T.C.

## YEDITEPE UNIVERSITY

## INSTITUTE OF HEALTH SCIENCES

## DEPARTMENT OF MOLECULAR MEDICINE

## INVESTIGATION OF CATECHOL-O-METHYLTRANSFERASE (COMT) GENE VAL158MET POLYMORPHISM IN OVARIAN CANCER

MASTER OF SCIENCES THESIS

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#### THESIS APPROVAL FORM

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#### APPROVAL

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I have gained all the information for this thesis within the bounds of academic and ethical rules, I have cited all the information and interpretations not obtained by this thesis, and I have listed these sources in the list of references again, I have pledge that this thesis work is my own work, from the planning of the thesis to the writing, I declare that there is no violation of the patent and copyrights during the study and writing of this thesis.

28.06.2019

İpek Yağmur ABAOĞLU

## DEDICATION

To my whole family

To my invaluable teacher Prof. Dr. Turgay İSBİR

To my friends who encourage and support me.

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## LIST OF SYMBOLS AND ABBREVIATIONS

BMI	:Body Mass Index
CI	:Confidence Interval
СТ	:Computed Tomography
COMT	:Catechol-O-Methyltransferase
CST®	:ChargeSwitch® technology
CYP1A1/1A2	:P 450 1A1, 1A2
CYP1B1	:Cytochrome P 4501B1
dsDNA	:Double-Stranded DNA
FIGO	:International Federation of Gynecology and
	Obstetrics
E1	:Estrone
E2	:Estradiol
E3	:Estriol
EDTA	:Ethylenediaminetetraacetic acid
GLOBACAN	:Global Cancer Observatory
GLOBACAN GSTP1	:Global Cancer Observatory :Glutathione-S-Transferase
GSTP1	:Glutathione-S-Transferase
GSTP1 MB	:Glutathione-S-Transferase :Membrane Bound
GSTP1 MB MB-COMT	:Glutathione-S-Transferase :Membrane Bound :Membrane-Bound Shape
GSTP1 MB MB-COMT MRI	:Glutathione-S-Transferase :Membrane Bound :Membrane-Bound Shape :Magnetic Resonance Imaging
GSTP1 MB MB-COMT MRI NAC	:Glutathione-S-Transferase :Membrane Bound :Membrane-Bound Shape :Magnetic Resonance Imaging :N-Acetylcysteine
GSTP1 MB MB-COMT MRI NAC NAT2	:Glutathione-S-Transferase :Membrane Bound :Membrane-Bound Shape :Magnetic Resonance Imaging :N-Acetylcysteine :N-acetyltransferase 2
GSTP1 MB MB-COMT MRI NAC NAT2 NEOC	:Glutathione-S-Transferase :Membrane Bound :Membrane-Bound Shape :Magnetic Resonance Imaging :N-Acetylcysteine :N-acetyltransferase 2 :Non-Epithelial Ovarian Cancers
GSTP1 MB MB-COMT MRI NAC NAT2 NEOC OR	:Glutathione-S-Transferase :Membrane Bound :Membrane-Bound Shape :Magnetic Resonance Imaging :N-Acetylcysteine :N-acetyltransferase 2 :Non-Epithelial Ovarian Cancers :Odd Ratio
GSTP1 MB MB-COMT MRI NAC NAT2 NEOC OR PET	<ul> <li>:Glutathione-S-Transferase</li> <li>:Membrane Bound</li> <li>:Membrane-Bound Shape</li> <li>:Magnetic Resonance Imaging</li> <li>:N-Acetylcysteine</li> <li>:N-acetyltransferase 2</li> <li>:Non-Epithelial Ovarian Cancers</li> <li>:Odd Ratio</li> <li>:Positron Emission Tomography</li> </ul>
GSTP1 MB MB-COMT MRI NAC NAT2 NEOC OR OR PET PARPs	<ul> <li>:Glutathione-S-Transferase</li> <li>:Membrane Bound</li> <li>:Membrane-Bound Shape</li> <li>:Magnetic Resonance Imaging</li> <li>:N-Acetylcysteine</li> <li>:N-acetyltransferase 2</li> <li>:Non-Epithelial Ovarian Cancers</li> <li>:Odd Ratio</li> <li>:Positron Emission Tomography</li> <li>:Poly (ADP-ribose) polymerases</li> </ul>

SAH	:S-adenosyl-L-homocysteine
S-COMT	:Soluble Shape
SNPs	:Single Nucleotide Polymorphisms
ssDNA	:Single-Stranded DNA
SPSS	:Statistical Package for Social Science
WHO	:World Health Organization
2/4-MeOE1/E2	:2/4-methoxy-estrone/estradiol



#### ABSTRACT

## Abaoğlu İ.Y. (2019). Investigation of Catechol-O-Methyltransferase (COMT) Gene Val158Met Polymorphism in Ovarian Cancer. Yeditepe University, Health Sciences Institude, Department of Molecular Medicine. Master Thesis. İstanbul.

Ovarian cancer is the seventh leading cause of death and is the eighth most common cancer among women. It has the highest mortality rate among all gynecological cancers. The prognosis of ovarian cancer is poor, especially when the disease is diagnosed at an advanced stage. The COMT enzyme synthesized from the Catechol-O-Methyltransferase (COMT) gene detoxifies the carcinogenic catechol estrogens. This enzyme catalyzes the methoxylation reaction to produce 2-methoxy estradiol. This metabolite induces apoptosis and suppresses carcinogenesis. In the COMT gene, the Val158Met polymorphism results in a decrease in the activity of this enzyme, resulting in the accumulation of carcinogenic catechol estrogens. This thesis study, it is aimed at examining the relationship of the disease with COMT Val158Met polymorphism which is thought to affect the risk of ovarian cancer. The two groups were seperated as a patient group with ovarian cancer (n = 47) and control group (n =47). Genotyping of both groups was determined by Real-Time PCR, and the statistical analysis of the data was performed by SPSS program. According to our results, GG genotype (homozygote wild type) was found in 12 (25.5%), GA genotype (heterozygote type) in 22 (46.8%), AA genotype (homozygote variant type) in 13 (27.7%) in the control group statistically. In the patient group, genotype distributions were determined 17 (37.8%), 19 (42.2%) and 9 (20%) respectively. There was no significant relationship in comparison with genotypes between patient and control groups (p=0.413). This study could provide a novel approach for the clinical treatment of ovarian cancer.

**Key words:** Polymorphism, Catechol-O-Methyltransferase, Catechol Estrogen, Ovarian Cancer.

#### **ABSTRACT** (Turkish)

## Abaoğlu İ.Y. (2019). Yumurtalık Kanserinde Katekol-O-Metiltransferaz (COMT) Gen Val158Met Polimorfizminin Araştırılması. Yeditepe Üniversitesi, Sağlık Bilimleri Enstitüsü, Moleküler Tıp Anabilim Dalı. Yüksek Lisans Tezi. İstanbul.

Yumurtalık kanseri dünyada yedinci önde gelen ölüm nedenidir ve kadınlar arasında en yaygın sekizinci kanserdir. Bütün jinekolojik kanserler arasında en yüksek ölüm oranına sahiptir. Yumurtalık kanseri olan hastalar için özellikle de hastalık ileri safhada teşhis edildiğinde prognozu kötüdür. Katekol-O-Metiltransferaz geninden sentezlenen COMT enzimi karsinojenik olan katekol östrojenlerini detoksifiye eder. Bu enzim metoksilasyon reaksiyonunu katalizleyerek 2-metoksi östradiol üretir. Bu metabolit, apoptozisi uyararak karsinojenezi baskılar. COMT geninde Val158Met polimorfizmi bu enzimin aktivitesinin azalmasına yol açarak karsinojenik katekol östrojenlerinin birikimine neden olur. Bu tez çalışmasında, yumurtalık kanseri riskini etkilediği düşünülen COMT Val158Met polimorfizmi ile hastalığın ilişkisinin incelenmesi amaçlanmıştır. Yumurtalık kanseri olan hasta grubu (n=47) ile kontrol grubu (n=47) olmak üzere iki gruba ayrıldı. Her iki grubun genotiplemesi gerçek zamanlı PZR ile belirlenmiş ve verilerin istatistiksel analizi SPSS programı kullanılarak gerçekleştirildi. Sonuçlarımıza göre, kontrol grubunda GG genotipi (homozigot yabani tip) 12 (% 25.5), GA genotipi (heterozigot tip) 22 (% 46.8), AA genotipi (homozigot varyant tip) 13 (% 27.7) bulundu. Hasta grubunda genotip dağılımı sırasıyla 17 (% 37.8), 19 (% 42.2) ve 9 (% 20) olarak tespit edildi. Hasta ve kontrol grubu arasında genotiplerle karşılaştırıldığında anlamlı bir ilişki bulunmadı (p = 0.413). Bu çalışma, yumurtalık kanserinin klinik tedavisi için yeni bir yaklaşım sağlayabilir.

Anahtar Kelimeler: Polimorfizm, Katekol-O-Metiltransferaz, Katekol Östrojeni, Yumurtalık Kanseri.

#### **1. INTRODUCTION and PURPOSE**

In clinical medicine, cancer is a multifactorial disease that is one of the very widespread diseases in the world and the main cause of morbidity globally (1,2).

The most frequent type of female malignancies is ovarian cancer which has the highest death rate and causes many new cases each year (3,4). Global Cancer Observatory (GLOBACAN) stated that there were 239.000 cancer cases which resulted in 152.000 deaths with ovarian cancer. Ovarian cancer is the seventh most frequent type of cancer (3.6 % of cases), also it is the eighth (4.3 %) in terms of cancer-related deaths in women.

In developed countries, the rate of occurence is highest, at more than 7.5 per 100.000. The risk of ovarian cancer mortality in people under 75 years old is two times more in developed countries than underdeveloped countries (5). According to findings by World Health Organization (WHO) in Turkey, there were 3.729 incidences and 2.191 mortality cases in 2018. Furthermore, it's expected that incidence (3.939 cases) and mortality cases (2.336) will increase among females in 2020 (6).

There are many risk factors which change the genetic predisposition to ovarian cancer, including consumption of alcohol, obesity, aging and hereditary ovarian cancer. In addition, the oxidative stress, inflammation, angiogenesis, and apoptosis could alter the progression of carcinoma. Thus, controlling these factors play a crucial role in cancer prevention (7,8).

The Catechol-O-Methyltransferase (COMT) gene is located on chromosome 22, which consists of six exons and its protein is synthesised as two different types called membrane-bound (MB) and cytosolic (S) forms. The molecular weight of the MB-COMT protein is 30kd, whereas S-COMT protein is 24kd (9). There is a large quantity of COMT in different human tissues such as liver, breast, ovarian and kidney tissues (10). COMT is a significant functional enzyme which metabolizes estrogens, and it is included in the conjugating and deactivation of estrogen catechol metabolite (11). The functional polymorphism in the COMT gene occurs with the base pair change from G to A.This polymorphism called Val158Met (rs4680) results in the replacement of the

valine with methionine (12). It is assumed that this exchange plays an important role in leading to three to four times reduction in COMT activity (13,14).

It has been shown that COMT is associated with the risk of hormonally affected cancers (12,15). Consequentially, the altered COMT gene expression with inactivating catechol estrogens has reduced via generating mediate products catechol estrogen metabolism (15,16). Owing to the production and accumulation of catechol estrogens that lead to have oncogenic activity, the people with COMT-Met polymorphism could have higher risk of ovarian cancer (17).

The objective of this study is to determine to possible relation between Val158Met polymorphism of COMT gene and ovarian cancer in a Turkish population. The COMT gene Val158Met polymorphism was analyzed by using the Real-Time Polymerase Chain Reaction method and we investigated genotypic and allelic differences between the healthy controls and patients with ovarian cancer. Moreover, our results could present scientific data for population variations as a factor. At the same time, this study could provide a novel approach for the clinical treatment of ovarian cancer.

#### **2. LITERATURE REVIEW**

#### 2.1. Cancer

Cancer is considered a genetic disease because of its relation to the genetic material of the cell. The hereditary or environmental factors may cause cancer. The basic mechanism of carcinogenesis is a non-lethal genetic damage in the cell. These damages are caused by inactivation and activation in genes that activate or suppress cell proliferation respectively, it occurs as a result of the change in DNA repair genes associated with the programmed death of the cell (apoptosis) (18).

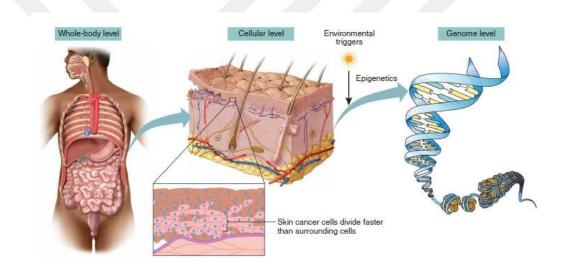


Figure 2.1. The representation of cancer levels (19).

Cancer can be seen various at levels, such as whole-body level, cellular level and genome level in the body (Figure 2.1.) (19).

Through the statistical analysis it is shown that the incidence of all kinds of cancers is 210.2 per hundred thousand between the years of 2010-2014, according to the standardized age and for both sexes in Turkey. The incidence at the standardized age is 173.6 per hundred thousand for females, and 246.8 per hundred thousand for males in 2014. In 2014, 163,417 individuals were diagnosed with carcinoma in Turkey. As the data of the last five years displayed, there is no increase or decrease in the frequency of cancer in both males and females (20).

On the one hand, it is estimated that there are 1,762,450 new cancer cases and 606,880 cancer deaths according to the American Cancer Society' studies in the United States in 2019 (21).

#### 2.2. Gynaecological Cancers

The three most prevalent gynecological malignancies are ovarian, cervical and uterine cancer. Gynecologic malignancies are commonly fatal in the current gynecologic cancer's treatment. The treatment options have provided extensions women's in life expectancy. New therapy strategies have accelerated survival rates due to novel molecular therapies (22).

In 2018, it was estimated that 110,070 women were diagnosed with gynecological cancer in the United States and this resulted 32.120 deaths. Molecular and genetic bases of the ovarian cancer which is the main subject of our study, accounts as the fifth most common cause of cancer-related death among women in the United States, while uterine cancer is the 6th most common cause of cancer-related deaths in women (23).

#### 2.3. Structure, Histological and Physiological Characteristics of Ovaries

The ovary-originated various cancer types are better clarified by understanding embryogenic development and the anatomy of the ovary (24). In the female, the pelvic reproductive organs are ovaries which produce sex hormones such as progesterone and estrogens (25). It is ovoid shaped and there are two ovaries located on both sides of the uterus (26). According to the examinations ovarian length is on average 13 mm, thickness is 4 mm, width is 5.7 mm, and volume is 125.88 mm<sup>3</sup> (27).

Structure of ovaries is included two types of components: cortex located outer and medulla located inner. In the connective tissue embedded part of the cortex is the follicles, that contain the female gametes called oocytes (28), and the cortex includes interstitial cells and corpora lutea (29). As for medulla, it is made up loose connective tissue and contains blood vein, lymphatic vessels, spiral arteries, hilus cells and neural network (Figure 2.2.) (25,29).

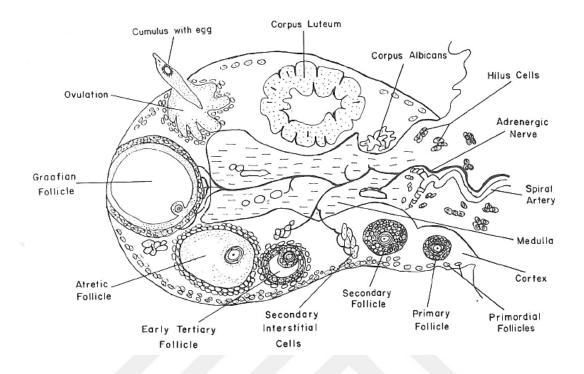


Figure 2.2. The scheme of the human ovary during years of reproduction (29).

#### 2.4. Ovarian Cancer

Ovarian cancer is the most common type among gynecological cancers and accounts for the highest rate of deaths related to gynecological cancers (30,31).

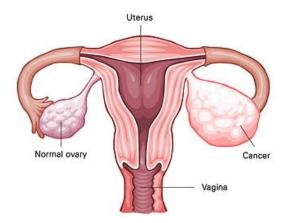


Figure 2.3. The structure of the normal and cancerous ovary (32).

Cancerous tumors stem from one non-normal cell, that cell causes the cancer disease, and spread all ovary (Figure 2.3.) (32).

#### 2.4.1. Incidence and Epidemiology of Ovarian Cancer

Approximately 23,000 new cases are diagnosed with the ovarian cancer in the worldwide every year. The deaths caused by ovarian cancer are estimated to be around 14.000 annually. Incidences of ovarian cancer have been reduced by 1% per year since the mid 1970s (33).

The occurrence of ovarian cancer varies by geographical area and age (35). Low rates of ovarian cancer have been determined in Africa and Asia while the highest rates have been revealed in the USA, Northern and Western Europe, but Japan has the lowest rate compared with other developed countries (36).

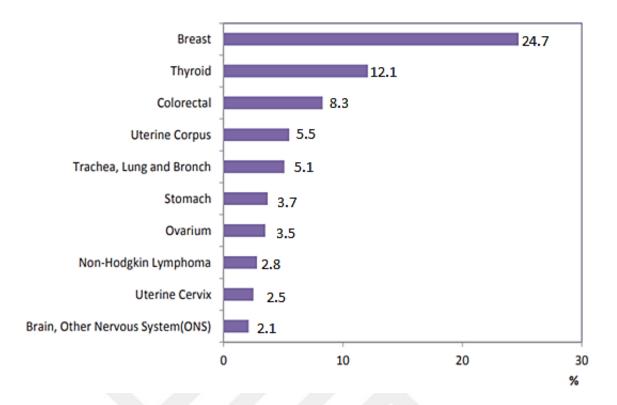
Five year survival rates are viewed in Table 2.1.,and almost 22,530 new cases of ovarian cancer are determined by the National Cancer Institute in 2019 (34,37).

**Table 2.1.** The comparison of various cancers diagnosed by National Cancer Institute

 between 2008 and 2014 (37).

	All stages	Local	Regional	Distant		All stages	Local	Regional	Distant
Breast (female)	90	99	85	27	Oral cavity & pharynx	65	84	65	39
Colon & rectum	65	90	71	14	Ovary	47	92	75	29
Colon	64	90	71	14	Pancreas	9	34	12	3
Rectum	67	89	70	15	Prostate	98	>99	>99	30
Esophagus	19	45	24	5	Stomach	31	68	31	5
Kidney†	75	93	69	12	Testis	95	99	96	74
Larynx	61	78	46	34	Thyroid	98	>99	98	56
Liver‡	18	31	11	2	Urinary bladder§	77	69	35	5
Lung & bronchus	19	56	30	5	Uterine cervix	66	92	56	17
Melanoma of the skin	92	98	64	23	Uterine corpus	81	95	69	16

According to the findings of the Republic of Turkey's Ministry of Health, there were approximately 2,361 women diagnosed with ovarian cancer in 2016 (38).



**Figure 2.4.** The percentage (%) distribution of the most common female cancer types in overall cancers, in 2015 (38).

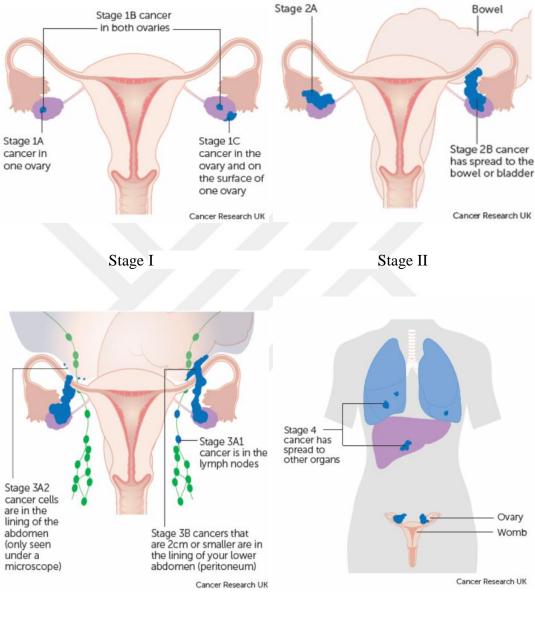
Figure 2.4. shows that the percentage of ovarian cancer is at 3.5 % in overall cancer among females during 2015 (38).

#### 2.4.2. The Stages of Ovarian Cancer

The clinical condition of malign tumors can be estimated by staging and classification of cancer and so that the appropriate treatment can be determined (39).

In 2014, the International Federation of Gynecology and Obstetrics (FIGO) reorganized the system for doctors to use for the staging of ovarian cancer. In the FIGO system, ovarian cancer consists of four stages, called Stage I, Stage II, Stage III and Stage IV, the first three stages have the sub-stages a, b and c. The stages are determined by whether or not cancer cells spread around the ovaries as well as the size of the cancer. In stage I, cancer cells are seen only in the ovaries and their spread is slow. In stage II and stage III, cells spread out of the ovaries. In stage IV, which is the last stage, cells are seen in the liver or lungs (Figure 2.5.) (40).

70% of the patients are diagnosed during the advanced stages, such as stage III and IV because ovarian cancer does not show any long-term symptoms especially in the early stages of the disease (36).



Stage III

Stage IV

Figure 2.5. The stages of ovarian cancer according to FIGO (40).

#### 2.4.3. The Classification of Ovarian Cancer

The ovarian tumors are categorized according to the different morphological and genetic characteristics. Ovarian cancers are divided into two groups: epithelial and non-epithelial cancers. Non-epithelial ovarian tumors are rare and define as germ cell tumors, sex cord-stromal cell tumours. Non-epithelial ovarian cancers (NEOC) are seen in all age groups, which formed part of 10-15% of ovarian cancers (41).

In addition, the epithelial ovarian tumors were reclassified by the World Health Organization (WHO) after International Federation of Gynecology and Obstetrics (FIGO)'s staging procedures for ovarian cancer in 2014. The new classification is based on current histopathological findings. The FIGO classification is dependent on the tumor grade, while the WHO classification is dependent on the tumor type (42).

The classification of the ovarian tumor by FIGO indicated that the surface epithelium and epithelial ovarian tumors were derived from the ovary, before 2014. According to FIGO, in the more consistent classification it has been shown that a new class called seromucinous tumors are added, but the previous class of the transitional cell tumors were erased from the classification. Thanks to the new classification, it was determined that high grade serous carcinomas were associated with tubal carcinogenesis. In accordance with WHO, classification emphasized that it is not possible to detect the origin of high-grade serous cancers (42,43). The current WHO classification of epithelial ovarian tumors includes six subgroups:

Serous tumors (Serous low-grade carcinoma and Serous high-grade carcinoma)

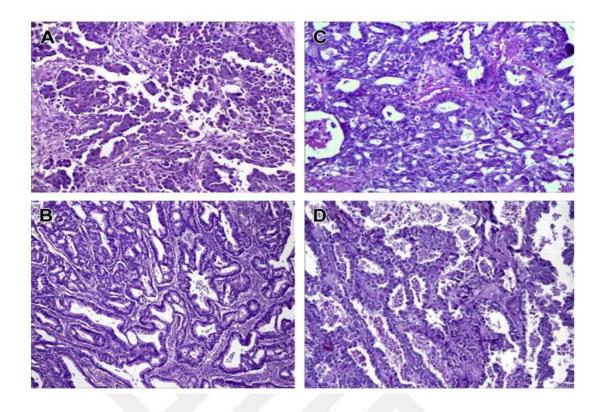
Mucinous tumors

Endometrioid tumors

Clear cell tumors

Transitional cell tumors

Seromucinous tumors (42).



**Figure 2.6.** The presentation of types of ovarian carcinoma (A) Serous tumor (B) Mucinous tumor (C) Endometrioid tumor (D) Clear Cell tumor. (Stain: hematoxylin and eosin) (44).

The FIGO classification ensures precision treatment for patients with ovarian cancer via the molecular classification of gynecologic malignancies. Moreover, these classifications are able to clarify to the patients' clinical risk accurately (45).

#### 2.4.4. Risk Factors of Ovarian Cancer

There are a number of risk factors for ovarian cancer. The first one is the family history, especially the first and second degree family members diagnosed with ovarian cancer. However, ovarian cancers are mostly detected as sporadic, although ovarian cancers are about 5% to 10%. On the other hand the age is also a potentially risk factor on ovarian cancer (46,47,48).

It was indicated that development of ovarian and breast malignancy which were diagnosed at an early stage were significantly associated with family history, by Whittemore et al. and Kauff et al. (49,50). The other risk factors are nulliparity and infertility, while the multiparity, oral contraceptives, and hysterectomy are protective factors. It has been shown that the risk of ovarian cancer is decreased by half for five years by using oral contraceptives (47).

Increased Risk	Decreased Risk
Delayed childbearing	Breastfeeding for 18 months or more
Early menarche	Early menopause
Endometriosis	Multiparity (risk decreases with each
Estrogen replacement therapy for more	additional pregnacy)
than five years	Hysterctomy
Family History suggesting genetic	Late menarche
predisposition	Low fat diet
Genetic syndromes	Tubal Ligation
Hight fat diet	
Late menopause	
Low parity	

**Table 2.2.** The factors affecting Ovarian cancer (51).

In Table 2.2. factors that increase and decrease the development of ovarian cancer are presented. The factors such as delayed childbearing, early menarche, low parity endometriosis and high-fat diet decrease the risk, while some factors such as early menopause, hysterectomy and low-fat diet increase the risk for ovarian cancer (51).

#### 2.4.5. The Diagnosis and Treatment of Ovarian Cancer

#### 2.4.5.1. Diagnosis of Ovarian Cancer

There are many tests such as imaging tests and blood tests used for the diagnosis of ovarian cancer. After a physical exam is performed by doctors, they may perform some tests for the accurate diagnosis of ovarian cancer. Imaging tests include Ultrasound, Computed Tomography (CT) scans, Barium Enema X-ray, Magnetic Resonance Imaging (MRI) scans, Chest x-ray, and Positron emission tomography (PET) scans, whereas other tests include Laparoscopy, Colonoscopy, and Biopsy (52).

The undetermined symptoms such as weight loss, abdominal distension and thamuria are seen at an early stage in individuals with ovarian cancer. Due to delays in ovarian cancer diagnosis, it can be diagnosed at an advanced-stage. Unfortunately, in this case the treatment of the disease is not possible (34,53). Early cancer diagnosis improves survival rates, thus new diagnosis strategies are needed for the early diagnosis of ovarian cancer and effective treatment (54).

The routine pelvic and transvaginal ultrasound examination and blood test, which includes the tumor marker cancer antigen 125, is recommended for individuals with a high risk for ovarian cancer. However, there are no effective results with ultrasound examination and blood test (55).

No effective results have been obtained from performing with ultrasound and CA125 in clinical trials. The studies in recent years imply that diagnostic criteria are inadequate to contribute to ovarian cancer screening, thus recent research continues to determine new candidate biomarkers for this disease (56).

#### 2.4.5.2. Treatment of Ovarian Cancer

The treatment options used for ovarian cancer are chemotherapy and surgery over the years. The effected tissue is staged, surgically debulked and chemotherapy is started (57,58). The emergency and more aggressive treatments are applied to the patients with advanced stage ovarian cancer, but it generally results in poor prognosis (59, 60).

Targeted therapy is one of the new treatment modalities, using some substances such as drugs that lead to minimal damage to noncancerous cells, while only cancerous cells are affected. To give an example, Catumaxomab and Poly (ADP-ribose) polymerases (PARPs) have different functions. Catumaxomab targeting tumor cells is a drug, specifically used for people who carry malignant ascites. Poly (ADP-ribose) polymerases (PARPs) are enzymes, that regulate cell survival and death. In ovarian cancer with BRCA1 and BRCA2 mutation this treatment is used, however, new approaches have determined that ovarian cancers may be resistent to PARP inhibitors (61,62).

#### 2.6. Catechol-O-Methyltransferase (COMT)

#### 2.6.1. General Characteristics and Chromosome Localization

The structural organization of the Catechol-O-Methyltransferase (COMT) gene was described by Lundström et al. (63). Lundstrom et al. used synthetic oligonucleotides to isolate cDNA clones. It was shown that the clones have an open reading frame, a 24.4-kD polypeptide is coded by the clones. The results obtained by DNA analysis considered that many organisms such as humans and monkeys may have 1 gene for COMT (64).

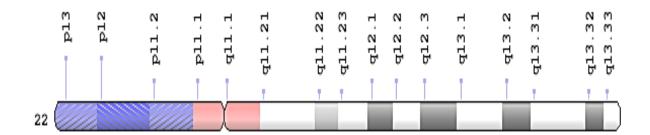


Figure 2.7. The chromosomal location of the COMT gene (65).

In Figure 2.7, the chromosomal location is shown as dark and light bands belonging to the chromosome in the gene. The dark and light bands are obtained by using with a chemical solution microscopically (65).

According to the southern blot analysis, COMT gene is located on chromosome 22 and the chromosomal region of q11.2. COMT gene includes six exons, of which exon 1 and 2 are noncoding (66,67). COMT gene products are expressed in many tissues like bone marrow, brain, bladder, heart, kidney, liver, lung, ovary and so on (68). The synthesized COMT enzyme participates in the DNA repair mechanism (69).

Many polymorphisms have been identified in the coding region of the COMT gene; these are codon 158 (G $\rightarrow$ A), codon 72 (G $\rightarrow$ T) and codon 62 (C $\rightarrow$ T). At codon 158 of the COMT gene Valine alters to Methionine. These alterations in the COMT gene lead to a decrease in the function of proteins (Figure 2.8.) (70,71).

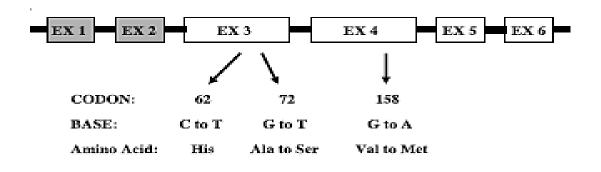
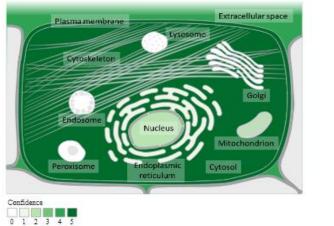


Figure 2.8. Schematic structure of COMT gene (72).

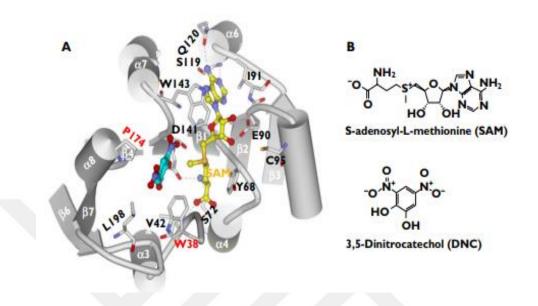
COMT proteins are synthesized from the COMT gene. COMT proteins have been shown to have two groups, these are a soluble shape (S-COMT), which is a shorter form containing 221 amino acids and a membrane-bound shape (MB-COMT) that is a longer form containing 271 amino acids (64,73). S-COMT and MB-COMT proteins differ from each other because they have different N-termini regions (68). The two distinct ATG (which are called start codons) are for promotors P1 and P2; these codons are located on exon 3 of the COMT gene (74).



Compartment	Confidence
plasma membrane	5
cytosol	5
extracellular	4
mitochondrion	2
nucleus	2
endoplasmic reticulum	1

Figure 2.9. Density of COMT protein in the human cell (75).

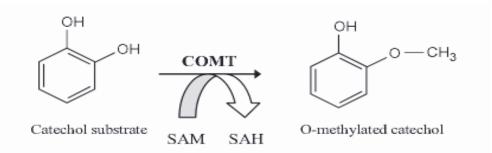
COMT protein is present in large amounts within the plasma membrane in contrast with the other compartments of the cell (75).



**Figure 2.10.** A)- Schematical representation of the tertiary structure of the COMT enzyme in the human cell, B)- The structures of S-Adenosylmethionine and 3,5-Dinitrocatechol (76,77).

In the tertiary structure of the COMT enzyme magnesium ion, chloride ion, S-Adenosylmethionine, and 3,5-Dinitrocatechol are present (77). The COMT protein has both the substrate-binding and SAM-binding sites (Figure 2.10.A) (76).

For the first time, Axelrod et al. explained the features of the COMT enzyme by using the soluble part of organ tissues such as the liver in 1958 (78,79).



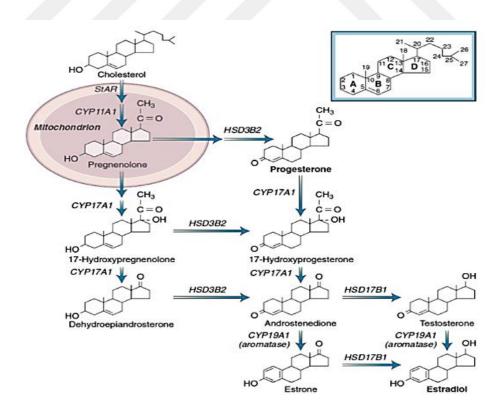
**Figure 2.11.** Representation of the COMT catalyzed reaction (SAM is explained as S-adenosyl-L-methionine and SAH is described as S-adenosyl-L-homocysteine) (80).

#### 2.6.2. Role of Catechol-O-Methyltransferase Enzyme in Estrogen Metabolism

#### 2.6.2.1. Estrogen Metabolism and Formation of Catechol Estrogens

Cholesterol has a 27-carbon and transforms steroid hormones which have 21-, 19-, and 18-carbons. Cholesterol moves into the mitochondrion in a cell to form pregnenolone or androstenedione as a precursor, after that it goes out from the mitochondria; pregnenolone produces and converts to estradiol step-by-step. Several enzymes such as CYP11A1, P450, CYP17A1, HSD3B2, CYP19A1 and HSD17B1 catalyze steroid hormone synthesis. Estrogens have 18-carbon, a phenol group at 17-carbon, two hydroxylic groups and an aromatic ring such as a phenol group at the third carbon (Figure 2.12.) (81,82,83).

In vivo, the estrogens estrone (E1), estradiol (E2), and estriol (E3) play a crucial role (84). The proliferation of ovarian cells is stimulated by these compounds secreted in ovaries at ovulation (85).



**Figure 2.12.** The representation of formation of steroid hormone in the human cells (83).

The catechols can stem from endogenous or exogenous substances, therefore the catecholamine and catechol estrogen are both found in endogenous and exogenous substances (86). Some evidence indicates that reactive catechol metabolites cause cancers originating from estrogens. The COMT enzyme, which is synthesized from the COMT gene, catalyzes O-Methylation, therefore, this enzyme deactivates catechol estrogens (87).

Catechol estrogens have reactive and carcinogenic effects. Various enzymes can detoxify these chemical compounds in cells. One of those enzymes is COMT, which formes a methoxy compound. This enzyme catalyzes the transfer of the methyl group in the coenzyme SAM to the hydroxyl group in the catechols (11).

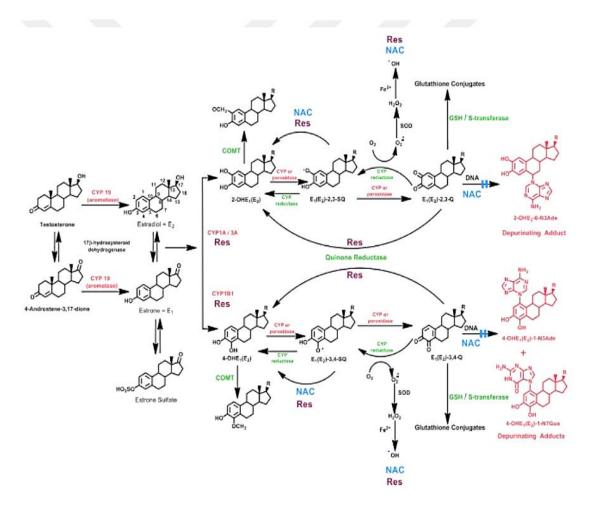


Figure 2.13. The formation of catechol estrogens (88).

In Figure 2.13., protective enzymes are shown in the green color and the enzymes that have a stimulatory effect for the formation of catechol estrogens are shown in red; enzymes and the others shown in red are products of the

apurinic/apyrimidinic site by depurinating DNA adducts in estrogen metabolism. N-Acetylcysteine (NAC) and Resveratrol (Res) affect estrogen metabolism in a positive way by blocking formation of DNA adducts with estrogen (88).

According to research regarding the estrogen metabolism (89,90), it has been implied that estrogen related cancers is initiated by formation carcinogenic catechol estrogens (91,92). The formation of these reactive catechol compounds can damage DNA (84) and have toxic effects against cells (93,94). But, these methoxy estrogens show no estrogenic effects and also lack a strong affinity to estrogen receptors, this reason why these estrogens are inactive (95). In-vitro, 2-methoxy-estrogen have effects on cell lines in cancer against angiogenic and proliferative (96-98) and it has been inhibited DNA synthesis and also mitosis (97).

# 2.6.2.2. Role of Catechol-O-Methyltransferase Val158Met Polymorphism in Ovarian Cancer

COMT, which is a phase II enzyme (78), is one of the estrogen-metabolizing enzymes (99) catalyzes the methylation of catechol estrogen (14). The other phase II enzymes are Glutathione-S-transferase 1 (GSTP1) and N-acetyltransferase 2 (NAT2). The active estrogen metabolites such as catechol estrogen are inactivated by using these enzymes (100,101).

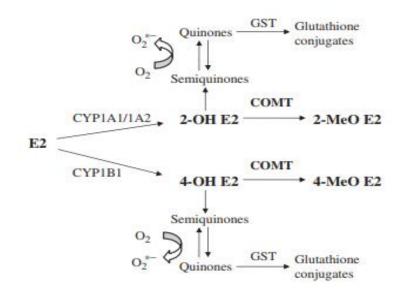


Figure 2.14. The diagram of COMT in estradiol biologically transformation (80).

The polymorphism derived from G to A transition, known as Val158Met in the COMT gene decreases its function. By that transition, valine amino acid converts to methionine amino acid in exon 4. It is shown that Met/Met genotype, which is the homozygous mutant, decreases the COMT enzyme activity by 3-4 times more than the Val/Val genotype called wild-type. The activity of the heterozygous genotype (known as Val/ Met) is intermediate. As a result of COMT, Val158Met polymorphism causes a decrease in enzymatic activity (14).

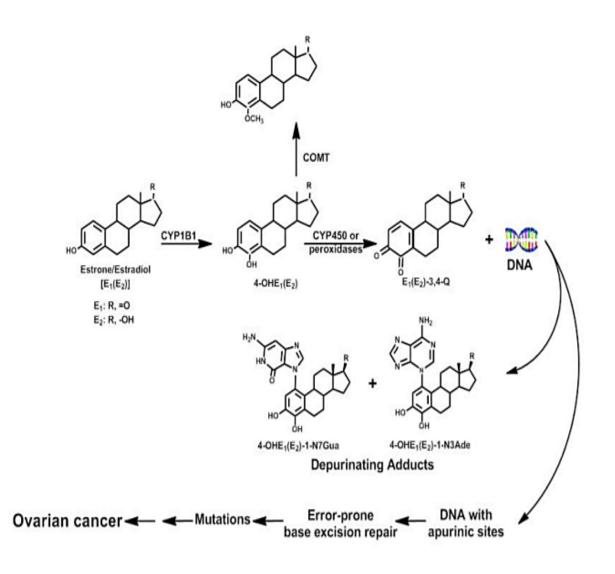


Figure 2.15. The role of the carcinogenic estrogens in cancer initiation (102).

The phase I enzymes involved in estrogen metabolism are cytochrome P450 1A1, 1A2 (CYP1A1/1A2 ) and cytochrome P 450 1B1 (CYP1B1) (100,101).

By these enzymes (CYP1A1/1A2 and CYP1B1) estrogens are converted to 2hydroxylated and 4-hydroxylated metabolites (the 2/4-OH catechol estrogens) via hydroxylation (103,104). The 2/4-OH catechol estrogens are converted 2/4-methoxyestrone/estradiol (2/4-MeOE1/E2) by the COMT enzyme, which catalyzes methylation reactions. Consequently, the 2/4-OH catechol estrogens are inactivated (104). The semiquinones and quinones are produced by oxidization reactions unless 2/4-methoxyestrone/estradiol are observed. If it is the case, then reactive oxygen species are obtained. These reactive molecules damage the DNA and initiate tumor (105).

In conclusion, COMT detoxifies catechol estrogens, which are carcinogenic metabolites. Furthermore, the metabolite 2-methoxy-estradiol is produced, which causes the suppression of angiogenesis and migration of endothelial cells and cytotoxin; it also stimulates apoptosis, which results in the suppression of carcinogenesis (106).

The aim of this study was to reveal the relationship between the COMT gene polymorphism and ovarian cancer in the Turkish population.

#### **3. MATERIALS AND METHODS**

#### **3.1. Sample Selection and Definition**

As part of the study, patients diagnosed with ovarian cancer (n=47) and a control group (n=47) were included. The patient group was composed of patients who were diagnosed by the department of Obstetrics and Gynecology, and the healthy control group consisted of healthy individuals. Ethical approval was obtained from Ethics Committee of Yeditepe University for the study (Ethics committee decision no: 915).

**Control Group:** The control group consisted of healthy individuals aged 18-85 years who were not diagnosed with ovarian cancer following the clinical examination.

**Patient Group:** The patients were composed of individuals within the Yeditepe University Hospital. They were diagnosed with ovarian cancer and their age range was between 18-85 years old.

#### 3.2. Materials and Devices Used in the Experiment

#### **3.2.1.** Materials Used in the DNA Isolation from the Peripheric Blood

The peripheral venous blood samples were set in tubes at -80 °C. The tubes contained Ethylenediaminetetraacetic acid (EDTA), which prevents blood clotting. DNA İsolation Robot (IPrep pure link, Invitrogen and the Thermo Fisher Scientific Inc) system was used for DNA isolation.

The mixture used in the DNA isolation composed of 1.12mg/ml Proteinase K, 8M Guanidine hydrochloride, 10.5 nM EDTA, 10.5 nM NaCl and 10.5 nM Tris-Cl of pH 8,8.

#### **3.2.2.** The Equipment Used in the Experiment

DNA Isolation Robot ( Iprep Purelink, Invitrogen, Thermo Fisher Scientific Inc), Nanodrop 2000 (Thermofisher Scientific Inc), Real-Time PCR ( LIGHT CYCLE 480 II Instrument, Roche Diagnostics, Fast Real Time 7500, Applied Biosystems), 7500 Fast Real-Time PCR Instrument, Plate Centrifuge (Hettich), Centrifuge (Centrifuge 22R-Beckman Coulter), +4 C° Refrigerator (Haier), -20 C° Refrigerator (Haier), Ultra-Pure Water ( Pure Lab Option Q,Elga), Vortex (V.I. Plus Biosan) and a Pipette Kit (Thermo Fisher Scientific Inc.) were used in this experiment.

#### 3.3. Methods

#### 3.3.1. Genomic DNA Isolation from Blood

All venous blood samples of the patient and control groups were taken into the tubes with EDTA in a volume of 5 ml. Blood samples were stored in a refrigerator at + 4 ° C until DNA isolation. DNA isolation was performed by using a robot of iPrep DNA extraction (Invitrogen), and from the blood genomic DNA isolation Kit with iPrep. DNA can be isolated from 350  $\mu$ l peripheric blood by this system, so 13 blood samples could be studied at the same time. One cartridge is used for each of samples, the cartridges are agitated for a period of time to bond the magnetic beads to the DNA efficiently before putting the samples to own cartridge.

The robot of iPrep works according to ChargeSwitch® technology (CST®), as an automated extraction method. A high amount of DNA can be prepared from samples with this method. In this method, amounts of genomic DNA are prepared from samples purely by using paramagnetic particles. These particles are surrounded by a DNAbinding surface. CST® (Charge Switch® Technology) extraction method has a unique principle when compared to the silica-based DNA extraction method. The charge of beads can be changed by the pH of its surrounding buffer. In the event of low pH conditions, the backbone of the DNA is negatively charged, then it binds to the positively charged beads. These charged beads are neutralized by using a low salt buffer that has a higher pH in order to allow for the elution of DNA. Purified nucleic acids pass into the wash buffer, then DNA samples are ready to use (107,108).

At the end of the experiment aqueous DNA samples were obtained and stored at +4 ° C in the refrigerator.

### 3.3.2. Measurement of DNA Purity

UV absorbance of nucleic acids is measured at 260 nm by UV spectroscopy that called NanoDrop. In this spectrophotometer cuvettes or capillaries are not required. DNA concentrations of both OD260/OD280 and OD260/OD230 proportions are determined by NanoDrop. Thanks to the NanoDrop device, the purity as well as the concentration of nucleic acid molecules such as DNA were able to observed. On the other hand, it has not been shown to distinguish several molecules like RNA, nucleotides, double-stranded DNA (dsDNA) or single-stranded DNA (ssDNA) because this device quantifies the absorbance of nucleic acids (109).

In this experiment, we quantified DNA using the NanoDrop 2000 (Thermofisher Scientific Inc). 1,5  $\mu$ l of DNA samples were used. The DNA samples before measurement were diluted in the ratio of 1/100. The sample was put into place for measurement by opening the arm then the device's arm was turned off. After each measurement, the surface was cleaned by distilled water, and thus it could be safe for the next measurement.

 $50 \ \mu\text{g}$  / ml of double-stranded DNA at a wavelength of 260 nm is equal to one Optical Density (OD) Unit. The purity of DNA samples was measured by analyzing the OD260 / OD280 ratio. The suitable ratio of OD260 / OD280 is between 1.7-1.9 when performing genotyping (110).

The following formula was used to calculate the concentration of DNA at 260 nm.

dsDNA concentration =  $50 \mu g/mL \times OD260 \times dilution$  factor

### 3.3.3. Real-Time PCR Conditions for COMT Polymorphism

Genotyping analysis was performed by using the 7500 Fast-Real-Time Polymerase chain reaction (Applied Biosystems) device with Real-Time PCR.

By Real-Time PCR, fluorescence dyes of probes are utilized to determine the single nucleotide polymorphisms (SNPs). It is a system that allows the genotyping by reading the fluorescence radiations. There are two TaqMan probes, one prob is labeled a FAM dye and the other prob is labeled a VIC dye. The fluorescent dye-bound DNA probes bind to the amplified region. The probes are hydrolyzed by Taq polymerase. Fluorescent signals can be easily detected. The probes with fluorescence dye used in the Real-Time PCR have two different wavelengths for allele specific, which are wildtype and mutant alleles, detection.

The primer sequence of COMT reported by Lundström et al. in 1991 is indicated below. This primer was determined according to the sequence of COMT gene in human cells and it was used in this experiment is used (64). In this method, the region containing this polymorphism was increased by using 5' –GGA GCT GGG GGC CTA CTG TG- 3' (Forward) and 5' –GCC CTT TTT CCA GGT CTG ACA- 3' (Reverse) primers. A region of the gene was generated by genotyping and COMT (rs4680) polymorphism was analyzed. The focused gene region of genotyping was rs4680 (G>A) for the COMT gene. Region specific primer and probe sets are used 'TaqMan Genotyping Assays' and fluorescence dyes of probes are given below. Allelic discrimination has been shown using the software of the 7500 Fast Real Time PCR tool.

### Forward 5' -GGA GCT GGG GGC CTA CTG TG-3'

### Reverse 5' -GCC CTT TTT CCA GGT CTG ACA- 3' (64).

## **3.3.3.1. Real Time Protocol**

Real-time PCR reagents and the mixture of reaction have been recorded in Table 3.1. Total volume for each sample was determined by the protocol.

The Material Used	Quantity
Master Mix	5 µl
TaqMan Genotyping Assay	0.5 µl
DNAse, RNAse Free water	3.5 µl
Template DNA	1 μ1

Table 3.1. The reaction mixture for the Real-Time PCR

The conditions for Real-Time PCR were arranged by waiting for 10 minutes at  $95^{\circ}$  C, accomplishing denaturation for 15 seconds at  $92^{\circ}$  C for each cycle and also connecting/elongation for 1 minute at  $60^{\circ}$  C for each cycle. As illustrated in Table 3.2., denaturation and connecting/elongation were completed for 40 cycles.

Table 3.2. The Real-Time PCR conditions

		40 Cycles	
	Waiting	Denaturation Connecting/Elong	
Temperature	95° C	92° C	60° C
Duration	10 minutes	15 seconds	1 minutes

# **3.3.4. Statistical Analysis**

Student's t-test was used to analyze numeric values. The data obtained from genotyping was evaluated using Chi-square and Fisher's Exact Tests via the SPSS 25.0 Program for statistical analysis. Chi-square and Fisher's Exact Tests were used for evaluating the distributions of genotypes and alleles among groups. p-values less than 0,05 were considered statistically significant.

The possible risk factors in ovarian cancer were evaluated by Logistic Regression Analysis.



## 4- RESULTS

# 4.1. The Obtained Findings Following Statistical Analysis

Demographic data belonging to the patient and healthy control groups were statistically analyzed as shown in Table 4.1.

**Table 4.1.** Demographic data related to the patients with ovarian cancer and healthy controls.( n: number of sample,  $\bar{x} \pm SD$ : mean value  $\pm$  Standard deviation, \*(S)= significantly different (p< 0.05), NS= non significant (p>0.05)).

		Control Group (n=47) %	Patient Group (n=47) %	p-value
Body Mass Ind	ex, $\overline{x} \pm SD$ (kg/m2)	23.04± 3.62	29.31± 5.37	< 0.0001* (S)
U	$\overline{x} \pm SD$ years)	51.11±12.86	54.87±12.55	0.154 (NS)
	ce Area, $\overline{x} \pm SD$ (m2)	1.66±0.14	1.77±0.15	< 0.0001* (S)
0	d Glucose, $\overline{x} \pm SD$ mg/dl)	86.45±7.60	108.52±33.28	< 0.0001* (S)
Alcohol	Yes %	(n=19) 40.4%	(n=1) 2.2%	< 0.0001*
consumption	No %	(n=28) 59.6%	(n=44) 97.8%	(S)
Smoking	Yes %	(n=26) 55.3%	(n=37) 82.2%	< 0.0001*
	No %	(n=21) 44.7%	(n=1) 17.8%	(S)
Menopause	Postmenopause %	(n=19) 40.4%	(n=38) 80.9%	< 0.0001*
	Premenopause %	(n=28) 59.6%	(n=9) 19.1%	(S)
History of	Yes %	(n=0) 0%	(n=12) 26.7%	< 0.0001*
Diabetics	No %	(n=47) 100%	(n=33) 73.3%	(S)
Domitar	$\leq 1$	(n=27) 54.4%	(n=12) 26.7%	< 0.0001*
Parity	>1	(n=20) 42.6%	(n=33) 73.3%	(S)
Pregnant State	≤ 1	(n=27) 54.4%	(n=10) 22.2%	< 0.0001*
	>1	(n=20) 42.6%	(n=35) 77.8%	(S)

As a result of the analysis, body mass index, body surface area and fasting blood glucose values in ovarian cancer patients were found to be significantly higher than the control group (Table 4.1). There were no differences between the patients and controls regarding mean age (p= 0.154). Other demographic data mentioned in Table 4.1 are diabetes (p<0.001), alcohol consumption (p<0.001), and smoking (p<0.001) and these risk factors were found to be statistically meaninful among groups. Smoking (p<0.001) and diabetes mellitus (p<0.001) were significantly higher in patients with ovarian cancer, while alcohol consumption (p<0.001) was significantly higher in the control group.

Table 4.1 shows that menopausal status (premenopausal and postmenopausal), pregnancy status (number of pregnancies  $\leq 1$  or number of pregnancies > 1) and parity (number of births  $\leq 1$  or number of births > 1) are statistically meaningful. In the patients with ovarian cancer, the rate of postmenopausal women (80.9%) was higher than the control group (40.4%) (Table 4.1). The controls whose number of births were less than or equal to one (number of births  $\leq 1$ ) (54.4%) were found to be statistically higher than the patient group (26.7%). The rate of postmenopausal women (80.9%) in the patients with ovarian cancer was higher than the control group. While the rate of pregnancy status (number of pregnancies  $\leq 1$ ) (54.4%) in the control group was higher than the patient group.

The demographic and histopathological parameters belonging to the patient group are given in Table 4.2. While the average value of age was determined as 29.31 in the patient group, the value of the BMI was obtained as 54.87. In the patients with ovarian cancer, the distribution of premenopause (80%) was analyzed to be high in comparison with postmenopause state (20%). When we evaluated in terms of relapse and metastasis of tumors, tumor statements were obtained respectively as 45.5% and 75%. The treatment parameters were resulted that 72.1% of the patients received adjuvant chemotherapy and 27.9 % of the patients did not receive adjuvant chemotherapy. While 36.4 % of the patients received neoadjuvant chemotherapy and 63.6 % of the patients did not receive neoadjuvant chemotherapy. 47.5% of the patients underwent debulking surgery. In Table 4.2. it can be observed that the rates of being

pregnant more than once (77.8%) and a number of births (> 1) were high (73.3%) in the patient.

**Table 4.2.** Demographic and histopathological parameters belonging to the patient group with ovarian cancer

Characteristic features	Average value ( $\overline{\mathbf{x}} \pm \mathbf{SD}$ )
Age	29.31± 5.37
Body Mass Index (kg/m2)	54.87±12.55
Characteristic features	Percentage (%) (n=47)
Menopause	
Postmenopause	20%
Premenopause	80%
Metastasis	
Yes	75%
No	25%
Relapse	
Yes	45.5%
No	54.5%
Adjuvant chemotherapy	
Yes	72.1%
No	27.9%
Neoadjuvant chemotherapy	
Yes	36.4%
No	63.6%
Surgery	
Staging	25%
Debulking	47.5%
Staging+Debulking	32.5%
Pregnant state	
≤1	22.2%
>1	77.8%
Parity	
≤1	26.7%
>1	73.3%

n: number of sample,  $\overline{x}\pm$  SD: mean value  $\pm~$  Standard deviation

The ratio of patients at stage III (42.9%) were found to be high as compared to stage I (23.8%), stage II (23.8%) and stage IV (14.3%). When we evaluated the tumors in terms of cell types, epithelial tumors were found at a high rate (92.3%) in comparison to the other types. Serous epithelial tumors from epithelial tumor types were calculated in a ratio of 56.4%. The ratio of sex-cord stromal tumors (5.3%) was found to be higher than germ cell tumors (2.6%) (Table 4.3).

<b>Table 4.3.</b> The percentage distributions of stages, cell types in the patient group	

Characteristic features	Percentage (%) (n=47)
Stages	
Stage I	23.8%
Stage II	23.8%
Stage III	42.9%
Stage IV	14.3%
Cell Types	
Epithelial tumors	92.3%
Serous tumors	56.4%
Mucinous tumors	10.3%
Endometrioid tumors	5.1%
Clear cell tumors	5.1%
Mixed epithelial tumors	15.4%
Germ cell tumors	2.6%
Sex-cord stromal tumors	5.1%
Germ cell tumors	2.6%
Sex-cord stromal tumors	5.3%

n: number of sample,  $\overline{x} \pm SD$ : mean value  $\pm$  Standard deviation

The ratio of patients with different genotypes in comparison with BMI values are given in Table 4.4. Body mass index ratio (GG, GA, AA, respectively, 27.37, 24.25, 27.10) was found to be nearly the same among all genotypes of COMT gene. Therefore, BMI ratios are statistically meaningful in the patient group. CA125, CEA, CA19\_9 and CA15 levels from metabolic parameters were not found to be statistically significant (0.186, 0.161, 0.829, 0.906, respectively) (Table 4.4).

**Table 4.4.** The distributions of BMI, CA125, CEA and CA19\_9 levels according to genotypes of COMT gene polymorphism in the patient group

	$\begin{array}{c} \mathbf{G}\mathbf{G}\\ (\overline{\mathbf{x}}\pm\mathbf{S}\mathbf{D}) \end{array}$	$\begin{array}{c} \mathbf{GA} \\ (\overline{\mathbf{x}} \pm \mathbf{SD}) \end{array}$	$\begin{array}{c} \mathbf{AA} \\ (\overline{\mathbf{x}} \pm \mathbf{SD}) \end{array}$	p-value
Body Mass Index (kg/m2)	27.37±5.73	24.25±4.74	27.10±5.91	0.041* (S)
CA125	486.88±548	1502.96±1985	1042 ±1951	0.186 (NS)
CEA	16.42±49.62	1.20±0.64	48.15±98.23	0.161 (NS)
CA19_9	265.01±886.51	175.69±469.07	45.34±45.52	0.829 (NS)
CA15	72.43±75.85	106.92±254.94	86.07±93.39	0.906 (NS)

n: number of sample,  $\overline{x} \pm SD$ : mean value  $\pm$  Standard deviation, \* (S)= significantly different (p<0.05), NS= non significant (p>0.05).

In Table 4.5., when we evaluated in terms of metastasis and relapse of ovarian cancer, there was no statistically significant data found (p=0.959, p=0.289 respectively). In the patients with GA and AA genotypes, the ratio of metastasis and relapse of disease was analyzed further in comparison with the GG genotype (Table 4.5.).

	GG % (n=29)	GA % (n=41 )	AA % (n=22)	p-value
Metastasis				
	76.5%	72.5%	75%	
Yes	n=13	n=13	n=6	
No	23.5% n=4	27.8% n=5	25% n=2	0.959 (NS)
Polongo				
Relapse				
	58.8%	66.7%	62.5%	
Yes	n=10	n=6	n=3	
				0.289
	41.2%	66.7%	62.5%	(NS)
No	n=7	n=12	n=5	

**Table 4.5.** The distribution of metastasis and relapse of ovarian cancer according to genotypes of COMT gene polymorphism in the patient group

n: number of sample,  $\overline{x} \pm SD$ : mean value  $\pm$  Standard deviation, \*(S)= significantly different (p<0.05), NS= non significant (p>0.05).

Table 4.6. shows a comparison between COMT genotypes and tumor stages within the patient group. Following analysis, there were no statistically meaningful findings (p=0.788). The patients' carrier A allele was found to be higher in comparison with carrier G allele (Table 4.6.).

**Table 4.6.** The percentage distributions of COMT genotypes in comparison with stageI, II, III and IV in the patient group.

Genotypes Stages	GG % (n=29)	GA % (n=41)	AA % (n=22)	p-value
Stage I	23.5% n=4	23.5% n=4	25% n=2	
Stage II	11.8% n=2	17.6% n=3	37.5% n=3	
Stage III	52.9% n=9	41.2% n=7	25% n=2	0.788 (NS)
Stage IV	11.8% n=2	17.6% n=3	12.5% n=1	

n: number of sample,  $\overline{x} \pm SD$ : mean value  $\pm$  Standard deviation, \* (S)= significantly different (p<0.05), NS= non significant (p>0.05).

Table 4.7. shows a comparison between COMT genotypes and various tumor types within the patient group. There were no statistically significant findings (p=0.788, p=0.478). In epithelial tumors, the homozygote wild type (GG), heterozygote (GA) and homozygote mutant type (AA) distributions were calculated respectively as 93.3%, 93.3% and 100% (Table 4.7.).

Genotypes Cell Types	GG % (n=29)	GA % (n=41)	AA % (n=22)	p-value
Epithelial tumors	93.3% n=14	93.3% n=14	100% n=8	0.788
Sex cord stromal tumors	6.7% n=1	6.7% n=1	0% n=0	(NS)
Mixed epithelial tumors	6.7% n=1	26.7% n=4	12.5% n=1	
Serous tumors	66.7% n=10	33.3% n=5	87.5% n=7	
Mucinous tumors	13.3% n=2	13.3% n=2	0% n=0	0.478
Endometrioid tumors	6.7% n=1	6.7% n=1	0% n=0	(NS)
Clear cell tumors	0% n=0	13.3% n=2	0% n=0	
Sex cord stromal tumors	6.7% n=1	6.7% n=1	6.7% n=0	

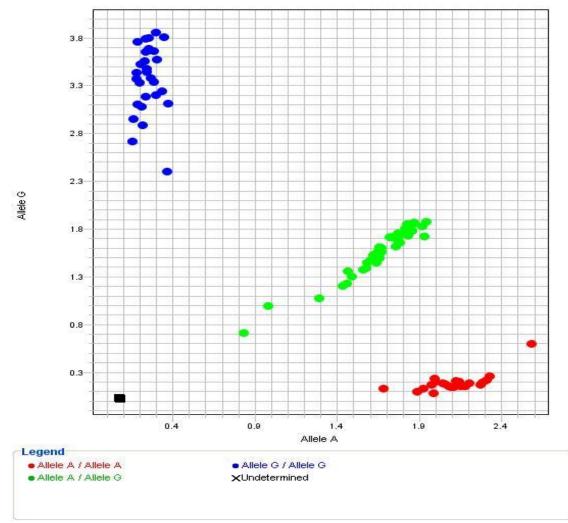
**Table 4.7.** The percentage distributions of COMT genotypes in comparison with tumor types in the patient group.

n: number of sample,  $\bar{x} \pm SD$ : mean value  $\pm$  Standard deviation, \* (S)= significantly different (p<0.05), NS= non significant (p>0.05).

### 4.2. Statistical Evaluation of Real-Time PCR Results

Allelic discriminations were analyzed automatically by the software of the 7500 Fast-Real Time PCR instrument. The readings and interpretations of the fluorescence irradiation are performed by dyes found in the probes. However, some samples could not be discriminated.

FAM dye shows blue color, while VIC dye shows green color. ROX is a reference color for comparing FAM and VIC dyes. Allelic discrimination was analyzed by examining and interpreting the radiance curves (Figure 4.1.).



Allelic Discrimination Plot

Figure 4.1. Allelic Discrimination Analysis of COMT genotype

GG: Homozygote Wild Type GA: Heterozygote AA: Homozygote Mutant Type

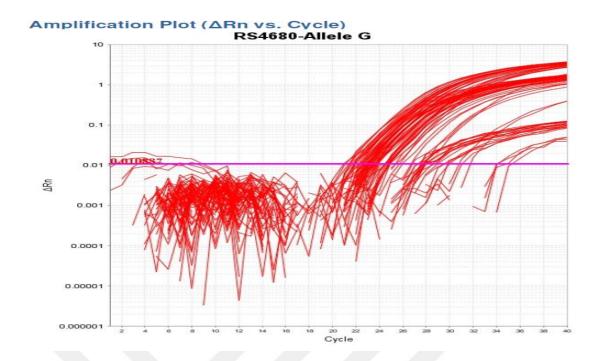


Figure 4.2. Amplification plot display of Allele G

The figure 4.2. shows amplification plots of Allele A. Threshold value (0.0108) is shown as a pink line.

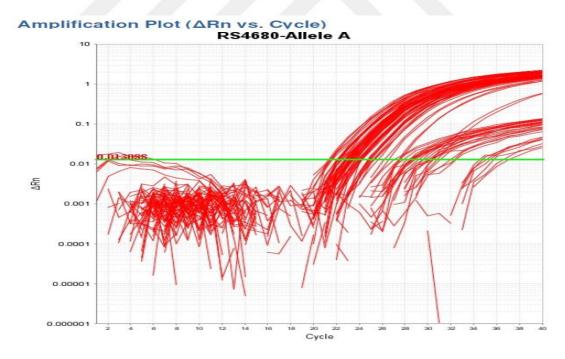


Figure 4.3. Amplification plot display of Allele A

The figure 4.3. shows amplification plots of Allele A. The fluorescent signal pass over the threshold to obtain any data. The threshold value (0.013) is shown as a green line.

In Figure 4.4.,4.5. and 4.6., Allele 1 (Allele A) shows as a blue color, while Allele 2 (Allele G) shows as a red color. Graphs are plotted depending on the quantity of radiation in the samples per cycle (Figure 4.5, Figure 4.6 and Figure 4.7).



Figure 4.4. The emission graph of the homozygote wildtype genotype

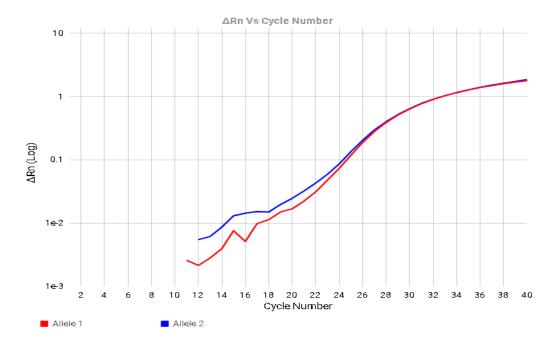


Figure 4.5. The emission graph of the heterozygote genotype

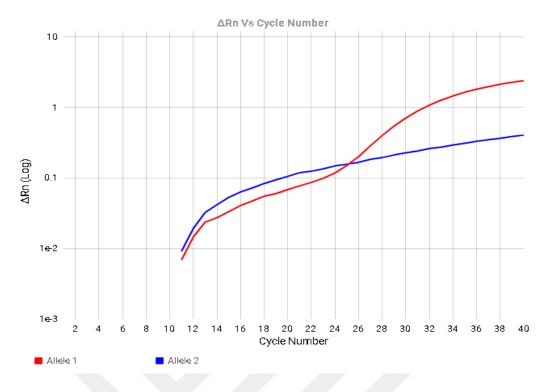


Figure 4.6. The emission graph of the homozygote mutant genotype

### 4.3. Analysis of Genotype and Allele Related to the Patient and Control Groups

In this experiment COMT Val158Met polymorphism in patients with ovarian cancer and healthy control groups. This study found no statistically significant findings among the two groups (p=0.413).

In Table 4.8., shows that there is no significant relation between the patient and control groups. The value of the homozygote mutant genotype is 0.216, that of heterozygote genotype is 0.658, while homozygote wild type genotype's value is 0.547.

According to our findings, homozygote wild type (GG), heterozygote (GA) and homozygote mutant type (AA) rates are calculated respectively as 37.8%, 42.2% and 20.0%. On the other hand, homozygote wild type (GG), heterozygote (GA) and homozygote mutant type (AA) rates are determined respectively as 25.5%, 46.8%, and 27.7% in the control group (Table 4.8.).

Table 4.8 shows that the value of G allele (56.4%) in patient group was found to be higher than the control group (48.9%). Accordingly, the value of A allele is also not statistically significant distinctly among groups like in G allele (p=0.301) (Table 4.8.).

Table 4.8.         The genotype and allele distributions for the COMT Gene between the	•
patient and control groups.	

COMT Genotypes	Control Group (n=47)	Patient Group (n=47)	p value	Odd Ratio (OR)	Confidence interval 95%
GG	(25.5%) n=12	(37.8%) n=17	0.216 (NS)	1.77	0.727-4.314
GA	46.8% n=22	(42.2%) n=19	0.658 (NS)	0.83	0.364-1.892
АА	(27.7%) n=13	(20%) n=9	0.547 (NS)	0.747	0.289-1.932
Alleles Distributions					
G	(48.9%) 46	(56.4%) 53	0.389 (NS)	1.529	0.579-4.037
А	(51.1%) 48	(39.4%) 37	0.301 (NS)	0.630	0.262-1.516

n: number of sample, \*(S)= significantly different (p<0.05), NS= non significant (p>0.05).

.

To sum up, when COMT gene Val158Met polymorphism is compared in terms of genotype and allele frequencies, no significant relation is obtained among groups (p=0.413) (Table 4.9.).

Table 4.9. shows that there is no statistically significant difference between individuals with homozygote variant type genotype (AA) in comparison to individuals with homozygote wild type genotype (GG) and heterozygote genotype (GA) (p=0.547). Also, there is no statistically significant difference between individuals with homozygote wild type genotype (GG) in comparison to individuals with homozygote variant type genotype (AA) and heterozygote genotype (GA) (p=0.206) (Table 4.9).

COMT Genotypes	Gr	ntrol oup =47)	Patient Group (n=47)		p-value	Odd Ratio OR	Confidence interval 95%	
	n	%	n	%				
GG	12	25.5	17	37.8	0.206	0.565	0.232-1.376	
GA+AA	35	74.5	28	62.2	(NS)	0.505	0.232-1.370	
AA	13	27.7	10	22.2	0.547	1.33	0.518-360	
GA+GG	34	72.3	35	77.8	(NS)	1.55	0.510-500	

 Table 4.9. Genotypes of COMT gene polymorphism according to logistic regression analysis

n: number of sample, \*(S)= significantly different (p< 0.05), NS= non significant (p>0.05).

### **5- DISCUSSION AND CONCLUSION**

Several studies have shown that cancer has a wide spread all around the world, and the death rate of cancer is increasing in males and females dramatically (111).

Ovarian cancer is one of the most common cancer types in women and is a heterogeneous disease, and morbidity rate of this disease is highest among gynecologic malignancies. There are many risk factors for ovarian cancer such as obesity, diabetes mellitus, aging, smoking and alcohol intake, inherited ovarian cancer. Several molecular signaling pathways play roles in ovarian cancer such as apoptosis, angiogenesis and oxidative stress; these mechanisms affect progression of this disease. In the United Kingdom, almost 7500 women are diagnosed annually. However, there is standard treatment including surgery and chemotherapy for ovarian cancer (7,112).

When the individuals with ovarian cancer are at a late-stage, 70% of patients are able to be diagnose. On the other hand, symptoms of ovarian cancer are not clear, and survival rate is almost 90% for 5-years during the first stages (113).

Ovarian cancer can not be diagnosed at the early stage due to a lack of a specific biomarker to detect this disease. To date, CA125 as a serum biomarker is the most commonly used. But, knowledge of CA125 of levels is not sufficient to diagnose ovarian cancer (114). In our study, we analyzed distributions of CA125 level according to genotypes of COMT gene polymorphism in the patient group. As a result, the homozygote wild type (GG), heterozygote (GA) and homozygote mutant type (AA) distributions were calculated according to CA125 levels respectively as  $486.88\pm548$ ,  $1502.96\pm1985$  and  $1042\pm1951$  in the patients. No association was found between the COMT gene and levels of CA125 (p=0.186).

It is considered that Catechol-O-Methyltransferase enzyme plays a significant role in estrogen metabolism based on many different studies (115-119). To give an example, Tolba et al. investigated that COMT might be as a biomarker, which can be important factor in suppressing tumor development and treatment of cancer (120). In estrogen metabolism, COMT prevents DNA damage, therefore, it is called the gatekeeper. Wu et al. reported that COMT transcription is decreased related to epigenetic changes such as DNA methylation (9). In light of these developments, our study will contribute to understanding molecular mechanisms for ovarian cancer. There are many case-control studies associated with ovarian cancer and COMT Val158Met polymorphisms, but most of these results have been inconclusive.

For example, Goodman et al. reported that COMT Val158Met polymorphism was not related to ovarian cancer risk due to a limited sample size. The lack of relation does not differ according to the individual's age, ovarian cancer histology or family history (117).

Holt et al. and Sellers et al. studied several enzymes such as CYP1A1, CYP1B1, COMT and SULT1A1 related to estrogen metabolism. The polymorphisms regarding these genes were performed by genotyping. The obtained data analyzed by using logistic regression tests. As a result of these studies, they found that there was no strong relationship between COMT genotypes or the other gene polymorphisms (CYP1A1, CYP1B1, SULT1A1) and ovarian cancer development in the American population (121,122).

Goodman et al. indicated that no meaningful association was seen between the COMT genotype and ovarian cancer risk. That study contained 108 cases and 106 controls from the German population. The ratio of heterozygote genotype carriers (50%) with ovarian cancer was found to be high in comparison with homozygote wild and variant type genotype carriers (25%, 25%) in the cases. There was no evidence that COMT Val158Met polymorphism increases the risk of ovarian cancer (p=0.73), and Goodman et al. implied that advance studies are required to explain different combinations of polymorphisms in estrogen metabolizing enzymes (116).

Similarly, Delort et al. stated that the ratio for ovarian cancer was 2.02 in French subgroups. In this study seven genes that play a role in estrogen metabolism were included. They proposed that advanced studies were required to determine risk factors for ovarian cancer. The homozygote wild type (GG), heterozygote (GA) and homozygote mutant type (AA) distributions were determined as 23.7%, 48%, and 28.3% in the control group, respectively. In the patient group, genotype distributions of GG, GA and AA were found to be 21.6%, 43.1% and 35.3%, respectively. In this study it has been worked with limited number of sample and this situtation was prevent our results (123).

Pan et al. showed relations between COMT Val158Met polymorphism and the development of ovarian carcinogenesis. As a result of meta-analysis, there was no significant correlation related to an increased risk for ovarian cancer. Similar to our results, it was found by the meta analysis that the ratio of the GG genotype was not different in comparison with the ratio of GA+AA genotypes (p=0.76) (124).

It was indicated by Du et al. that there are relations between COMT Val158Met polymorphism and susceptibility of ovarian carcinogenesis. 1,293 cases and 2,647 controls were included regarding Caucasians and other populations in this metaanalysis. They performed a subgroup analysis in regards to ethnicity. In consequence of genotype analysis, there were no meaninful associations for developing ovarian cancer (p>0.05) (17).

In addition to these studies, it is indicated by Garner et al. that a decreased risk for mucinous ovarian tumors is associated with American females who carry variant COMT alleles (A) (OR 0.28, 95% CI 0.13–0.61) (125). Our findings showed that GG genotype was found in 2 (13.3%), GA genotype in 2 (13.3%), and AA genotype in 0 (0%) of the patients with mucinous tumors. Consequently, our results support the findings accomplished Garner et al.

Smoking affects on tumor formation in ovarian cancer especially, and it is the known as the first risk factor. In our study, smoking was found statistically meaningful (p<0.001). Smoking was significantly higher in patients with ovarian cancer (82.2%) than in the control group (55.3%). These results show that smoking may be an important risk factor for developing ovarian cancer (126).

Moreover, it was reported by Goodman et al. that the risk of ovarian cancer increases 2.2 fold in Hawaiian smokers with a low-activity COMT allele. They showed that is no association between individuals with homozygotes wild type genotype (Val/Val) and ovarian cancer risk (p=0.17) (117).

Ovarian cancer development is influenced by various risk factors such as BMI, age, tumor histology, family history of patients with ovarian cancer and smoking. Therefore, there are many factors that create a risk for ovarian cancer. In a nutshell, it is not sufficient to diagnose individuals simply because they carry the Val158Met polymorphism in the COMT gene.

Our findings correspond with the studies accomplished by Delort et al., Holt et al., Goodman et al., Du et al., Sellers et al. and Pan et al. In our study, we examined the presence of the COMT gene Val158Met polymorphism among patients with ovarian cancer and a control group in the Turkish population. Patients with ovarian cancer (n=47) and healthy controls (n=47) were included in this study. First of all, DNA isolation was performed by using a robot of iPrep DNA extraction and with the blood genomic DNA isolation Kit by iPrep. UV absorbance and the purity of nucleic acids was measured at 260 nm by the NanoDrop device. Genotyping analysis was performed by using a 7500 Fast-Real Time Polymerase chain reaction device utilizing Real-Time PCR method. The demographic and histological data were evaluated statistically. Student T-test, Chi-square and Fisher's Exact Tests were used for analyzing the distributions of genotypes and alleles among groups.

The distributions of genotypes and alleles were analyzed for ovarian cancer among the patient and control groups. In our study, the GG genotype was found in 12 (25.5%), GA genotype in 22 (46.8%), and AA genotype in 13 (27.7%) regarding the control group. In the patient group, GG, GA and AA genotypes were determined 17 (37.8%), 19 (42.2%) and 9 (20%) respectively. GA heterozygote genotype carriers were observed in 42.2% of the patient group compared to other genotypes (GG and AA). There was no significant variation in comparison with genotypes between patient and controlgroups(p=0.413).

### In consequence of this research;

This study is the first and only study showing the relationship between ovarian cancer and the COMT gene Val158Met polymorphism within the Turkish population. It is represented that there are no statistically meaningful relationships among genoytpes belonging to the patient and control groups in this study.

It is proposed that this study should be performed further in the future due to some limitations such as patients'medical history. COMT may be proposed as a new biomarker for ovarian cancer. This study should be further carried out by increasing the sample size of the study groups to determine a more exact knowledge for this disease.

Although COMT polymorphism does not have a direct relationship with ovarian cancer patients, and so this polymorphism does not carry any risk for the Turkish population.

### **6. REFERENCES**

- 1) Thompson MW, McInnes RR, Willard HF. *Genetics in Medicine*. 6th ed.Genetics and Cancer. W.B. Saunders Company: Philadelphia; 2001.
- 2) Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jema A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA: CA Cancer J Clin.* 2018; 0: 1–31.
- 3) Hennessy BT, Coleman RL, Markman M. Ovarian cancer. *Lancet.* 2009; 374: 1371-1382.
- 4) Jemal A, Bray F, Center MM, et al. Global cancer statistics. *CA Cancer J Clin*. 2011; 61: 69-90.
- 5) Ferlay J, Soerjomataram I, Dikshit R, et al. Cancer incidence and mortality worldwide: sources, methods and major patterns in GLOBOCAN 2012. *Int J Cancer*. 2015; 136: 359-386.
- 6) World Health Organization, International Agency for Research on Cancer, Global Cancer Observatory, Access 01.04.2019, Available at, https://gco.iarc.fr/tomorrow/home.
- 7) Zare H, Shafabakhsh R, Reiter RJ, Asemi Z. Melatonin is a potential inhibitor of ovarian cancer: molecular aspects. *J Ovarian Res.* 2019; 12(1): 26.
- 8) Dal NA, Ertem G. Jinekolojik Kanserler Farkındalık Ölçeği Geliştirme Çalışması. *Itobiad: Journal of the Human & Social Science Researches*. 2017; 6(5).
- 9) Wu Q, Odwin-Dacosta S, Cao S, et al. Estrogen down regulates COMT transcription via promoter DNA methylation in human breast cancer cells. *Toxicol Appl Pharmacol.* 2019; 367:12-22.
- 10) Mannisto P, Ulmanen I, Lundstrom K, et al. Characteristics of catechol Omethyltransferase (COMT) and properties of selective COMT inhibitors. *Prog Drug Res.* 1992; 39: 291-350.
- 11) Guldberg HC, Marsden CA. Catechol-O-methyl transferase: pharmacological aspects and physiological role. *Pharmacol Rev.* 1975; 27: 135-206.
- 12) Mao C, Wang XW, Qiu LX, et al. Lack of association between catechol-Omethyltransferase Val108/158Met polymorphism and breast cancer risk: a metaanalysis of 25,627 cases and 34,222 controls. *Breast Cancer Res Treat.* 2010; 121: 719-725.
- 13) Weinshilboum RM, Raymond FA. Inheritance of low erythrocyte catechol-omethyltransferase activity in man. *Am J Hum Genet*.1977; 29: 125.
- 14) Dawling S, Roodi N, Mernaugh RL, et al. Catechol-Omethyltransferase (COMT)-mediated metabolism of catechol estrogens comparison of wild-type and variant COMT isoforms. *Cancer Res.* 2001; 61: 6716-6722.
- 15) Hirata H, Hinoda Y, Okayama N, et al. COMT polymorphisms affecting protein expression are risk factors for endometrial cancer. *Mol carcinog*.2008; 47: 768-774.
- 16) Tian C, Liu L, Yang X, et al. The Val 158 Met polymorphism in the COMT gene is associated with increased cancer risks in Chinese population. *Tumor Biol.* 2014; 35: 3003-3008.

- 17) Du JZ, Dong YL, Wan GX, et al. Lack of association between the COMT rs4680 polymorphism and ovarian cancer risk: Evidence from a meta-analysis of 3,940 individuals. *Asian Pac J Cancer Prev*.2014; 15: 7941–7945.
- 18) Ayhan A, Durukan T, Günalp S, Gürgan T, Önderoğlu LS, Yaralı H, Yüce K. ed. *Temel Kadın Hastalıkları ve Doğum Bilgisi*, 2nd ed. Güneş Tıp Kitapevleri: Ankara; 2008.
- 19) Lewis R. *Human Genetics Concepts And Applications*. 11th ed. Cancer Genetics and Genomics. McGraw-Hill Education: New York; 2015.p.351-371.
- 20) T.C. Sağlık Bakanlığı Halk Sağlığı Genel Müdürlüğü, 2014 yılı Türkiye Kanser İstatistikleri, Access 14.04.2019, Available at https://hsgm.saglik.gov.tr/depo/birimler/kanser-db/istatistik/2014 RAPOR.\_uzuuun.pdf.
- Siegel RL, Miller KD, Jemal A. Cancer Statistics, CA Cancer J Clin. 2019 ;69:7– 34.
- 22) L.Richardson D, New and Novel Therapies for Gynecologic Cancers. *Semin Oncol Nurs*.2019;35:217-219.
- Siegel RL, Miller KD, Jemal A. Cancer statistics. CA Cancer J Clin. 2018;68: 7– 30.
- 24) Chen VW, Ruiz B, Killeen JL, Cote TR, Wu XC, Correa CN. Pathology and classification of ovarian tumors. *Cancer* 97.2003: 2631–2642.
- 25) Miranda, A. M. & Schnatz, R. H. Ovary Anatomy [serial online]. Medscape. Accessed 23.03.2019. Available at: https://emedicine.medscape.com/article/1949171-overview#a1.
- 26) Heintz APM, Odicino F, Maisonneuve P, et al. Carcinoma of the ovary. In: 26<sup>th</sup>annual report on the results of treatment in gynecological cancer. Int J Gynecol Obstet 95.2006.
- 27) Forabosco A, Sforza C, De Pol A, Vizotto L, Marzona L, Ferrario VF. Morphometric study of the human neonatal ovary. *Anat Rec.* 1991;231:201-8.
- 28) Sowers MF, et al. *Menopause: Biology and Pathobiology*. In: Menopause: biology and pathobiology. Academic Press, New York, NY, 2000. p.175-188.
- 29) Felig P, Frohman LA. *Endocrinology and Metabolism*, The McGraw-Hill Companies :United States. 2001. p.708.
- 30) Beard CM, Hartmann LC, Atkinson EJ, O'brien PC, Malkasian GD, Keeney GL, Melton LJ. The epidemiology of ovarian cancer: a population-based study in olmsted county, Minnesota, 1935-1991. Ann Epidemiol. 2000;10:14-23.
- Badgwell D, Bast R. Early detection of ovarian cancer. *Dis Markers*. 2007; 23: 397-410.
- 32) Ovarian Cancer Surgery [serial online].KK Women's and Children's Hospital . Accessed 17.04.2019, Available at: https://www.kkh.com.sg/patientcare/conditions-treatments/ovarian-cancer-surgery
- 33) Greenlee RT, Murray T, Bolden S, Wingo PA. Cancer statistics. CA Cancer J Clin. 2000;50:7–33.
- 34) Cancer Facts & Figures 2019. Access 18.04.2019. Available at: https://www.cancer.org/content/dam/cancer-org/research/cancer-facts-andstatistics/annual-cancer-facts-and-figures/2019/cancer-facts-and-figures-2019.pdf

- 35) Gong, L, Wang C, Gao Y, Wang J. Decreased expression of microRNA-148a predicts poor prognosis in ovarian cancer and associates with tumor growth and metastasis. *Biomed Pharmacother*.2016; 83: 58-63.
- 36) Runnebaum IB and Stickeler E. Epidemiological and molecular aspects of ovarian cancer risk. *J Cancer Res Clin Oncol*.2001; 127(2): 73-79.
- 37) Noone AM, Howlader N, Krapcho M, et al. (eds). SEER Cancer Statistics Review, 1975-2015, National Cancer Institute, Bethesda, MD, http://seer.cancer.gov/ csr/1975\_2015/, based on November 2017 SEER data submission, posted to the SEER website April 2018.
- 38) T.C. Sağlık Bakanlığı, Türkiye Halk Sağlığı Kurumu, Kanser Daire Başkanlığı, Access 18.04.2019. Available at: https://dosyasb.saglik.gov.tr/Eklenti/30148,ingilizcesiydijiv1pdf.pdf?0.
- 39) Espina V, Liotta AL. (Eds). *Molecular Profiling: Methods and Protocols*. New York: Humana Press; 2012.
- 40) Cancer Research U.K. 2019. Ovarian Cancer [serial online]. Cancer Research UK. Access 22.04.2019. Available at at:https://www.cancerresearchuk.org/about-cancer/ovarian-cancer.
- 41) Boussios S, Zarkavelis G, Seraj E, Zerdes I, Tatsi K, Pentheroudakis G. Nonepithelial Ovarian Cancer: Elucidating Uncommon Gynaecological Malignancies. *Anticancer Res*, 2016;36(10):5031-5042.
- 42) Meinhold-Heerlein I, Fotopoulou C, Harter P, et al. The new WHO classification of ovarian, fallopian tube, and primary peritoneal cancer and its clinical implications. *Arch Gynecol Obstet*.2016;293:695–700.
- 43) Kurman RJ, Carcangiu ML, Herrington CS, et al. WHO Classification of Tumours of Female Reproductive Organs. In WHO Classification of Tumours. 4. Aufl. Lyon: WHO Press; 2014.
- 44) Ahmed Q, Alosh B, Bandyopadhyay S, et al. Gynecologic cancers: molecular updates. *Clin Lab Med*. 2013; 33(4): 912.
- 45) Abdulfatah E, Ahmed Q, Alosh B et al. Gynecologic Cancers: Molecular Updates 2018, *Clin Lab Med.* 2018; 38: 421–438.
- 46) Hartge P, Whittemore AS, Itnyre J, et al: Rates and risks of ovarian cancer in subgroups of white women in the United States. *Obstet Gynecol* 1994;84:760– 764.
- 47) Holschneider CH, Berek JS. Ovarian cancer: epidemiology, biology, and prognostic factors. *Semin Surg Oncol*. 2000;19:3-10.
- 48) Boyd J, Rubin SC: Hereditary ovarian cancer: molecular genetics and clinical implications. *Gynecol Oncol*. 1997;64:196–206.
- 49) Whittemore AS, Balise RR, Pharoah PDP, et al. Oral contraceptive use and ovarian cancer risk among carriers of BRCA1 or BRCA2 mutations. *Br J Cancer*.2004; 91(11):1911–1915.
- 50) Kauff ND, Domchek SM, Friebel TM, et al. Risk-reducing salpingooophorectomy for the prevention of BRCA1- and BRCA2-associated breast and gynecologic cancer: a multicenter, prospective study. J Clin Oncol.2008; 26(8): 1331–1337.
- 51) Montes AF, et al. Epidemiology and etiology of ovarian cancer. In: Ovarian Cancer-Basic Science Perspective. *IntechOpen*. 2012.

- 52) American Cancer Society, Access 24.04.2019, Available at https://www.cancer.org/cancer/ovarian-cancer/detection-diagnosis-staging/how-diagnosed.html.
- 53) Nakamura K, Sawada K, Yoshimura A, Kinose Y, Nakatsuka E. & Kimura T. Clinical relevance of circulating cell-free microRNAs in ovarian cancer. *Mol Cancer*.2016;15: 48.
- 54) Mayo Clinic, Access 23.04.2019, Available at https://www.mayoclinic.org/diseases-conditions/ovarian-cancer/symptoms-causes/syc-20375941.
- 55) Buys SS, Partridge E, Black A, Johnson CC, Lamerato L, Isaacs C. Effect of screening on ovarian cancer mortality: the Prostate, Lung, Colorectal and Ovarian (PLCO) cancer screening randomized controlled trial. *JAMA*. 2011;305:2295–2302.
- 56) Gupta KK, Gupta VK, et al. Ovarian cancer: screening and future directions, Int J Gynecol Cancer. 2019;29:195–200.
- 57) Jayson GC, Kohn EC, Kitchener HC, Ledermann JA. Ovarian cancer. *Lancet*. 2014;384:1376–1388.
- 58) Stewart C, Ralyea C, Lockwood S. Ovarian Cancer: An Integrated Review, *Semin Oncol Nurs.* 2019;35:151-156.
- 59) Dehal A, Smith JJ, Nash GM, Cytoreductive surgery and intraperitonealchemotherapy: an evidence-based reviewepast, present and future, *J. Gastrointest. Oncol.*2016; 7(1): 143–157.
- 60) Zhang L, Huang J, Yang N, Greshock J, Megraw MS, Giannakakis A, et al. MicroRNAs exhibit high frequency genomic alterations in human cancer. *Proc. Natl. Acad. Sci. U. S. A.*2006; 103(24): 9136–9141.
- 61) American Cancer Society, Access 23.04.2019, Available https://www.cancer.org/content/dam/CRC/PDF/Public/8773.00.pdf.
- 62) Heiss MM, Murawa P, Koralewski P, et al. The trifunctional antibody catumaxomab for the treatment of malignant ascites due to epithelial cancer: Results of a prospective randomized phase II/III trial. *Int J Cancer*.2010; 127(9):2209-21.
- 63) Männistö PT, Kaakkola S. Catechol-O-methyltransferase (COMT):biochemistry, molecular biology, pharmacology, and clinical efficacy of the new selective COMT inhibitors. *Pharmacol Rev.* 1999; 51: 593–628.
- 64) Lundstrom K, Salminen M, Jalanko A, Savolainen R, Ulmanen I. Cloning and characterization of human placental catechol-O-methyltransferase cDNA. *DNA Cell Biol.* 1991;10:181-189.
- 65) NIH, U.S. National Library of Medicine, Available 24.04.2019, Access https://ghr.nlm.nih.gov/chromosome/22#idiogram.
- 66) Park BL, Shin HD, Cheong HS, Park CS, et al. Association analysis of COMT polymorphisms with schizophrenia and smooth pursuit eye movement abnormality. *J. Hum. Genet.* 2009; 54(12): 709.
- 67) Grossman MH, Emanuel BS, Budarf ML. Chromosomal mapping of the human catechol-O-methyltransferase gene to 22q11.1–q11.2. *Genomics*. 1992; 12(4): 822–825.

68) NCBI, Access 18.04.2019 Available https://www.ncbi.nlm.nih.gov/gene/1312#gene-expression.

- 69) Zhou Q, et al. Association between the COMT Val158Met polymorphism and risk of cancer: evidence from 99 case-control studies. *Onco Targets Ther.* 2015; 8: 2791–2803.
- 70) Lachman HM, Papolos DF, Saito T, Yu YM, Szumlanski CL, Weinshilboum RM. Human catechol-O-methyltransferase pharmacogenetics: description of a functional polymorphism and its potential application to neuropsychiatric disorders. *Pharmacogenetics*. 1996; 6(3): 243 250.
- 71) Saito S, Iida A, Sekine A, et al. Identification of 197 genetic variations in six human methyltransferase genes in the Japanese population. *J Hum Genet*. 2001; 46: 529.
- 72) Tanaka Y, Sasaki M, Shiina H, et al. Catechol-O-methyltransferase gene polymorphisms in benign prostatic hyperplasia and sporadic prostate cancer. *Cancer Epidemiol Biomarkers Prev.* 2006; 15(2): 238-244.
- 73) NIH, U.S. National Library of Medicine, Access 24.04.2019, Available at https://ghr.nlm.nih.gov/gene/COMT.
- 74) Tenhunen J, Salminen M, Lundstrom K, Kiviluoto T, Savolainen R, Ulmanen I. Genomic organization of the human catechol O-methyltransferase gene and its expression from two distinct promoters. *Eur J Biochem*. 1994; 223: 1049-1059.
- 75) Gene Cards Human Gene Database. COMT. Access 01.04.2019, Available at https://www.genecards.org/cgi-bin/carddisp.pl?gene=COMT.
- 76) Ma Z, Liu H, Wu B. Structure-based drug design of catecholO-methyltransferase inhibitors for CNS disorders. Br. *J. Clin. Pharmacol.* 2013, 77, 410–420.
- 77) NCBI, Access 24.04.2019, Available at https://www.ncbi.nlm.nih.gov/Structure/pdb/5LSA.
- 78) Axelrod J, Tomchichk R. Enzymatic O-methylation of epinephrine and other catechols. *J Biol Chem*. 1958; 233: 702–705.
- 79) Axelrod J, Senoh S, Witkop B. O-Methylation of catechol amines in vivo. *J Biol Chem.* 1958; 233: 697–701.
- 80) Sak K. The Val158Met polymorphism in COMT gene and cancer risk: role of endogenous and exogenous catechols. *Drug Metab Rev.* 2017; 491: 56–83.
- 81) Carr BR, MacDonald PC, Simpson ER, The role of lipoproteins in the regulation of progesterone secretion by the human corpus luteum, *Fertil Steril 38*. 1982: 303-311.
- 82) Miller WL, Strauss JF, Molecular pathology and mechanism of action of the steroidogenic acute regulatory protein, StAR, J. Steroid Biochem. Mol. Biol. 1999; 69(1-6):131-41.
- 83) Melmed S, Polonsky KS, Larsen PR, Kronenberg HM, *Williams Textbook of Endocrinology*. 12th ed. Saunders press: Philadelphia; 2011. pp. 599–602.
- 84) Zhu BT, Conney AH. Functional role of estrogen metabolism in target cells: review and perspectives. Carcinogenesis. 1998a;19(1): 1-27.
- 85) Lukanova A and Kaaks R. Endogenous hormones and ovarian cancer: epidemiology and current hypotheses. *Cancer Epidemiol Biomarkers Prev.* 2005; 14: 98-107.

- 86) Zou LW, Xu XJ, Liu T, et al. No association between COMT Val158Met polymorphism and prostate cancer risk: A meta-analysis. *Genet. Test. Mol. Biomarkers.* 2013; 17: 78–84.
- 87) Lavigne JA, Helzlsouer KJ, Huang HY, et al. An association between the allele coding for a low activity variant of catechol-O-methyltransferase and the risk for breast cancer. *Cancer Res.* 1997; 57: 5493-5497.
- 88) Cavalieri EL, Rogan EG, Zahid M. Critical depurinating DNA adducts: estrogen adducts in the etiology and prevention of cancer and dopamine adducts in the etiology and prevention of Parkinson's disease. *Int. J. Cancer.* 2017; 141: 1078– 1090.
- 89) Cavalieri EL, Kumar S, Todorovic R, et al. Imbalance of estrogen homeostasis in kidneyand liver of hamsters treated with estradiol: implications for estrogen-induced initiation ofrenal tumors. *Chem Res Toxicol*. 2001; 14: 1041–5042.
- 90) Cavalieri EL, Devanesan P, Bosland MC, et al. Catechol estrogen metabolites and conjugates indifferent regions of the prostate of Noble ratstreated with 4hydroxyestradiol: implications for estrogen-induced initiation of prostate cancer. *Carcinogenesis*. 2002; 23: 329–33.
- 91) Cavalieri E, Rogan E, Chakravarti D. The role of endogenous catechol quinones in the initiation cancer and neurodegenerative diseases. *Methods Enzymol.* 2004; 382: 293-319.
- 92) Zhao Z, Kosinska W, Khmelnitsky M, et al. Mutagenic activity of 4hydroxyestradiol, but not 2-hydroxyestradiol, in BB2 rat embryonic cells, and the mutational spectrum of 4-hydroxyestradiol. *Chem Res Toxicol*. 2006; 19: 475-479.
- 93) Stokes AH, Hastings TG, Vrana KE. Cytotoxic and genotoxic potential of dopamine. *J Neurosci Res*.1999; 55: 659 665.
- 94) Bolton JL, Trush MA, Penning TM, Dryhurst G, Monks TJ. Role of quinones in toxicology. *Chem Res Toxicol.* 2000;13:135-60.
- 95) Merriam GR, MacLusky NJ, Picard MK, Naftolin F. Comparative properties of the catechol estrogens, I: methylation by catechol-O-methyltransferase and binding to cytosol estrogen receptors. *Steroids*. 1980; 36:1-11.
- 96) Lottering ML, Haag M, Seegers JC. Effects of 17 beta-estradiol metabolites on cell cycle events in MCF-7 cells. *Cancer Res.* 1992; 52: 5926-5932.
- 97) Fotsis T, Zhang Y, Pepper MS, et al. The endogenous oestrogen metabolite 2methoxyoestradiol inhibits angiogenesis and suppresses tumour growth. *Nature*. 1994; 368: 237-239.
- 98) Klauber N, Parangi S, Flynn E, Hamel E, D'Amato RJ. Inhibition of angiogenesis and breast cancer in mice by the microtubule inhibitors 2methoxyestradiol and taxol. *Cancer Res.* 1997; 57: 81-86.
- 99) Ashton KA, Meldrum CJ, McPhillips ML, et al. The association of the COMT V158M polymorphism with endometrial/ovarian cancer in HNPCC families adhering to the Amsterdam Criteria. *Hered Cancer Clin Pract*. 2006; 4: 94-102.
- 100) Cheng TC, Chen ST, Huang CS, et al. Breast cancer risk associated with genotype polymorphism of the catechol estrogen-metabolizing genes: a multigenic study on cancer susceptibility. *Int J Cancer*. 2005; 113: 345-353.

- 101) Mitrunen K, Hirvonen A. Molecular epidemiology of sporadic breast cancer. The role of polymorphic genes involved in oestrogen biosynthesis and metabolism. *Mutat Res.* 2003; 544: 9-41.
- 102) Zahid M, Beseler CL, Hall JB, LeVan T, Cavalieri EL, Rogan EG. Unbalanced estrogen metabolism in ovarian cancer. *Int J Cancer*. 2014;134:2414–2423.
- 103) Matos A, Castelao C, Pereira da Silva A, et al. Epistatic interaction of CYP1A1 and COMT polymorphisms in cervical cancer. *Oxid Med Cell Longev*. 2016: 2769804.
- 104) Ball P, Knuppen R, Haupt M, Breuer H. Interactions between estrogens and catechol amines. 3. Studies on the methylation of catechol estrogens, catechol amines and other catechols by the catechol-O-methyltransferases of human liver. *J. Clin. Endocrinol. Metab.* 1972; 34: 736-746.
- 105) Cavalieri EL, Stack DE, Devanesan PD, Todorovic R, et al. Molecular origin of cancer: catechol estrogen-3,4- quinones as endogenous tumor initiators. *Proc. Natl. Acad. Sci. U. S. A.* 1997; 94: 10937-10942.
- 106) Zhu BT, Conney AH. Is 2-methoxyestradiol an endogenous estrogen metabolite that inhibits mammary carcinogenesis? *Canc. Res.* 1998b; 58: 2269-2277.
- 107) Witt S, Neumann J, Zierdt H, Gébel G, Röscheisen C. Establishing a novel automated magnetic bead-based method for the extraction of DNA from a variety of forensic samples. *Forensic Sci. Int Genet.* 2012; 6(5): 539-547.
- 108) Invitrogen, ChargeSwitch Forensic DNA Purification Kits (user manual), Version A, 2005, Access 10.04.2019, Available at: http://tools.thermofisher.com/content/sfs/manuals/chargeswitch\_forensicdna\_ma n.pdf.
- 109) Nakayama Y, Yamaguchi H, Einaga N, Esumi M. Pitfalls of DNA Quantification Using DNA-Binding Fluorescent Dyes and Suggested Solutions. *PLoS One.* 2016; 11(3): e0150528.
- 110) Li X, et al. Comparison of three common DNA concentration measurement methods. *Anal Biochem.* 2014;451:18–24.
- 111) Mali AV, Padhye SB, Anant S, Hegde MV, Kadam SS. Anticancer and antimetastatic potential of enterolactone: Clinical, preclinical and mechanistic perspectives. *Eur J Pharmacol.* 2019; 852: 107-124.
- 112) Grayson K, Gregory E, Khan G, Guinn BA. Urine Biomarkers for the Early Detection of Ovarian Cancer - Are We There Yet?. *Biomark Cancer*. 2019; 11: 1179299X19830977.
- 113) Dochez et al. Biomarkers and algorithms for diagnosis of ovarian cancer: CA125, HE4, RMI and ROMA, a review. *J. Ovarian Res.* 2019; 12: 28.
- 114) Duffy MJ. Use of Biomarkers in Screening for Cancer. Adv. Exp. Med.. Biol. 2015; 867: 27–39.
- 115) Lin H, Pizer E, Morin PJ. A frequent deletion polymorphism on chromosome 22q13 identified by representational difference analysis of ovarian cancer. *Genomics.* 2000: 69; 391–394.
- 116) Goodman JE, Lavigne JA, Hengstler JG, Tanner B, Helzlsouer KJ and Yager JD. Catechol-O-methyltransferase polymorphism is not associated with ovarian cancer risk. *Cancer Epidemiol. Biomarkers Prev.* 2000: 9; 1373-1376.

- 117) Goodman MT, McDuffie K, Kolonel LN, Terada K, Donlon TA, Wilkens LR, Guo C, Le Marchand L. Case-control study of ovarian cancer and polymorphisms in genes involved in catecholestrogen formation and metabolism. *Cancer Epidemiol. Biomarkers Prev.* 2001; 10(3): 209-216.
- 118) Qiu J, Du Z, Liu J, Zhou Y, Liang F, Lü Q. Association between polymorphisms in estrogen metabolism genes and breast cancer development in Chinese women: A prospective case-control study. *Medicine (Baltimore)*. 2018; 97(47): e13337.
- 119) Hung R J, Boffetta P, Brennan P, Malaveille C, Gelatti U, Placidi D, Carta A, Hautefeuille A, Porru S. Genetic polymorphisms of MPO, COMT, MnSOD, NQO1, interactions with environmental exposures and bladder cancer risk. *Carcinogenesis*. 2004: 25; 973–978.
- 120) Tolba MF, Omar HA, Hersi F, Nunes ACF, Noreddin AM. The impact of Catechol-O-methyl transferase knockdown on the cell proliferation of hormone-responsive cancers. *Mol Cell Endocrinol*. 2019; 488: 79-88.
- 121) Holt SK, Rossing MA, Malone KE, et al. Ovarian cancer risk and polymorphisms involved in estrogen catabolism. *Cancer Epidemiol Biomarkers Prev*.2007; 16: 481–489.
- 122) Sellers TA, Schildkraut JM, Pankratz VS, et al. Estrogen bioactivation, genetic polymorphisms, and ovarian cancer. *Cancer Epidemiol Biomarkers Prev*.2005; 14: 2536–2543.
- 123) Delort L, Chalabi N, Satih S, et al. Association between genetic polymorphisms and ovarian cancer risk. *Anticancer Res.2008;* 28: 3079–3081.
- 124) Pan W, Liao H. Correlations between the COMT gene rs4680 polymorphism and susceptibility to ovarian cancer. *Genetics and Molecular Research*. 2015; 14 (4): 16813-16818.
- 125) Garner EI, Stokes EE, Berkowitz RS, et al. Polymorphisms of the estrogenmetabolizing genes CYP17 and catechol-O-methyltransferase and risk of epithelial ovarian cancer. *Cancer Res.* 2002; 62: 3058–3062.
- 126) Wang YX, Zhu N, Zhang CJ, et al. Friend or foe: Multiple roles of adipose tissue in cancer formation and progression. Journal of Cellular Physiology Early View. *J. Cell Physiol.* 2019; 1–14.

### 7. APPENDICES

### 7.1. Forms

### 7.1.1. Biological Materials Transfer Form

### KLİNİK ARAŞTIRMALARDA KULLANILACAK BİYOLOJİK MATERYAL TRANSFER ANLAŞMASI

### Araştırmanın Açık Adı: Yumurtalık Kanserinde Katekol-O-Metiltransferaz (COMT) Geni Val158Met Polimorfizminin Araştırılması

#### Protokol Numarası :

#### Araştırmanın Özeti

Yumurtalık kanseri en yaygın kadın kanserlerinden biridir ve jinekolojik kanserler arasında önde gelen ölüm nedenidir. 205.000'in üzerinde yeni vaka ve 125.000 ölüm, yumurtalık kanserinden kaynaklanmaktadır. Etiyoloji açısından, östrojen katabolizmasından sorumlu genlerdeki polimorfizmlerin, hücresel genotoksik 4-hidroksile edilmiş katekol östrojenleri ve antianjiyojenik 2-metoksestradiol seviyelerini değiştirdiği ve böylece yumurtalık kanseri gelişme riskini etkilediği bilinmektedir. Katekol-O-metiltransferaz (COMT), östrojenin katekol metabolitinin bağlanması ve inaktivasyonunda yer alan önemli bir östrojen katalizleyen enzimdir. COMT; yumurtalık, göğüs, karaciğer ve böbrek dokuları dahil olmak üzere çeşitli insan dokularında yüksek miktarlarda bulunur. Katekol-O-metiltransferaz (COMT) geni 22q11.2 kromozomunda yer alır ve kodlanmayan ekson 1 ve 2 olmak üzere altı ekzondan oluşur. rs4680 polimorfizmi, ekson 4'te kodon 158'de bir G'den A'ya dönüşümüdür ve bu da valin yerine bir metiyonin değişimine yol açar; bu, azalan aktiviteye sahip bir termolabil enzim ile sonuçlanır. rs4680 ile yumurtalık kanserine karşı yatkınlık arasındaki ilişki üzerine birçok vaka kontrol çalışması yayınlanmıştır. Bununla birlikte, bu calışmalar araştırma kaliteşindeki yaryaşyonlar, küçük örnek boyutu veva deneklerin farklı bölgesel ve etnik altvapısı da dahil olmak üzere bazı sınırlamalara sahiptir. Bu nedenle, bu calısmalardan elde edilen sonuclar cok sağlam değildir ve sınırlı güvenilirliğe sahiptir. Ancak bu calısmanın sonucları, bu yıkıcı hastalığa yönelik temel arastırma ve klinik tedavi icin daha güvenilir kanıtlar sağlayacaktır.

Bu çalışmanın amacı, katekol-O-metiltransferaz (COMT) genindeki tek nükleotid polimorfizm rs4680 ile yumurtalık kanserine yatkınlık arasındaki korelasyonların sistematik incelemesinin yapılmasıdır. Yayımlanan ilgili vaka-kontrol çalışmaları için bir bilgisayar araştırması gerçekleştirilecektir.

İşbu anlaşma ile biyolojik materyali gönderen araştırmacı ve kurum "Yumurtalık Kanserinde Katekol-O-Metiltransferaz (COMT) Geni Val158Met Polimorfizminin Araştırılması" isimli araştırmada kullanılmak üzere gönderilecek "5ml ve araştırma" amacı ile kullanılması üzere 26 Ağustos Yerleşkesi Yeditepe Üniversitesi Kayışdağı / İSTANBUL adresindeki Yeditepe Üniversitesi Moleküler Tıp Anabilim Dalı ' ndaki merkezine göndermeden önce GÖNDERİCİ ve ALICI'dan aşağıdaki koşulları kabul etmesi istenmektedir;

- 1. Gönderilen biyolojik materyaller yalnızca yukarıda yazılı amaç için kullanılabilir. ALICI biyolojik materyallerin alınma amacından dışında ikincil amaç için kullanılma isteklerini GÖNDERİCİ'ye yazılı olarak bildirecektir.
- Biyolojik materyaller ALICI'ya gönderilmeden önce biyolojik materyalin sağlandığı gönüllülere ait Türkiye İlaç ve Tıbbi Cihaz Kurumu ve Etik Kurul'un onayladığı tüm kullanım amaçlarına yönelik olarak düzenlenmiş bilgilendirilmiş gönüllü olur formunun her bir gönüllüden alınmış olması gerekmektedir.
- 3. ALICI biyolojik materyali GÖNDERİCİ'nin yazılı izni olmadan üçüncü kişi/kurumlara vermeyecektir.

- 4. Biyolojik materyaller GÖNDERİCİ tarafından bireyin kimlik ve tanımlayıcı bilgileri olmaksızın ALICI'ya gönderilecektir.
- 5. ALICI biyolojik materyalleri Birleşmiş Milletler İnsan Genomu ve İnsan Hakları Evrensel Beyannamesine uygun olarak kullanacaktır.
- 6. Bu anlaşma ile gönderilecek biyolojik materyalin araştırma için kullanılacak olduğu ve biyolojik materyal kullanımına ait risklerin var olduğu ALICI tarafından kabul edilmektedir. Söz konusu risklere karşı uygun önlemlerin alınması gerekmektedir.
- 7. GÖNDERİCİ ve ALICI gönderilen biyolojik materyalin herhangi bir şekilde ticari kazanç kaynağı olarak kullanılamayacağı ancak elde edilebilecek fikri mülkiyet ve patent haklarının bu durumdan istisna olduğu kabul etmektedir. GÖNDERİCİ ve ALICI söz konusu haklarını araştırma başlangıcında karşılıklı olarak belirleyecektir.
- 8. ALICI bu anlaşmanın sonlanması veya biyolojik materyalin sağlandığı gönüllülere ait anlaşmanın 2. maddesinde belirtilen olurun geri çekilmesi halinde bütün materyalleri geri vermeyi veya ortadan kaldırmayı ve bunu belgelemeyi kabul eder.
- 9. Bu anlaşma, araştırmanın sonlanması, ilgili mevzuat hükümlerine uyulmaması veya ilgili tarafların anlaşma hükümlerine uymaması durumlarında son bulacaktır.
- **10.** Bu anlaşmanın yürütülmesinde ALICI ve GÖNDERİCİ yetkilileri sorumludur. Anlaşmazlık halinde ihtilafın çözümü için her iki ülke mahkemeleri de yetkilidir.

### BİYOLOJİK MATERYALİ GÖNDEREN ARAŞTIRMACI BİLGİSİ

Adı Soyadı ve Unvanı	ini : Prof. Dr. Rukset Attar				
Uzmanlık Alanı	: Kadın Hastalıkları ve Doğum				
Kurumu	: Yeditepe Üniversitesi Hastanesi				
Adresi	: İçerenköy Mah. Hastane Yolu Sok. 4,4/1				
	34718 Ataşehir / İstanbul				
Telefon	: 05378401900				
Faks					
E-posta	: rattar@yeditepe.edu.tr				
BİYOLOJİK MATERYALİ ALAN ALICI BİLGİSİ					
Adi Savadi ya Uniyani					

Adı Soyadı ve Unvanı	Prof. Dr. Turgay Isbir
Uzmanlık Alanı Kurumu Adresi	: Moleküler Tıp : Yeditepe Üniversitesi : Yeditepe Üniversitesi Moleküler Tıp Anabilim Dalı 26 Ağustos Yerleşkesi Kayışdağı
Telefon Faks E-posta	: 0533 282 37 26 : : turgay.isbir@yeditepe.edu.tr

Bu anlaşmada belirtilen koşulları okudum ve anladım. Gönderilen materyalde bu anlaşmada belirtilen koşullara uyacağımı taahhüt ederim.

		ALICI			
	Gönderen Araştırmacı	Gönderen Destekleyici Firma Yetkilisi veya Yasal Temsilcisi	Eğitim Görevlisi / Ana Bilim Dalı Başkanı	Kurum Amiri / Rektör veya Yetkilendirdiği Makam	Alıcı Kurum Yetkilisi
El Yazısı ile Adı Soyadı Unvanı					
Tarih					

İmza			

**Not:** Bu anlaşmada yer alan alıcı kurum yetkilisinin imzası yerine alıcı kurum tarafından verilecek olan ve içerik olarak bu anlaşmadaki hükümlere benzer hükümleri içeren imzalı "end use certificate" "son kullanım sertifikası" de kabul edilebilir.

#### AGREEMENT FOR TRANSFER OF BIOLOGICAL MATERIAL TO BE USED IN CLINICAL TRIALS

Full name of the clinical trial:

Protocol code

Summary of the clinical trial:

Click here on enter text

By this agreement, the investigator and the institution who send the biological material requires the CONSIGNEE and CONSIGNOR to agree on the below terms before sending (*Specify biological material type and amount*) which shall be used for to be dispatched to the address of .

- 1. Delivered biological materials shall be used only for the above-mentioned purposes. CONSIGNEE shall use those materials only for secondary purposes, which are initially approved by the CONSIGNOR in written.
- 2. Prior to the dispatch of the biological materials to the CONSIGNEE, Turkish Medicines and Medical Devices Agency and Ethics Committee approved informed consent forms, which belong to the persons for whom the biological material is provided, should be obtained. This consent form should explain all the purposes of use of the biological samples.
- 3. CONSIGNEE cannot provide the biological material to the third parties without prior written approval of the CONSIGNOR.
- 4. Biological materials shall be dispatched by the CONSIGNOR to the CONSIGNEE without the identity or any descriptive information of the individuals.
- 5. CONSIGNEE shall use the biological materials in accordance with as the United Nations Human Genome and Universal Declaration of Human Rights.
- 6. CONSIGNEE acknowledges and agrees that the biological materials to be dispatched under this agreement shall be utilized for research purposes and have some risks associated with their usage. Appropriate preventive actions should be taken for those risks.
- 7. CONSIGNOR and CONSIGNEE shall mutually agree that the biological materials cannot be used as a source for any commercial profit and the rights relating to a joint publication or a patent right that may arise may be the only exception for that. CONSIGNOR and CONSIGNEE shall mutually agree on those rights prior to trial initiation.
- 8. CONSIGNEE agrees to return or dispose of all materials and to evidence such acts accordingly in the event of termination of the agreement or withdrawal of written consent of the volunteer referred in Item 2.
- 9. This agreement shall be terminated in the event of, termination of the trial, violation on the terms of related regulations or noncompliance with agreement clauses of either of the parties.
- **10.** CONSIGNEE and CONSIGNOR shall be responsible from the execution of this Agreement and performances hereunder. In case of conflict, both countries of the parties' courts are authorized.

#### INFORMATION REGARDING THE INVESTIGATOR SENDING THE BIOLOGICAL MATERIAL

:

Name Surname and Title
Specialization
Institution
Address
Telephone
Fax
E-mail

INFORMATION REGARDING THE CONSIGNEE RECEIVING THE BIOLOGICAL MATERIAL

Name Surname and Title : Specialization :

Institution	:
Address	:
Telephone	:
Fax	:
E-mail	:

# I read and understood the terms under this agreement. I hereby agree and undertake that I will act in accordance with the terms of this agreement with respect to the dispatched materials.

		CONSIGNEE			
	Consignor Investigator	Consignor Sponsor Company Official or Legal Representative	Chief / Head of the Department	Chief Officer of the Institution / Rector or Assigned Person	Consignee Institution Official
Name Surname and Title in Handwriting					
Date					
Signature					

**Note:** Instead of the signature of the consignee representative, a signed "end use certificate" including clauses similar to this agreement's to be issued by the consignee institution may also be accepted.

# 7.1.2. Volunteer Form

# Asgari Bilgilendirilmiş Gönüllü Olur Formu

# Sayın Hastamız,

- Bu belge bilgilendirilme ve aydınlatılmış onam haklarınızdan yararlanabilmenizi amaçlamaktadır.
- Size gerçekleştirilebilecek klinik araştırmalar amaçlı girişimler konusunda, tüm seçenekler ile bu girişimlerin yarar ve muhtemel zararları konusunda anlayabileceğiniz şekilde **bilgi alma hakkınız ve bir kopyasını isteme hakkınız** vardır.
- Yasal ve tıbbi zorunluluk taşıyan durumlar dışında **bilgilendirmeyi reddedebilirsiniz**. Yazılı bildirmek koşulu ile bilgi almama veya yerinize güvendiğiniz bir kimsenin bilgilendirilmesini talep etme hakkına sahipsiniz.
- klinik araştırmalara katılım konusunda bilgilendirildikten sonra bunu kabul edebilirsiniz. Ya da k**arar verebilmek için uygun zaman talep edebilirsiniz.**
- Hayatınız veya hayati organlarınız tehlikede olmadığı sürece onamınızı (yazılı talep etme koşulu ile) **dilediğiniz zaman geri alabilir** ya da önceden kabul etmediğiniz herhangi bir tanı/tedavi amaçlı girişimi **tekrar talep edebilirsiniz**.
- Hastanemizde verilen hizmetleri **Hastane Tanıtım Broşürü**nden edinebilirsiniz. Ayrıca Hastane personeli hakkında <u>http://www.yeditepe.edu.tr</u> web sayfasından daha detaylı bilgilere ulaşabilirsiniz.
- Burada belirtilenlerden başka sorularınız varsa bunları yanıtlamak görevimizdir.

# TANIMLAMA

# 1. Araştırmanın Adı: Yumurtalık Kanserinde Katekol-O-Metiltransferaz (COMT) Geni Val158Met Polimorfizminin Araştırılması

# Araştırmaya Katılımcı Sayısı: 100

# Bu araştırmanın Amacı:

Yumurtalık kanseri en yaygın kadın kanserlerinden biridir ve jinekolojik kanserler arasında önde gelen ölüm nedenidir. 205.000'in üzerinde yeni vaka ve 125.000 ölüm, vumurtalık kanserinden kaynaklanmaktadır. Etiyoloji açısından, östrojen katabolizmasından sorumlu genlerdeki polimorfizmlerin, hücresel genotoksik 4hidroksile edilmiş katekol östrojenleri ve antianjiyojenik 2-metoksestradiol seviyelerini değiştirdiği ve böylece yumurtalık kanseri gelişme rişkini etkilediği bilinmektedir. Katekol-O-metiltransferaz (COMT), östrojenin katekol metabolitinin bağlanması ve inaktivasyonunda yer alan önemli bir östrojen katalizleyen enzimdir. COMT; yumurtalık, göğüs, karaciğer ve böbrek dokuları dahil olmak üzere çeşitli insan dokularında yüksek miktarlarda bulunur. Katekol-O-metiltransferaz (COMT) geni 22q11.2 kromozomunda yer alır ve kodlanmayan ekson 1 ve 2 olmak üzere altı ekzondan oluşur. rs4680 polimorfizmi, ekson 4'te kodon 158'de bir G'den A'ya dönüşümüdür ve bu da valin yerine bir metiyonin değişimine yol açar; bu, azalan aktiviteye sahip bir termolabil enzim ile sonuçlanır. rs4680 ile yumurtalık kanserine karşı yatkınlık arasındaki ilişki üzerine birçok vaka kontrol çalışması yayınlanmıştır. Bununla birlikte, bu çalışmalar araştırma kalitesindeki varyasyonlar, küçük örnek boyutu veya deneklerin farklı bölgesel ve etnik altyapısı da dahil olmak üzere bazı sınırlamalara sahiptir. Bu nedenle, bu çalışmalardan elde edilen sonuçlar çok sağlam değildir ve sınırlı güvenilirliğe sahiptir. Ancak bu çalışmanın sonuçları, bu yıkıcı hastalığa yönelik temel araştırma ve klinik tedavi için daha güvenilir kanıtlar sağlayacaktır. Bu çalışmanın amacı, katekol-O-metiltransferaz (COMT) genindeki tek nükleotid polimorfizm rs4680 ile yumurtalık kanserine yatkınlık arasındaki korelasyonların sistematik incelemesinin yapılmasıdır. İlgili vaka-kontrol çalışmaları için bir bilgisayar araştırması gerçekleştirilecektir.

# Süresi: 1 Yıl

# İzlenecek Yöntem/Yöntemler:

Hasta (n=) ve kontrol (n=) grubu olmak üzere iki gruptan oluşması planlanmıştır.

# **DNA İzolasyonu:**

Yumurtalık kanseri olduğu tespit edilen hastalardan 5ml EDTA 'lı tüplere yaklaşık 350 ml 'lik periferik kan alınacaktır.

DNA izolasyonu için iPrep DNA ( iPrep Pure link , invitrogen ) izolasyon robotu kullanılacaktır.

Elde edilen DNA ' ların konsantrasyonları nanodrop ( Thermoscientific ) cihazı ile belirlenecektir.

Saflık ölçümleri için NanoDrop cihazı ile spektrofotometrik olarak ölçüm yapılacaktır. DNA 'lar çalışma gününe kadar +4°C 'de bekletilecektir.

# Genotip Analizi :

COMT genotiplemesi Applied bioscience 7500 Fast Real Time cihazı kullanılarak

Gerçek zamanlı PZR ile yapılacak ve hedef varyasyonlarındaki olası mutasyonların

belirlenebilmesi için özel primer dizileri kullanılacaktır.

Çalışmanın istatistiksel analizi için SPSS 24.0 sürümü kullanılacaktır.

Hasta grubu ve kontrol grubu katılımcıların karşılaştırılmalarında allel fraksiyonlarının ve genotiplerin karşılaştırılması için ki kare testi kullanılacaktır.

Hasta ve kontrol grubu anlamlılık ölçümü için student 's t test kullanılacaktır. Anlamlılık seviyesi p < 0.05 kabul edilecektir.

#### Araştırma Sonunda Beklenen Fayda

Yumurtalık Kanserinde Katekol-O-Metiltransferaz (COMT) Geni Val158Met Polimorfizminin Etkisinin Belirlenmesi

Bu Çalışmada Herhangi Bir Alternatif Tedavi ya da Girişimde Bulunulmayacaktır.

Bu Araştırma Gönüllüler İçin Hiçbir Risk Teşkil Etmemekte ve Hiçbir Rahatsızlığa Sebep olmamaktadır.

#### ONAM (RIZA)

Bilgilendirilmiş Gönüllü Olur Formundaki tüm açıklamaları okudum. Bana, yukarıda konusu ve amacı belirtilen araştırma ile ilgili yazılı ve sözlü açıklama aşağıda adı belirtilen hekim tarafından yapıldı. Araştırmaya gönüllü olarak katıldığımı, istediğim zaman gerekçeli veya gerekçesiz olarak araştırmadan ayrılabileceğimi ve kendi isteğime bakılmaksızın araştırmacı tarafından araştırma dışı bırakılabileceğimi biliyorum. Bu durumda hastanenin çalışma düzeni ve hastalara verilen bakımda aksaklık olmayacağı konusunda bilgilendirildim. Bu araştırmaya katılırken zorlama, maddi çıkar ve ast üst ilişkisine dayalı herhangi bir baskı olmaksızın bu çalışmaya katıldığımı beyan ederim. Bu bilimsel çalışmanın devamı esnasındaki süreçle ilgili olarak ayrıca eklenen çalışma protokolü ile bilgilendirildim. Tarafımdan alınan kan ve doku örneklerinin daha sonra başka araştırma çalışmalarında kullanılmasında herhangi bir sakınca yoktur.

Söz konusu araştırmaya, hiçbir baskı ve zorlama olmaksızın kendi rızamla katılmayı kabul ediyorum.

#### Gönüllünün:

Adı Soyadı:

İmzası:

Tarih:

#### Açıklamaları Yapan Kişinin:

Adı Soyadı:

İmzası:

Tarih:

## Gerekiyorsa Olur İşlemine Tanık Olan Kişinin:

Adı Soyadı:

İmzası:

Tarih:

## Gerekiyorsa Yasal Temsilcinin :

Adı Soyadı:

İmzası:

Tarih:

# Klinik Araştırma Proje Koordinatörü İletişim Bilgileri:

Adı Soyadı: Uzmanlık alanı: Kurumu: E-posta adresi: Telefon numarası:

### 7.1.3. Case Report Form

### 1- ÇALIŞMANI ADI: Yumurtalık Kanserinde Katekol-O-Metiltransferaz (COMT) Geni Val158Met Polimorfizminin Araştırılması

ÇALIŞMAYA / ARAŞTIRMAYA DAHİL <u>EDİLME</u> KRİTERLERİ

Deney Grupları için;

- Gönüllü Olma
- Yumurtalık Kanseri Hastası Olma
- 18 85 yaş aralığında olma
- Yukarıda belirtilenler haricinde bir hastalığa sahip olmama

Kontrol Grubu için ;

- Gönüllü olma
- Sağlıklı olma (yukarıda belirtilenlerde dahil olmak üzere hiçbir hastalığa sahip olmama)
- 18 85 yaş aralığında olma

### ÇALIŞMAYA /ARAŞTIRMAYA DAHİL EDİLMEME KRİTERLERİ

- Gönüllü olmama
- 18 85 yaş aralığı dışında olma
- Yukarıda belirtilenler dışında bir hastalığa sahip olma

Sorumlu Araştırmacı



Sayı : 37068608-6100-15-1560 Konu: Klinik Araştırmalar Etik kurul Başvurusu hk.

25/10/2018

#### İlgili Makama (İpek Yağmur Abaoğlu)

Yeditepe Üniversitesi Biyokimya, Moleküler Tıp Anabilim Dalı Prof. Dr. Turgay İsbir'in koordinatör, Yeditepe Üniversitesi Hastanesi Kadın Hastalıkları ve Doğum Anabilim Dalı Prof. Dr. Rukset Attar'ın sorumlu olduğu "Yumurtalık Kanserinde Katekol-O-Metiltransferaz (COMT) Geni Vall58Met Polimorfizminin Araştırılması" isimli araştırma projesine ait Klinik Araştırmalar Etik Kurulu (KAEK) Başvuru Dosyası (1532 kayıt Numaralı KAEK Başvuru Dosyası ), Yeditepe Üniversitesi Klinik Araştırmalar Etik Kurulu tarafından 24.10.2018 tarihli toplantıda incelenmiştir.

Kurul tarafından yapılan inceleme sonucu, yukarıdaki isimi belirtilen çalışmanın yapılmasının etik ve bilimsel açıdan uygun olduğuna karar verilmiştir ( KAEK Karar No: 915 ).

Prof. Dr. Turgay ÇELİK Yeditepe Üniversitesi Klinik Araştırmalar Etik Kurulu Başkanı

Yeditepe Üniversitesi 26 Ağustos Yerleşimi, İnönü Mahallesi Kayışdağı Caddesi 34755 Ataşehir / İstanbul T. 0216 578 00 00 www.**yeditepe**.edu.tr **F.** 0216 578 02 99

## 8. RAW DATA

# COMTgeneotype \* patient\_control\_group

### Crosstab

			hasta_kontr	ol_grubu	
			kontrol	over ca	Total
COMTgeneotype	GG	Count	12	17	29
		% within COMTgeneotype	41,4%	58,6%	100,0%
		% within hasta_kontrol_grubu	25,5%	37,8%	31,5%
		% of Total	13,0%	18,5%	31,5%
	GA	Count	22	19	41
		% within COMTgeneotype	53,7%	46,3%	100,0%
		% within hasta_kontrol_grubu	46,8%	42,2%	44,6%
		% of Total	23,9%	20,7%	44,6%
	AA	Count	13	9	22
		% within COMTgeneotype	59,1%	40,9%	100,0%
		% within hasta_kontrol_grubu	27,7%	20,0%	23,9%
		% of Total	14,1%	9,8%	23,9%
Total		Count	47	45	92
		% within COMTgeneotype	51,1%	48,9%	100,0%
		% within hasta_kontrol_grubu	100,0%	100,0%	100,0%
		% of Total	51,1%	48,9%	100,0%

			Asymptotic		
			Significance (2-		Exact Sig. (1-
	Value	df	sided)	Exact Sig. (2-sided)	sided)
Pearson Chi-Square	1,597ª	1	,206		
Continuity Correction <sup>b</sup>	1,080	1	,299		
Likelihood Ratio	1,602	1	,206		
Fisher's Exact Test				,263	,149
Linear-by-Linear Association	1,580	1	,209		
N of Valid Cases	92				

# COMTgeneotypeGG \* patient\_control\_group

## Crosstab

			hasta_kont	rol_grubu	
			kontrol	over ca	Total
COMTgentypeGG	yok	Count	35	28	63
		% within COMTgentypeGG	55,6%	44,4%	100,0%
		% within hasta_kontrol_grubu	74,5%	62,2%	68,5%
		% of Total	38,0%	30,4%	68,5%
	var	Count	12	17	29
		% within COMTgentypeGG	41,4%	58,6%	100,0%
		% within hasta_kontrol_grubu	25,5%	37,8%	31,5%
		% of Total	13,0%	18,5%	31,5%
Total		Count	47	45	92
		% within COMTgentypeGG	51,1%	48,9%	100,0%
		% within hasta_kontrol_grubu	100,0%	100,0%	100,0%
		% of Total	51,1%	48,9%	100,0%

	Value	df	Asymptotic Significance (2- sided)	Exact Sig. (2-sided)	Exact Sig. (1- sided)
Pearson Chi-Square	1,597ª	1	,206		
Continuity Correction <sup>b</sup>	1,080	1	,299		
Likelihood Ratio	1,602	1	,206		
Fisher's Exact Test				,263	,149
Linear-by-Linear Association	1,580	1	,209		
N of Valid Cases	92				

# COMTgenotypeGA \* patient\_control\_group

			hasta_kon	trol_grubu	
			kontrol	over ca	Total
COMTgenotypeGA	yok	Count	25	26	51
		% within COMTgenotypeGA	49,0%	51,0%	100,0%
		% within hasta_kontrol_grubu	53,2%	57,8%	55,4%
		% of Total	27,2%	28,3%	55,4%
	var	Count	22	19	41
		% within COMTgenotypeGA	53,7%	46,3%	100,0%
		% within hasta_kontrol_grubu	46,8%	42,2%	44,6%
		% of Total	23,9%	20,7%	44,6%
Total		Count	47	45	92
		% within COMTgenotypeGA	51,1%	48,9%	100,0%
		% within hasta_kontrol_grubu	100,0%	100,0%	100,0%
		% of Total	51,1%	48,9%	100,0%

#### Crosstab

	Value	df	Asymptotic Significance (2- sided)	Exact Sig. (2-sided)	Exact Sig. (1- sided)
Pearson Chi-Square	,196ª	1	,658		
Continuity Correction <sup>b</sup>	,054	1	,816		
Likelihood Ratio	,196	1	,658		
Fisher's Exact Test				,680	,408
Linear-by-Linear Association	,194	1	,660		
N of Valid Cases	92				

#### Crosstab

			hasta_kont	rol_grubu	
			kontrol	over ca	Total
COMTgenotypeAA	yok	Count	34	35	69
		% within COMTgenotypeAA	49,3%	50,7%	100,0%
		% within hasta_kontrol_grubu	72,3%	77,8%	75,0%
		% of Total	37,0%	38,0%	75,0%
	var	Count	13	10	23
		% within COMTgenotypeAA	56,5%	43,5%	100,0%
		% within hasta_kontrol_grubu	27,7%	22,2%	25,0%
		% of Total	14,1%	10,9%	25,0%
Total		Count	47	45	92
		% within COMTgenotypeAA	51,1%	48,9%	100,0%
		% within hasta_kontrol_grubu	100,0%	100,0%	100,0%
		% of Total	51,1%	48,9%	100,0%

COMTgenotypeAA \* patient\_control\_group

			Asymptotic		
			Significance (2-		Exact Sig.
	Value	df	sided)	Exact Sig. (2-sided)	(1-sided)
Pearson Chi-Square	,362ª	1	,547		
Continuity Correction <sup>b</sup>	,130	1	,718		
Likelihood Ratio	,363	1	,547		
Fisher's Exact Test				,633	,360
Linear-by-Linear Association	,359	1	,549		
N of Valid Cases	92				

# COMTalleleG \* patient\_control\_group

#### Crosstab

			hasta_kontrol_grubu		
			kontrol	over ca	Total
COMTallelG	yok	Count	13	9	22
		% within COMTallelG	59,1%	40,9%	100,0%
		% within hasta_kontrol_grubu	27,7%	20,0%	23,9%
		% of Total	14,1%	9,8%	23,9%
	var	Count	34	36	70
		% within COMTallelG	48,6%	51,4%	100,0%
		% within hasta_kontrol_grubu	72,3%	80,0%	76,1%
		% of Total	37,0%	39,1%	76,1%
Total		Count	47	45	92
		% within COMTallelG	51,1%	48,9%	100,0%
		% within hasta_kontrol_grubu	100,0%	100,0%	100,0%
		% of Total	51,1%	48,9%	100,0%

	Value	df	Asymptotic Significance (2- sided)	Exact Sig. (2-sided)	Exact Sig. (1-sided)
Pearson Chi-Square	,741ª	1	,389		
Continuity Correction <sup>b</sup>	,380	1	,538		
Likelihood Ratio	,745	1	,388		
Fisher's Exact Test				,467	,269
Linear-by-Linear Association	,733	1	,392		
N of Valid Cases	92				

## COMTalleleA \* patient\_control\_group

			hasta_kont	trol_grubu	
			kontrol	over ca	Total
COMTalleIA	yok	Count	13	17	30
		% within COMTallelA	43,3%	56,7%	100,0%
		% within hasta_kontrol_grubu	27,7%	37,8%	32,6%
		% of Total	14,1%	18,5%	32,6%
	var	Count	34	28	62
		% within COMTallelA	54,8%	45,2%	100,0%
		% within hasta_kontrol_grubu	72,3%	62,2%	67,4%
		% of Total	37,0%	30,4%	67,4%
Total		Count	47	45	92
		% within COMTallelA	51,1%	48,9%	100,0%
		% within hasta_kontrol_grubu	100,0%	100,0%	100,0%
		% of Total	51,1%	48,9%	100,0%

### Crosstab

## Chi-Square Tests

	Value	df	Asymptotic Significance (2- sided)	Exact Sig. (2-sided)	Exact Sig. (1- sided)
Pearson Chi-Square	,741ª	1	,389		
Continuity Correction <sup>b</sup>	,380	1	,538		
Likelihood Ratio	,745	1	,388		
Fisher's Exact Test				,467	,269
Linear-by-Linear Association	,733	1	,392		
N of Valid Cases	92				

Group Statistics

	hasta_kontrol_grubu	Ν	Mean	Std. Deviation	Std. Error Mean
yıl	over ca	47	54,87	12,555	1,831
	kontrol	47	51,11	12,863	1,876
kg/m2	over ca	39	29,3128	5,37255	,86030
	kontrol	47	23,0468	3,62233	,52837
m2	over ca	37	1,7727	,15493	,02547
	kontrol	47	1,6645	,14643	,02136
mg/dl akş	over ca	42	108,52	33,288	5,136
	kontrol	47	86,45	7,604	1,109

		Levene's Test for Equality of Variances		t-test for Equality of Mea	
		F	Sig.	t	df
yıl	Equal variances assumed	,125	,724	1,436	92
	Equal variances not assumed			1,436	91,946
kg/m2	Equal variances assumed	5,945	,017	6,430	84
	Equal variances not assumed			6,206	64,494
m2	Equal variances assumed	,093	,761	3,278	82
	Equal variances not assumed			3,256	75,295
mg/dl akş	Equal variances assumed	15,691	,000	4,422	87
	Equal variances not assumed			4,201	44,826

#### Independent Samples Test

#### Independent Samples Test

		t-test for Equality of means			
		Sig. (2-tailed)	Mean Difference	Std. Error	
		Sig. (2-tailed)	Mean Difference	Difference	
yıl	Equal variances assumed	,154	3,766	2,622	
	Equal variances not assumed	,154	3,766	2,622	
kg/m2	Equal variances assumed	,000	6,26601	,97456	
	Equal variances not assumed	,000	6,26601	1,00960	
m2	Equal variances assumed	,002	,10823	,03302	
	Equal variances not assumed	,002	,10823	,03324	
mg/dl akş	Equal variances assumed	,000	22,077	4,992	
	Equal variances not assumed	,000	22,077	5,255	

### t-test for Equality of Means

#### Independent Samples Test

#### t-test for Equality of Means

#### 95% Confidence Interval of the Difference

		Lower	Upper
yıl	Equal variances assumed	-1,441	8,973
	Equal variances not assumed	-1,441	8,973
kg/m2	Equal variances assumed	4,32800	8,20402
	Equal variances not assumed	4,24941	8,28262
m2	Equal variances assumed	,04255	,17391
	Equal variances not assumed	,04202	,17445
mg/dl akş	Equal variances assumed	12,155	31,999
	Equal variances not assumed	11,492	32,662

			hasta_kont	rol_grubu		
			kontrol	over ca	Total	diab
değişenmenapoz	premenapoz	Count	28	9	37	etes
		% within değişenmenapoz	75,7%	24,3%	100,0%	*
		% within hasta_kontrol_grubu	59,6%	19,1%	39,4%	pati
		% of Total	29,8%	9,6%	39,4%	ent_
	postmenapoz	Count	19	38	57	cont rol_
		% within değişenmenapoz	33,3%	66,7%	100,0%	gro
		% within hasta_kontrol_grubu	40,4%	80,9%	60,6%	up
		% of Total	20,2%	40,4%	60,6%	-
Total		Count	47	47	94	
		% within değişenmenapoz	50,0%	50,0%	100,0%	
		% within hasta_kontrol_grubu	100,0%	100,0%	100,0%	
		% of Total	50,0%	50,0%	100,0%	

## $changing menopause \ ^* \ patient\_control\_group \ Crosstabulation$

#### Crosstab

		hasta_kontrol_grubu			
			kontrol	over ca	Total
diyabet	yok	Count	47	33	80
		% within diyabet	58,8%	41,3%	100,0%
		% within hasta_kontrol_grubu	100,0%	73,3%	87,0%
		% of Total	51,1%	35,9%	87,0%
	var	Count	0	12	12
		% within diyabet	0,0%	100,0%	100,0%
		% within hasta_kontrol_grubu	0,0%	26,7%	13,0%
		% of Total	0,0%	13,0%	13,0%
Total		Count	47	45	92
		% within diyabet	51,1%	48,9%	100,0%
		% within hasta_kontrol_grubu	100,0%	100,0%	100,0%
		% of Total	51,1%	48,9%	100,0%

smoking \* patient\_control\_group

			hasta_kontrol_grubu		
			kontrol	over ca	Total
sigara_kullanımı	kullanmıyor	Count	26	37	63
		% within sigara_kullanımı	41,3%	58,7%	100,0%
		% within hasta_kontrol_grubu	55,3%	82,2%	68,5%
		% of Total	28,3%	40,2%	68,5%
	kullanıyor	Count	21	8	29
		% within sigara_kullanımı	72,4%	27,6%	100,0%
		% within hasta_kontrol_grubu	44,7%	17,8%	31,5%
		% of Total	22,8%	8,7%	31,5%
Total		Count	47	45	92
		% within sigara_kullanımı	51,1%	48,9%	100,0%
		% within hasta_kontrol_grubu	100,0%	100,0%	100,0%
		% of Total	51,1%	48,9%	100,0%

#### Crosstab

## 9. CURRICULUM VITAE

## **Personal Information**

Name	Ipek Yagmur	Surname	ABAOGLU
Birth Place	Istanbul	Date of Birth	23/01/1995
Nationality	Turkish	Identity Number	
E-mail	yagmurabaoglu@gmail.com	Phone	

### Education

Degree	Department	Institution	Graduation Year
Doctorate			
Master			
University	Molecular Biology and Genetics	Yıldız Technical University	2017
High School	Science-Math	Hasan Sabriye Gümüş Anatolian High School	2013

## Scientific Work

## Additional Education & Certificates

Thermofisher Scientific Days 2018 Workshop, September 2	.018
indimonster Scientific Dujs 2010 Workshop, September 2	010

## **Computer Knowledge**

Program	Usage
Office Programs	Very good
Statistical Package for the Social Sciences (SPSS)	Good

