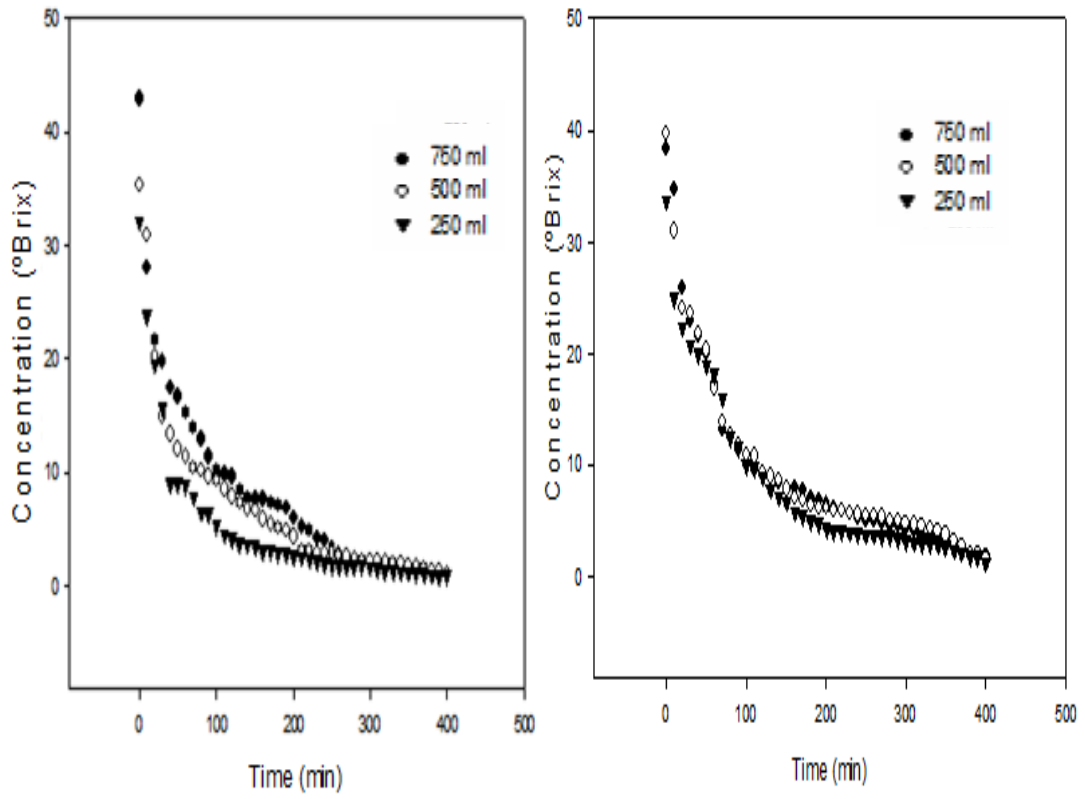
\* x= Time (min)

\* y= Concentration (°Brix)

Table 3.19 presents the regression analysis at different sizes for sumac concentrates. The determination coefficient (R<sup>2</sup>) values changed between 0.911 and 0.968.

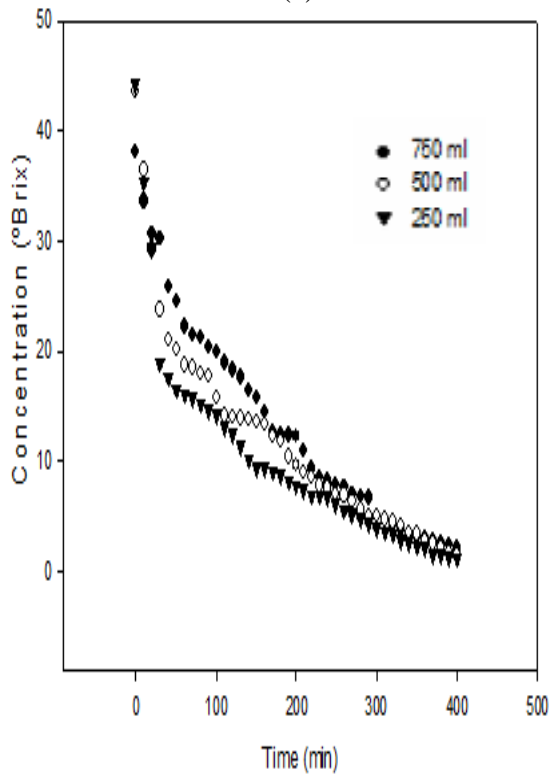
Figures 3.14 illustrates the variation of concentration with time for apple concentrates at different sizes. The size of samples being thawed was observed to affect the degree of separation. The rate of melting increased with increasing surface area of sample. Because a larger surface area contacts with the surrounding air. It can absorb more heat, so more melting occurs. Even though larger surface area melts fast, it may take more time to melt. Because more ice is present during the melting, its surface area decreases so more time requires to melt.

Table 3.20 shows that the regression analysis at different sizes for apple concentrates. The determination coefficient (R<sup>2</sup>) values changed between 0.900 and 0.986. By statistical analyses of the results, the most effective degree of separation of juice from ice was observed at lower temperatures regardless sample size.



(a)

(b)



(c)

Figure 3.14. Variation of concentration with time for apple concentrates at different sizes: (a) initial concentration 1 °Brix, (b) initial concentration 5 °Brix, (c) initial concentration 10 °Brix



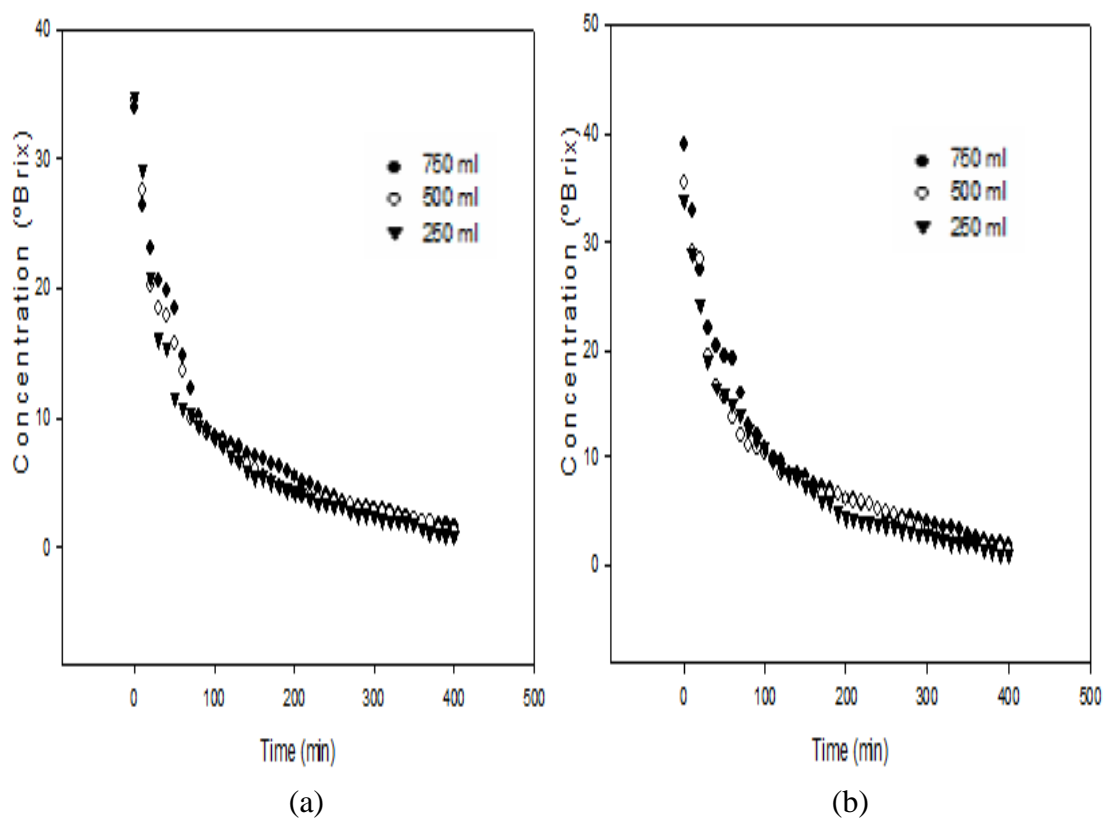
Table 3.20. Regression analyses for apple concentrates (initial concentration 1, 5 and °Brix) at different sizes

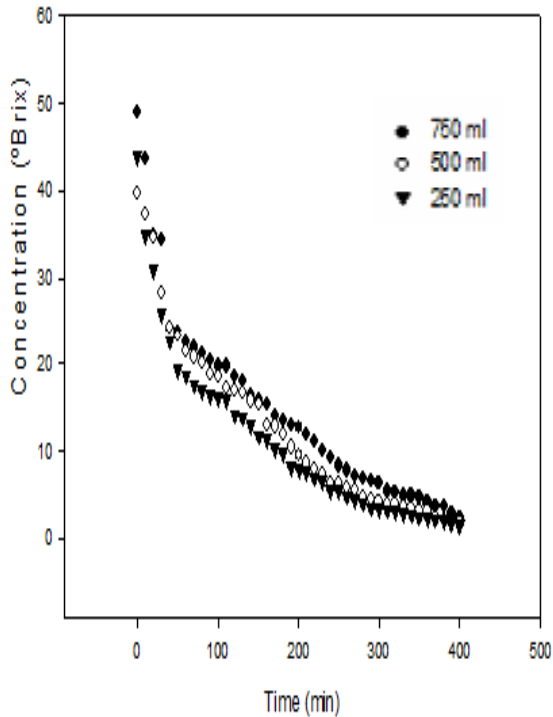
Initial Concentration (°Brix)	mL	Equation	R <sup>2</sup>
1	250	$y = 28.57 \cdot \exp(-0.018 \cdot x)$	0.940
	500	$y = 28.80 \cdot \exp(-0.012 \cdot x)$	0.908
	750	$y = 31.86 \cdot \exp(-0.010 \cdot x)$	0.924
5	250	$y = 29.22 \cdot \exp(-0.009 \cdot x)$	0.971
	500	$y = 32.20 \cdot \exp(-0.009 \cdot x)$	0.931
	750	$y = 33.19 \cdot \exp(-0.009 \cdot x)$	0.937
10	250	$y = 33.52 \cdot \exp(-0.008 \cdot x)$	0.900
	500	$y = 34.27 \cdot \exp(-0.007 \cdot x)$	0.935
	750	$y = 35.50 \cdot \exp(-0.006 \cdot x)$	0.986

\* x= Time (min)

\* y= Concentration (°Brix)

Figures 3.15 illustrates the variation of concentration with time for grape concentrates at different sizes. Concentration decreased by time so exponential decay model was selected.





(c)

Figure 3.15. Variation of concentration with time for grape concentrates at different sizes: (a) initial concentration 1 °Brix, (b) initial concentration 5 °Brix, (c) initial concentration 10 °Brix

Table 3.21. Regression analyses for grape concentrates (initial concentration 1, 5 and °Brix) at different sizes

Initial Concentration (°Brix)	mL	Equation	R <sup>2</sup>
1	250	$y = 28.86 \cdot \exp(-0.012 \cdot x)$	0.921
	500	$y = 28.89 \cdot \exp(-0.011 \cdot x)$	0.942
	750	$y = 28.90 \cdot \exp(-0.010 \cdot x)$	0.953
5	250	$y = 29.41 \cdot \exp(-0.010 \cdot x)$	0.969
	500	$y = 29.82 \cdot \exp(-0.010 \cdot x)$	0.931
	750	$y = 33.34 \cdot \exp(-0.009 \cdot x)$	0.955
10	250	$y = 36.13 \cdot \exp(-0.008 \cdot x)$	0.961
	500	$y = 37.19 \cdot \exp(-0.007 \cdot x)$	0.981
	750	$y = 40.75 \cdot \exp(-0.007 \cdot x)$	0.947

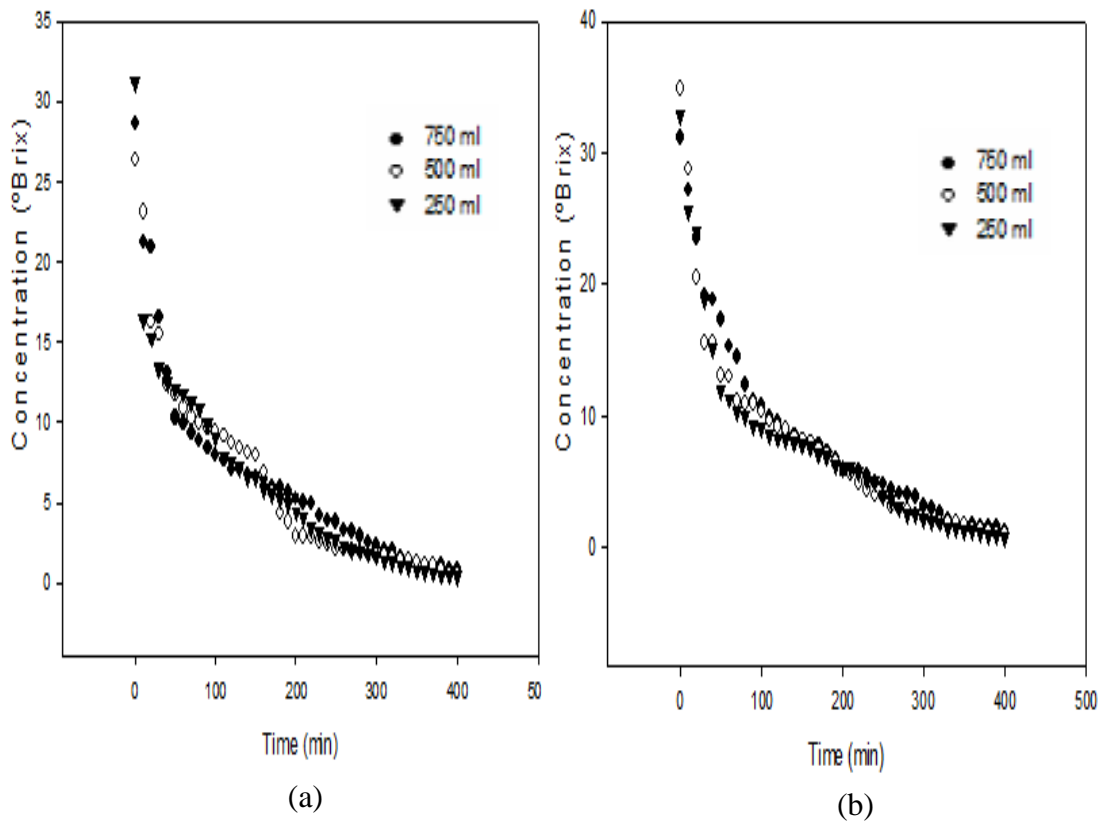
\* x= Time (min)

\* y= Concentration (°Brix)

Table 3.21 shows that the regression analysis at different sizes for grape concentrates. The determination coefficient ( $R^2$ ) values changed between 0.921 and 0.981. The regression analysis shows that when volume size increased, higher °Brix value was obtained.

This is due to the decrease in  $(\text{surface area}/\text{volume})_{\text{SAMPLES}}$  as the size increases, which causes a relatively less heat transfer and consequently slower rate of thawing, resulting in a more concentrated juice leaving the ice front.

Figures 3.16 illustrates the variation of concentration with time for carrot concentrates at different sizes.



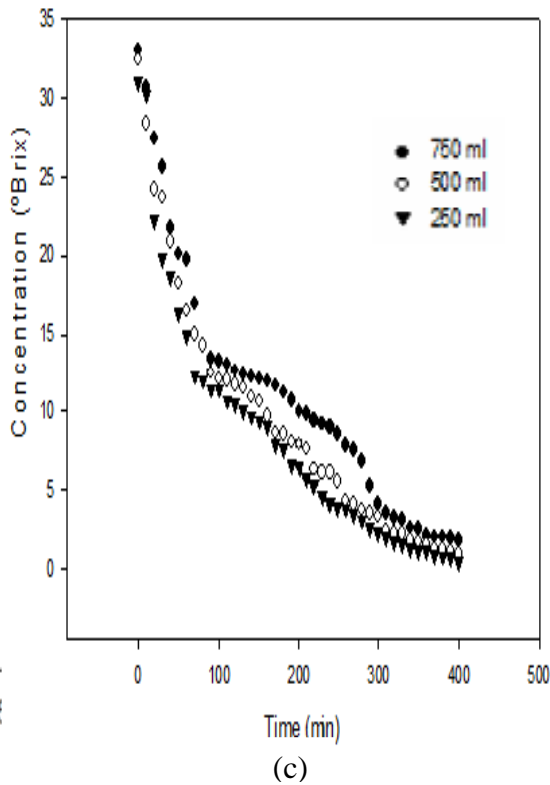


Figure 3.16. Variation of concentration with time for carrot concentrates at different sizes: (a) initial concentration 1 °Brix, (b) initial concentration 5 °Brix, (c) initial concentration 10 °Brix

Table 3.22. Regression analyses for carrot concentrates (initial concentration 1, 5 and °Brix) at different sizes

Initial Concentration (°Brix)	mL	Equation	R <sup>2</sup>
1	250	$y = 21.67 \cdot \exp(-0.009 \cdot x)$	0.907
	500	$y = 22.29 \cdot \exp(-0.009 \cdot x)$	0.950
	750	$y = 22.62 \cdot \exp(-0.009 \cdot x)$	0.918
5	250	$y = 26.32 \cdot \exp(-0.009 \cdot x)$	0.918
	500	$y = 26.90 \cdot \exp(-0.009 \cdot x)$	0.913
	750	$y = 27.23 \cdot \exp(-0.008 \cdot x)$	0.972
10	250	$y = 27.97 \cdot \exp(-0.008 \cdot x)$	0.965
	500	$y = 29.05 \cdot \exp(-0.007 \cdot x)$	0.978
	750	$y = 29.78 \cdot \exp(-0.006 \cdot x)$	0.953

\* x= Time (min)

\* y= Concentration (°Brix)

Equation coefficient are shown in Table 3.22. Equation coefficient increases, increasing sample size. According to regression analyses, sample size affects the degree of separation.

### 3.9. Effect of Time on the Concentration

According to data, generally concentration (°Brix) decreased at 10–15 minutes instantaneously for lower concentrated juices. After these times, concentration decreased slowly. Figures (3.17–3.21) show the variation of concentration with temperature for 10 °Brix grape concentrate at different time intervals.

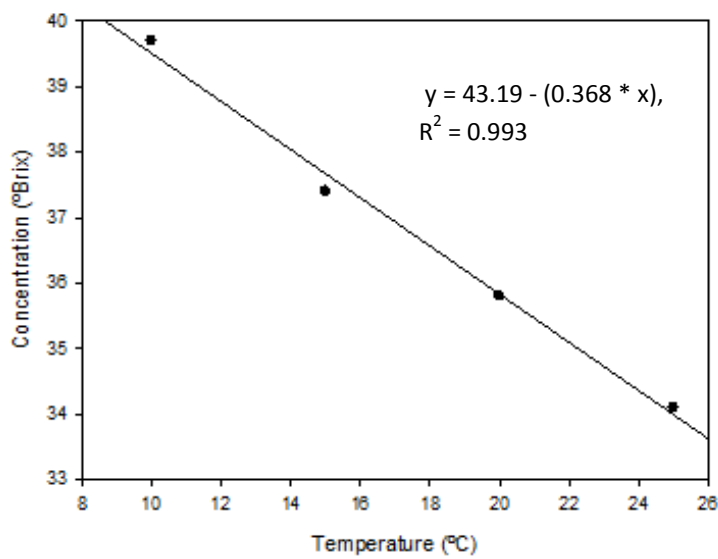


Figure 3.17. Variation of concentration versus temperature for 10 °Brix grape concentrate at 0 min

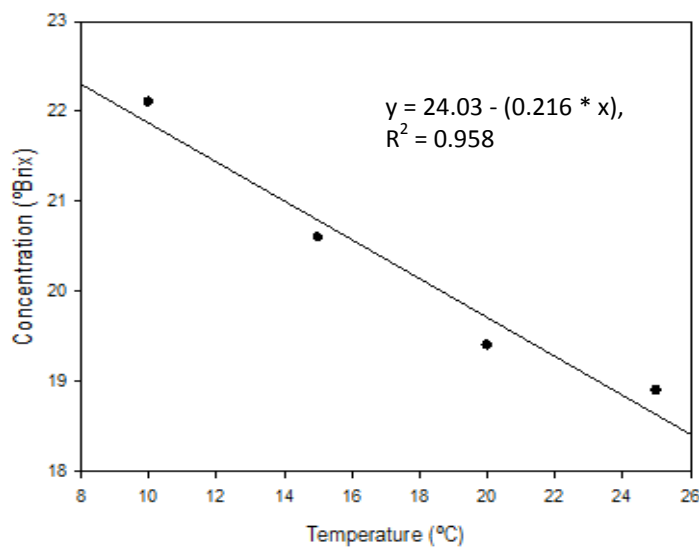


Figure 3.18. Variation of concentration versus temperature for 10 Brix grape concentrate at 25 min

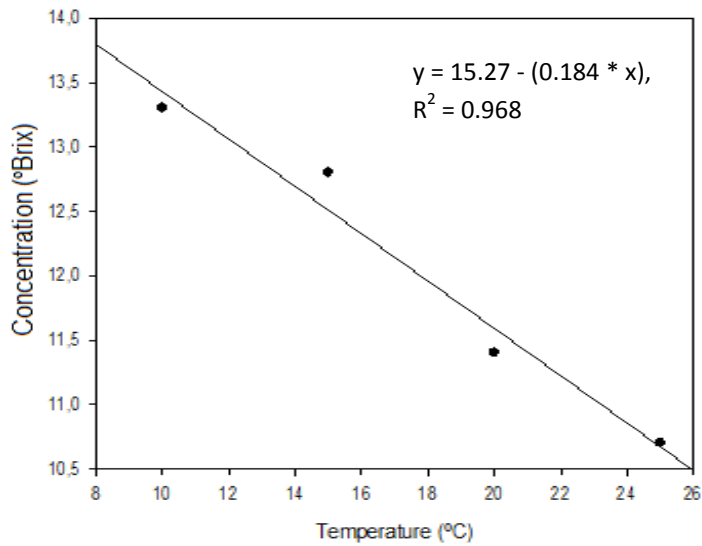


Figure 3.19. Variation of concentration versus temperature for 10 °Brix grape concentrate at 50 min

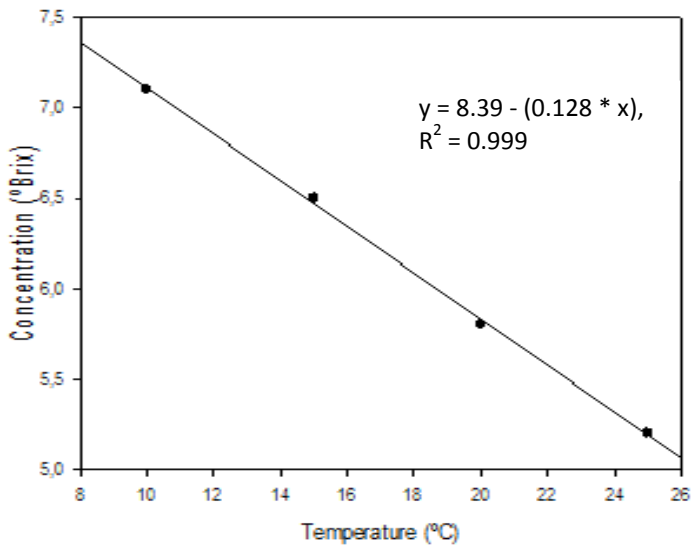


Figure 3.20. Variation of concentration versus temperature for 10 °Brix grape concentrate at 75 min

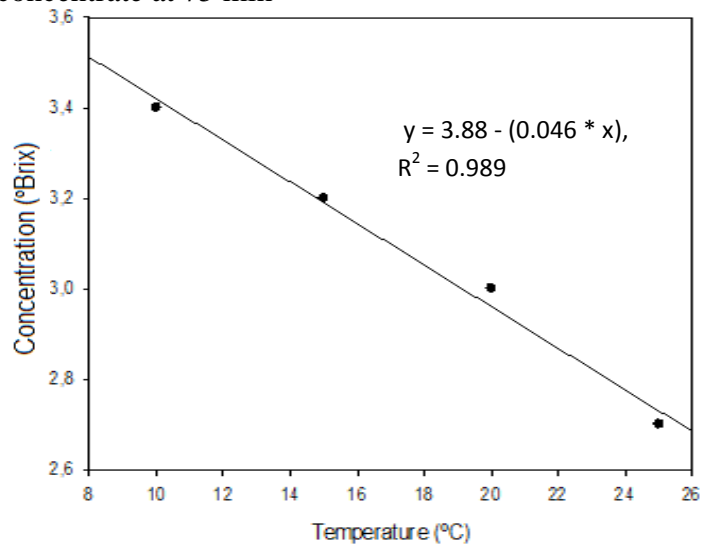


Figure 3.21. Variation of concentration versus temperature for 10 °Brix grape concentrate at 100 min

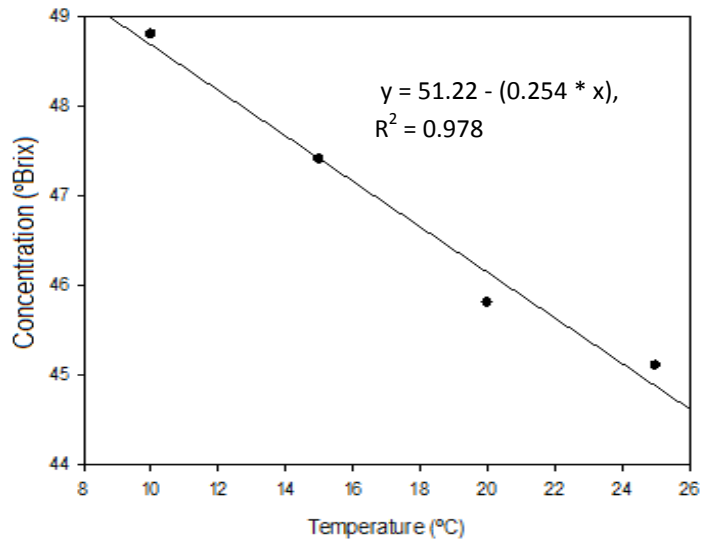


Figure 3.22. Variation of concentration versus temperature for 40 °Brix grape concentrate at 0 min

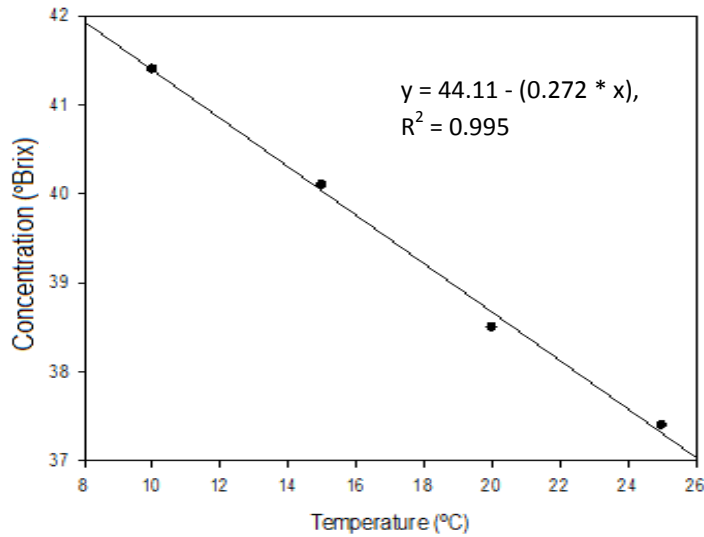


Figure 3.23. Variation of concentration versus temperature for 40 °Brix grape concentrate at 15 min

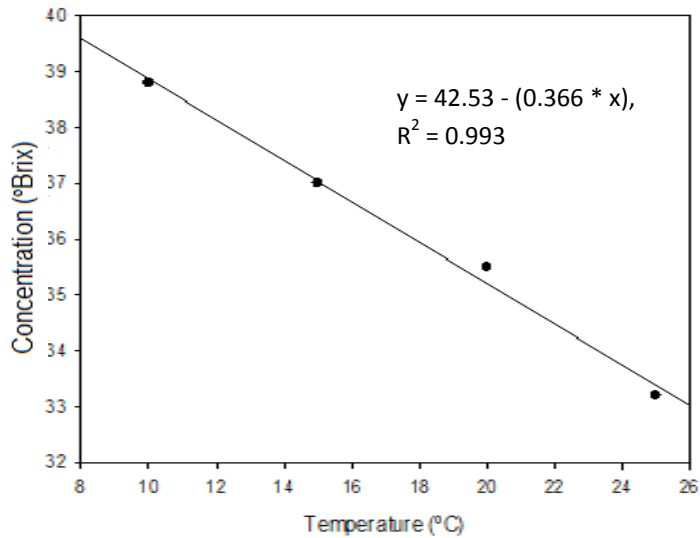


Figure 3.24. Variation of concentration versus temperature for 40 °Brix grape concentrate at 30 min

The separation of higher concentrated juices was slowly at each time interval. Figures 3.22–3.24 illustrate the change of concentration with temperature for 40 °Brix grape concentrate at different time intervals.

Figures (3.25–3.30) show the variation of concentration with temperature for 10 °Brix carrot concentrate at different time intervals. These plots show that freeze concentration is suitable to lower concentrated juices.

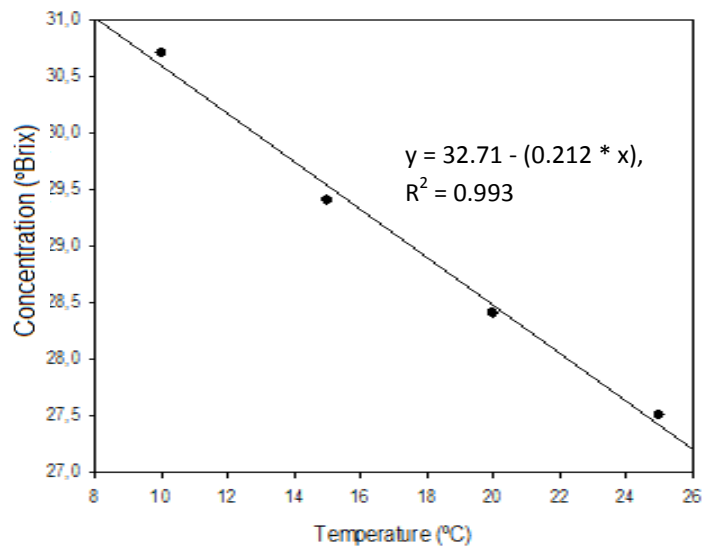


Figure 3.25. Variation of concentration versus temperature for 10 °Brix carrot concentrate at 0 min

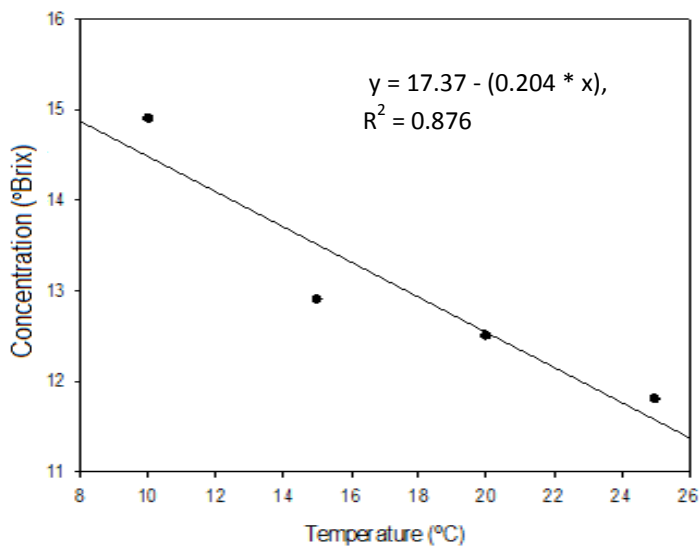


Figure 3.26. Variation of concentration versus temperature for 10 °Brix carrot concentrate at 25 min



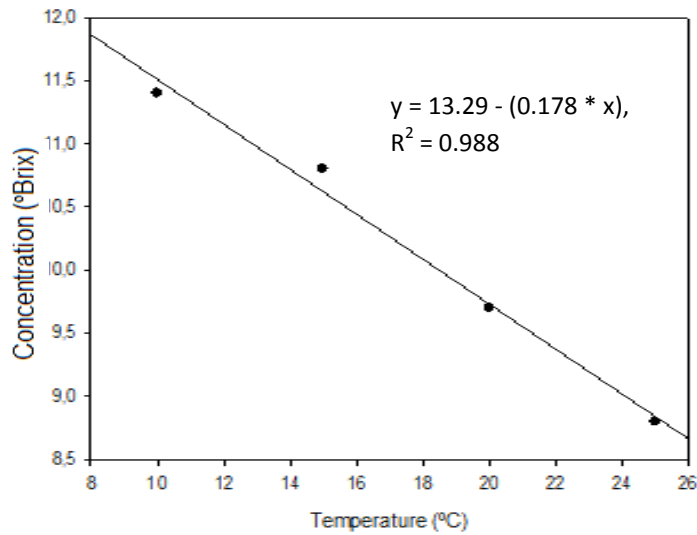


Figure 3.27. Variation of concentration versus temperature for 10 °Brix carrot concentrate at 50 min

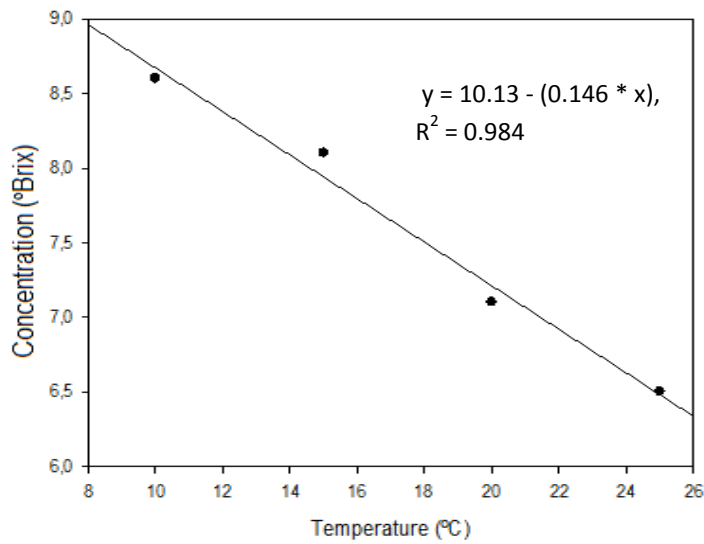


Figure 3.28. Variation of concentration versus temperature for 10 °Brix carrot concentrate at 75 min

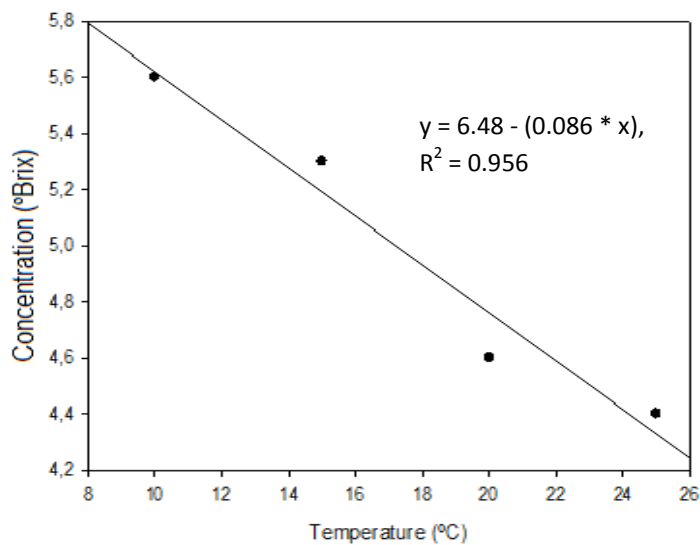


Figure 3.29. Variation of concentration versus temperature for 10 °Brix carrot concentrate at 100 min

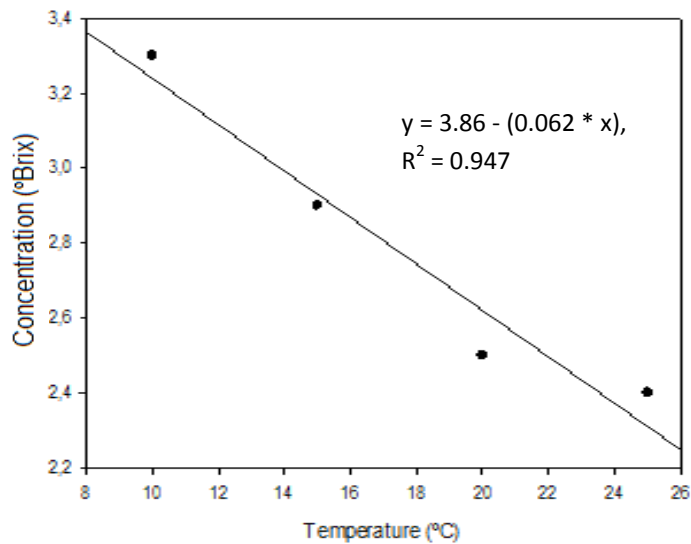


Figure 3.30. Variation of concentration versus temperature for 10 °Brix carrot concentrate at 125 min

The separation of higher concentrated juices was slowly at each time interval. Figures 3.31–3.33 illustrate the change of concentration with temperature for 40 °Brix carrot concentrate at different time intervals. The degree of separation is very low for higher concentrated carrot juices. Therefore, the degree of separation was slower at each time interval for higher concentrated juices.

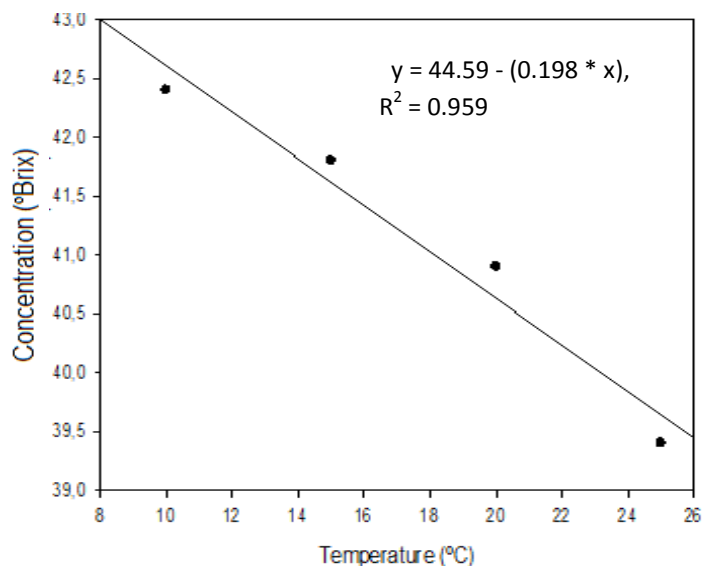


Figure 3.31. Variation of concentration versus temperature for 40 °Brix carrot concentrate at 0 min

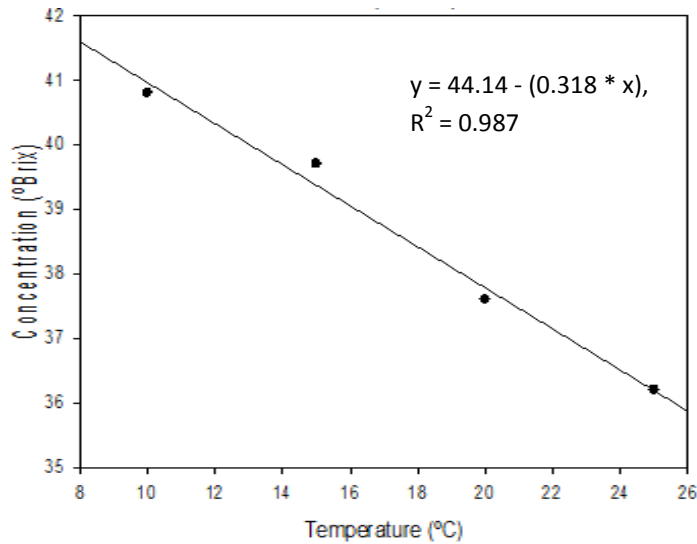


Figure 3.32. Variation of concentration versus temperature for 40 °Brix carrot concentrate at 15 min

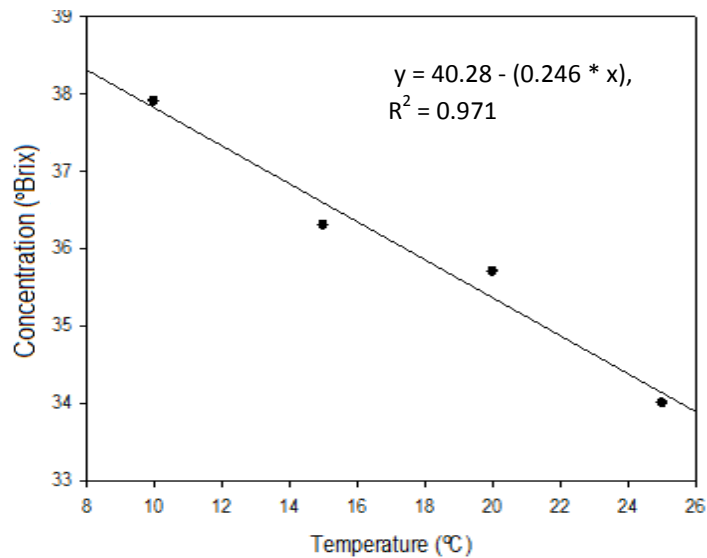


Figure 3.33. Variation of concentration versus temperature for 40 °Brix carrot concentrate at 30 min

Although the degree of separation is improved with the size of the samples, the time of separation, increases in parallel. This necessitates optimizing the size of the samples to achieve the highest degree of separation in shortest time. The degree of separation did not change appreciably with sample size in the first 60-90 minutes, after this time degree of separation was observed to be different for each size. After 60-90 minutes, the separation degree was decreased slowly.

### 3.10. Effect of Temperature

The influence of temperature on the rheological parameters of fruit juices have been studied in many articles. Temperature influence on viscosity has been found to be

related to soluble solids content, and experimental data for fixed concentrations can be related to the temperature using the Arrhenius-Guzman equation. Therefore, the variation in concentration with temperature on the flow behavior of fluid foods can be described by the exponential type Arrhenius relationship.

$E_a$  is defined as the energy that must be overcome in order for a chemical reaction to occur. Activation energy may also be defined as the minimum energy required starting a chemical reaction.

The Arrhenius equation gives the quantitative basis of the relationship between the activation energy and the rate at which a reaction proceeds.

The measured concentration results were used to calculate the values of parameter (flow activation energy  $E_a$ ) in Arrhenius equation. Figure 3.34 shows that these results are suitable to first order reaction. This figure illustrates first order reaction for 35 °Brix carrot concentrate. First order reaction was observed to be the best fitted model for all data. From the Arrhenius equation, the activation energy can be expressed as:

$$k = A e^{-E_a/RT} \quad (1)$$

where  $A$  is pre-exponential factor,  $R$  is the universal gas constant (8.314 J/mol K),  $T$  is the temperature (in Kelvin) and  $k$  is the reaction rate coefficient. This equation suggests that the activation energy is dependent on temperature.  $E_a$  (kJ/mol K) can be evaluated from the reaction rate coefficient at any temperature.

Take the natural log of both side in equation (1):

$$\ln k = \ln A - E_a/RT \quad (2)$$

Equation (2) has the form of a straight line as in Figure 3.35. It shows that temperature effect on concentration of 20 °Brix sumac concentrate. The slope of this graph gives  $-E_a/R$ .

A linear plot is obtained between rate constant and inverse of temperature. The slope of the straight line gives the activation energy.

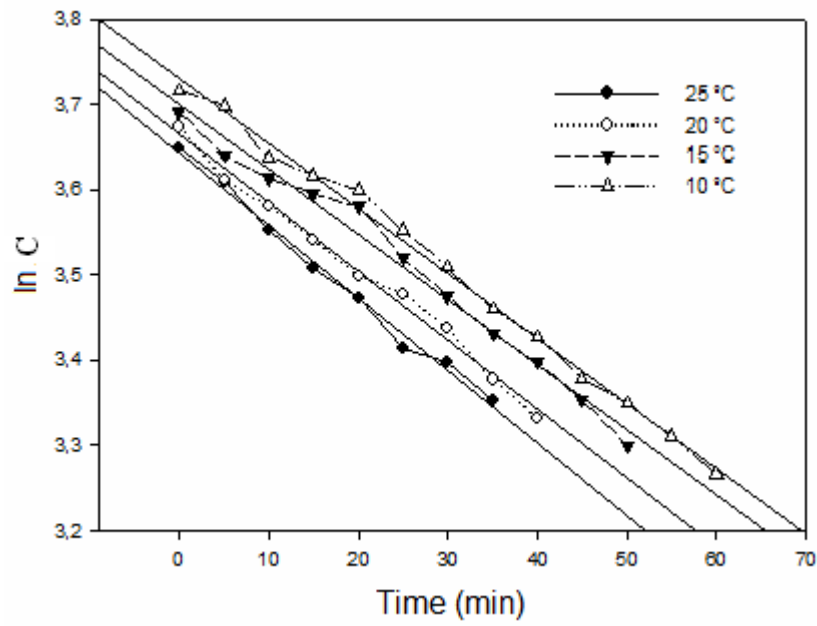


Figure 3.34. First order reaction for 35 °Brix carrot concentrate

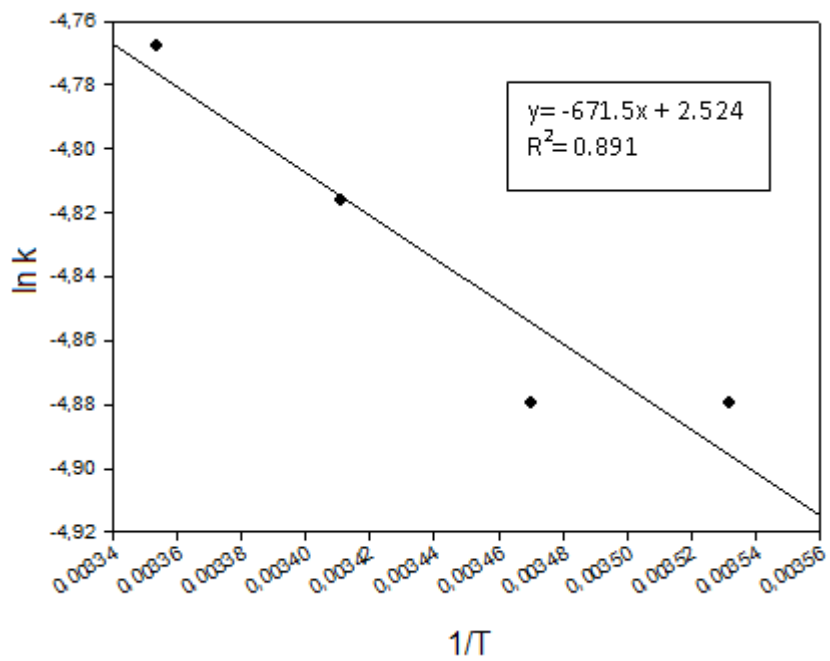


Figure 3.35. Arrhenius plot for 35 °Brix carrot concentrate

Table 3.23. Activation energy for different concentrated juices

<b>Fruit Juices</b>	<b>Initial Concentration (°Brix)</b>	<b>Activation Energy (<math>E_a</math>) (kJ/mol K)</b>	<b>R<sup>2</sup></b>
<b>Sumac</b>	10	5.57	0.699
	15	5.54	0.922
	20	4.46	0.892
	25	5.23	0.884
	30	6.09	0.826
	35	1.23	0.882
	40	1.93	0.824
	<b>Apple</b>	10	6.81
15		6.94	0.982
20		3.28	0.916
25		2.89	0.892
30		2.79	0.941
35		4.61	0.656
40		5.71	0.895
<b>Grape</b>		10	1.85
	15	3.32	0.921
	20	3.44	0.963
	25	3.25	0.893
	30	1.67	0.894
	35	5.05	0.896
	40	6.37	0.996
	<b>Carrot</b>	10	5.62
15		4.18	0.934
20		2.26	0.947
25		3.97	0.993
30		6.73	0.983
35		5.59	0.963
40		5.40	0.952

Table 3.23 shows  $E_a$  values for sumac, apple, grape, and carrot concentrates. The value of  $E_a$  did not change significantly for concentrated juices. However,  $E_a$  values changed significantly for 35 and 40 °Brix grape concentrate.  $E_a$  values decreased for 35 and 40 °Brix sumac concentrate. According to Wang (2007), the general tendency is for the activation energy to increase with the soluble solids concentration. This observation indicates that the effect of temperature on concentration of fruit juice concentrates is not pronounced for this study.

## CHAPTER 4

### CONCLUSION

In this study, the application of a modified version of freeze concentration, called differential freeze concentration, was applied to concentrate sumac, apple, grapes, and carrot juices.

The outcomes of this work can be summarized as follows:

1. When all juices were thawed, the more concentrated portion was always observed to separate from the ice front at the beginning of thawing. The concentration of the juice leaving the ice phase was observed to decrease with time.
2. The concentration of the juice leaving the ice phase was seen to be strongly dependent on the initial concentration of the frozen juice. The lower the initial concentration of the juice, the higher degree of separation was achieved. This method is, therefore, not feasible for samples having an initial concentration higher than 20 °Brix.
3. The more concentrated region of the frozen samples was visually observed to be near their centers. This is in agreement with the fact that, mass transfer occurs from high to low concentration regions.
4. The decay exponential model fitted best to the data obtained from the differential freeze concentration experiments for all juices.
5. The size of samples being thawed was observed to affect the degree of separation. The larger the size of samples, the higher the degree of separation realized. This is due to the decrease in  $(\text{surface area}/\text{volume})_{\text{SAMPLES}}$  as the size increases, which causes a relatively less heat transfer and consequently slower rate of thawing, resulting in a more concentrated juice leaving the ice front.
6. Although the degree of separation is improved with the size of the samples, the time of separation, increases in parallel. This necessitates optimizing the size of the



samples to achieve the highest degree of separation in shortest time.

7. The degree of separation was observed to be influenced also by the nature of the juice being concentrated. In general, more acidic samples resulted in a higher degree of separation. Thus the less acidic carrot juice exhibited lower degree of separation than that of sumac juice.

#### **4.1. Suggestion for Future Work**

Freeze concentration presents new opportunities in a wide variety of applications. Freeze concentration can be supported with the other methods to get higher concentrated samples. However, it has some problems like cost which prevent commercial application. The technique is easier and cheaper to use and adapt to small scale batch processes. A mathematical model, from which the transport properties of the present systems are determined, can be developed for the future work.

## REFERENCES

Aider, M., de Halleux, D., and Akbache, A. (2007). Whey cryoconcentration and impact on its composition. *Journal of Food Engineering*, 82(1), 92–102.

Aider, M., de Halleux, D., and Melnikova, I. (2007). Skim milk whey cryoconcentration and impact on the composition of the concentrated and ice fractions. *Food and Bioprocess Technology*, 10.1007/s11947-007-0023-0.

Aider, M., and Halleux D. (2008). Passive and microwave-assisted thawing in maple sap cryoconcentration technology. *Journal of Food Engineering*, 85: 65–72.

Aider, M., and Halleux D. (2009). Cryoconcentration technology in the bio-food industry. *Food Science and Technology*, 42: 679–685.

Atwood, G. R. (1973). Studies in melt crystallization. In E. S. Perry, & C. J. Van Oss (Eds.), *Separation and purification methods* (pp. 297–369). New York: Marcel Dekker.

Braddock, R. J., and Marcy, J. E. (1985). Freeze concentration of pineapple juice. *Journal of Food Science*, 50, 1636–1939.

Burdo, O. G., Kovalenko, E. A., and Kharenko, D. A. (2007). Development of freeze concentration device. *Communication of the Odessa state academy of food technologies*, Odessa, Ukraine.

Canovas, B., Tapia G. V, Cano, M. S., Martí'n-Belloso, M. P., and Martí'nez, A. (2005). *Novel food processing technologies* (1st ed.). CRC Press, Technology & Industrial Arts, p. 720.

Cassano, A., Conidi, C., Timpone, R., D'Avella, M., and Drioli, E. (2007). A membrane-based process for the clarification and the concentration of the cactus pear juice. *Journal of Food Engineering* 80: 914–921.

Conrad, K.M., Mast, M.G., and Ball, H.R. (1993). Concentration of liquid egg white by vacuum evaporation and reverse osmosis. *Journal of Food Science*, 58: 1017–1020.

Deshpande, S. S., Bolin, H. R., and Salunkhe, D. K. (1982). Freeze concentration of fruit juices. *Food Technology* (May), 68–82.

Elaya, M.M.O., and Gunasekaran, S. (2002). Gelling properties of egg white produced using a conventional and a low-shear reverse osmosis process. *Journal of Food Science*, 67: 725–729.

Englezos, P. (1992). The Freeze Concentration Process and its Applications.

Flesland, O. (1995). Freeze concentration by layer crystallization. *Drying Technology*, 13, 1713–1739.

Gea Process Engineering. <http://www.gea-messo-pt.com>.

Gilbert, S. W. (1991). Melt crystallization: process analysis and optimization. *AIChE Journal*, 37, 1205–1218.

Gu, X., Watanabe, M., Suzuki, T., and Miyawaki, O. (2008). Limiting Partition Coefficient in a Tubular Ice System for Progressive Freeze Concentration. *Food Science and Technology*, Vol. 14 (2008), No. 3 pp. 249-252.

Hartel, R. W., and Espinel, L. A. (1993). Freeze concentration of skim milk. *Journal of Food Engineering*, 20, 101–120.

Heist, J. A. (1979). Freeze concentration. *Chemistry Engineering*, 86(10), 72–82.

Heldman R. D., and Lund B. D. (2007). Handbook of Food Engineering, Second Edition. *Food Science and Technology* ( Taylor and Francis); 161.

Huige, N. J., and Thijssen, H. A. C. (1972). Production of large crystals by continuous ripening in a stirred tank. *Journal of Crystal Growth*, 13/14, 483–487.

Jusoh, M., Yunus, R.M., and Abu Hassan, M.A. (2009). Performance Investigation on a New Design for Progressive Freeze Concentration System. *Journal of Applied Sciences* 9 (17): 3171-3175.

Karel, M. (1975). Membrane Separation Processes and Freeze Concentration in the 1980s [and Discussion]. *Great Britain Proc R. Soc. Lond. B.* 191, 21-48.

Keshani, S., Luqman Chuah, A., Nourouzi, M. M., Russly, A.R., and Jamilah, B. (2010). Optimization of concentration process on pomelo fruit juice using response surface methodology (RSM). *International Food Research Journal*, 17: 733-742.

Kessler, H. G. (1986). In *Concentration and Drying of Foods* (Ed: D. MacCarthy), Elsevier Applied Science, Barking, Essex.

Kramer, A., Wani, K., Sullivan, J. H., and Shomer, I. (1971). Freeze concentration by directional cooling. *Journal of Food Science*, 36(2), 320–322.

Lee, Y.C., and Lee, S.W. (1999). Quality changes during storage in Korean Clear Pear Juices concentrated by three methods. *Journal of Food Quality* 22: 565-571.

Lemmer, S., Klomp R., Ruemekorf, R., and Scholz, R. (2000). Preconcentration of Wastewater through the Niro Freeze Concentration Process. *German in Chem. Ing. Tech.* 72: 403-405.

Matthews, J. S., and Coggeshall, N. D. (1959). Concentration of impurities from organic compounds by progressive freezing. *Analytical Chemistry*, 31, 1124–1125.

Matthews, O., William H., and Gael P. A. Comparison of Freeze Concentration in Celsius-Paks and a Conventional Freeze-Thaw System. *Sartorius Stedim Biotech, GmbH W/sart-000, G Publication No.: SL-1057-e07121 Order No.: 85032-534-39.*

Mazura, J., Rosli, M. Y., Mohd Ariffin, Abu Hassan. Development of a New Crystallisation Chamber for a Progressive Freeze Concentration System.

Mellor, J. D. (1978) *Fundamentals of Freeze-drying*. Academic Press, London, pp. 257–288.

Miyawaki, O., Abe, T., and Yano, T. (1992). Freezing and ice structure formed in protein gels. *Bioscience Biotechnology and Biochemistry*, 56, 953–957.

Miyawaki, O., Liu, L., and Nakamura, K. (1998). Effective partition constant of solute between ice and liquid phases in progressive freeze-concentration. *Journal of Food Science*, 63, 756–758.

Miyawaki, O., Liu, L., Shirai, Y., Sakashita, S., Kagitani, K. (2003). Tubular ice system for scale-up of progressive freeze-concentration. *Journal of Food Engineering*, 69 (2005) 107–113.

Miyawaki, O., Liu, L., Shirai, Y., and Kagitani, K. (2005). Tubular Ice System For Scale-up of Progressive Freeze concentration, *Journal of Food Engineering*, Vol (69), 107-113.

Miyawaki, O. (2009). Progressive Freeze-Concentration: A New Method for High Quality Concentration of Liquid Food. *Foods Food Ingredients*, Vol. 214, No.2, 2009.

- Muller, J. G. (1967) Freeze concentration of food liquids: theory, practice and economics. *Food Technology*, 21, 49–52, 54–56, 58, 60, 61.
- Muller, M., and Sekoulov, I. (1992). Waste water reuse by freeze concentration with a falling film reactor. *Water Science and Technology*, 26, 1475–1482.
- Myerson, A. S., and Ginde, R. (1993). Handbook of Industrial Crystallization. A. S. Myerson, Edition. Butterworth-Heinemann, Stoneham, pp. 60-61.
- Nebbia, G., and Menozzi, G.N. (1968). Early Experiments on Water Desalination by Freezing. *Desalination*, 5, 49-54.
- Parker, A. (1942). Potable Water from Sea-water. *Nature*, 149,184-186.
- Pederson Carl, S., and Beattie Harold, G. (1947). Concentration of Fruit Juices By Freezing. *New York State Agricultural Experiment Station*, Bulletin No. 727, Geneva, New York.
- Petzold G., and Aguilera J. M. (2009). Ice Morphology: Fundamentals and Technological Applications in Foods. *Food Biophysics*, 4:378–396.
- Pradistsuwana, C., Theprugsa, P., and Miyawaki, O. (2003). Measurement of Limiting Partition Coefficient in Progressive Freeze-Concentration. *Food Science Technology, Res.*, 9 (2), 190–192.
- Ramaswamy, H., and Marcotte, M. (2006a). Separation and Concentration. *In Food processing Principles and Applications*, p.317-379. United States of America: CRC Press.
- Ramteke, R.S., Singh, N.I., Rekhaand, M. N., and Eipeson, W. E. (1993). Methods for concentration of fruit juices: A critical evaluation. *Journal of Food Science and Technology*, 30: 391-402.
- Rane, M. V., and Jabade, S. (2005). Freeze concentration of sugarcane juice in a jaggery making process. *Applied Thermal Engineering*, 25, 2122–2137.
- Raventos, M., Hernandez, E., Auleda, J., and Ibarz, A. (2006). Concentration of Aqueous Sugar Solutions in a Multi- plate Cryoconcentrator. *Journal of Food Engineering*, Vol. 79, 577-585.
- Sanchez, J., Ruiz, Y., Auleda, J. M., Hernandez, E., and Raventos, M. (2009). Review. Freeze Concentration in the Fruit Juices Industry. *Food Science and Technolog*, 0(00):0001-0013.
- Sanchez, J., Hernandez, E., Auleda, J.M., Raventos, M., Dette S. S., and Jansen H. (2009). Freeze Concentration of Black Currant Juice. *Chemical Engineering Technology*, 33, No. 5, 762–766.

- Sanchez, J., Hernandez, E., Auleda, J.M., and Raventos, M. (2010). Freeze concentration of whey in a falling-film based pilot plant: Process and characterization. *Journal of Food Engineering*, 103 (2011) 147–155.
- Sanchez, J., Hernandez, E., Auleda, J.M., and Raventos, M. (2011). Review: Freeze Concentration Technology Applied to Dairy Products. *Food Science and Technology International*, 17: 5.
- Shamsudin, R. (2000). Design, concentration and Testing of freeze concentration, PhD Thesis, University Putra Malaysia.
- Shapiro, J. (1961). Freezing-out, a safe technique for concentration of dilute solutions. *Science*, 133, 2063–2064.
- Stepakoff, G. L., Siegelman, D., Johnson, R., and Gibson, W. (1974). Development of a eutectic freezing process for brine disposal. *Desalination*, 15(1), 25–38.
- Thijssen, H.A.C. (1975). Current developments in the freeze concentration of liquid foods. In Goldblith, S.A., Rey L. and Rothmayr, W. W. (Ed.), *Freeze Drying and Advanced Food Technology*, p. 481–501. New York: Academic Press.
- Vaessen, R. J. C., Himawan, C., and Witkamp, G. J. (2002). Scale formation of ice from electrolyte solutions on a scraped surface heat exchanger plate. *Journal of Crystal Growth*, 237–239, 2172–2177.
- Vaessen, R. J. C., Janse, B. J. H., Seckler, M. M., and Witkamp, G. J. (2003). Evaluation of the performance of a newly developed eutectic freeze crystallizer: scraped cooled wall crystallizer. *Chemical Engineering Research and Design*, 81(10), 1363– 1372.
- Vaessen, R. J. C., Seckler, M. M., and Witkamp, G. J. (2003). Eutectic freeze crystallization with an aqueous KNO<sub>3</sub>–HNO<sub>3</sub> solution in a 100-L cooled disc column crystallizer. *Industrial & Engineering Chemistry Research*, 42(20), 4874–4880.
- Van der Ham, F., Witkamp, G. J., De Graauw, J., and Van Rosmalen, G. M. (1998). Eutectic freeze crystallization: application to process streams and waste water purification. *Chemical Engineering and Processing*, 37, 207–213.
- Wakisaka, M., Shirai, Y. and Sakashita, S. (2001). Ice crystallization in a pilot-scale freeze wastewater treatment system. *Chemical Engineering and Processing*, 40, 201–208.
- Wang, W., and Xu S. (2007). Degradation kinetics of anthocyanins in blackberry juice and concentrate. *Journal of Food Engineering*, 82 (2007) 271–275.
- Wenzelberger, E. P. (1951). Apparatus for the Dehydration of Water Bearing Substances. *The Commonwealth Engineering*, United States Patent, 2552525.

Werezak, GN. (1969). Unusual methods of separation. *AIChE Symp Series*, 65(91). 6-18.

Widehem, P., and Cochet, N. (2003). *Pseudomonas syringae* as an ice nucleator-application to freeze- concentration. *Process. Biochem.* 39: 405-410.

Yuping, S., and Baomin, L. (2001). US Patent No. 6305178. Continuous system of freeze concentration for aqueous solutions.