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UNIVERSITY OF DICLE
INSTITUTE OF NATURAL AND APPLIED SCIENCES**

**SCREENING OF ANGUCYCLINE ANTIBIOTICS AS POTENTIAL
DRUG CANDIDATES AGAINST MRSA BY DOCKING ANALYSIS**

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**ANGUSİKLİN ANTİBİYOTİKLERİN DOKİNG ANALİZLERİ
YARDIMIYLA MRSA'YA KARŞI POTANSİYEL İLAÇ ADAYI
OLARAK TARANMASI**

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YÜKSEK LİSANS TEZİ

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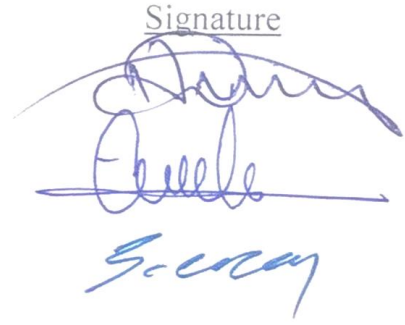
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Screening of Angucycline Antibiotics as Potential Drug Candidates Against MRSA by Docking Analysis, Submitted by Hazem Abbas Tofiq AL-BUSTANY in partial fulfillment of the requirements for the degree of Master of Science in Biology/Molecular Biology.

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CONTENTS

	Page
Acknowledgement	I
Contents	II
Abstract in Turkish (OZET)	IV
Abstract	V
List of Tables	VI
List of Figures	VII
List of Appendix	IX
Abbreviations	X
1. INTRODUCTION	1
2. LITERATURE REVIEW	5
2.1. Current Status of MRSA	5
2.2. Treatment approaches for MRSA	7
2.3. Targeted pathway	9
2.4. Secondary Metabolites	11
2.4.1. Function and Importance of Secondary Metabolites	12
2.4.2. Genetic of Secondary Metabolites	14
2.5. Angucycline Compounds	15
2.6. Actinomycetes, especially genus streptomyces (Angucycline producers)	17
2.7. Protein (Receptor) Structure & Function	19
2.7.1. Introduction to Proteins	19
2.7.2. Protein Structure	21
2.7.3. Protein Data Bank	22
2.8. Ligands	23
2.9. Protein-Ligand Interaction	23
2.9.1. Non-covalent binding	24
2.9.2. Electrostatic Interaction	24
2.9.3. Hydrophobic Interaction	25
2.9.4. Binding Affinity and Binding Site	26
2.9.5. Affinity prediction problem	27

2.10. Molecular Docking	27
2.10.1. AutoDock Vina	28
3. MATERIALS AND METHODS	31
4. RESULTS AND DISCUSSION	39
5. CONCLUSION	65
6. REFERENCES	67
APPENDIX I	75
APPENDIX II	90
APPENDIX III	105
APPENDIX IV	119

ÖZET

ANGUSİKLİN ANTİBİYOTİKLERİN DOKİNG ANALİZLERİ YARDIMIYLA MRSA'YA KARŞI POTANSİYEL İLAÇ ADAYI OLARAK TARANMASI

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Metisilin-dirençli *Staphylococcus aureus* (MRSA), ağırlıklı hastane enfeksiyonlarından kaynak alan önemli bir patojendir. β -laktamlar, aminoglikozitler ve kinolonlar gibi farklı antibiyotiklere direnç geliştirmişlerdir. Bundan dolayı, MRSA enfeksiyonlarına karşı, bunların virulans faktörlerini hedef alan yeni ilaçların keşfine büyük ihtiyaç vardır. Stafiloksantin MRSA'nın bir virulans faktörü olup, bu pigmentin biyosentezinin ilk aşaması dehidroskualen sentaz (CrtM) enzimi tarafından sentezlenmektedir. Bu sebeple, CrtM, virulansı zayıflatmak suretiyle MRSA'ya karşı kullanılabilir. Aktinomisetler; Gram-pozitif, filamentli bakteriler olup, farklı biyolojik aktivitelere sahip sekonder metabolitleri yüksek kapasitede üretirler. Bunlar arasında, polisiklik aromatik bileşikler olan ve tip-II poliketid sentazlar tarafından sentezlenen angusiklin antibiyotikler geniş bir yer kaplamaktadır. Bu çalışma, aktinomisetler tarafından üretilen 157 angusiklin bileşiğinin, AutoDock Vina programı kullanılarak, MRSA CrtM enzimi (PDB ID: 3ACW and 3W7F) üzerine inhibisyon etkisinin değerlendirilmesini kapsamaktadır. Doking analizleri sonucunda, incelenen bileşikler içerisinde; Moromycin A (56), Saquayamycin B (58), Saquayamycin A (145) ve Saprolymycin E (29) bileşiklerinin, enzimin substratı olan farnesyl diphosphate (doking skoru -8.3 kcal/mol) ile kıyaslandığında, CrtM ile iyi etkileşim göstererek, sırasıyla; -14.8, -14.4, -13.7 ve -13.7 kcal/mol gibi yüksek doking skorlarına sahip olduğu tespit edildi. Bununla beraber, kesin sonuçların elde edilmesi için moleküler dinamik simülasyonları ve in vitro deneylerin yapılması gerekmektedir..

Anahtar Kelimeler : Aktinomisetler, Angusiklin Antibiyotikler, MRSA, Stafiloksantin, CrtM, Moleküler Docking, AutoDock Vina.

ABSTRACT

SCREENING OF ANGUICYCLINE ANTIBIOTICS AS POTENTIAL DRUG CANDIDATES AGAINST MRSA BY DOCKING ANALYSIS

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Methicillin-resistant *Staphylococcus aureus* (MRSA) is one of the major pathogens, mainly caused by hospital infections. It has also developed resistance to various antibiotics such as β -lactams, aminoglycosides, and quinolones. Therefore, it is necessary to discover new drugs against MRSA infections by targeting their virulent factors. It is through that staphyloxanthin is a virulent factor of MRSA as dehydrosqualene synthase (CrtM) involves in the first step of its biosynthesis. For this reason, the CrtM enzyme is a potential target against MRSA by weakening its virulence. Actinomycetes are Gram-positive, filamentous, bacteria known for their significant capacity for the production of secondary metabolites with diverse biological activities. Among these, the polycyclic aromatic compounds which are known as angucycline antibiotics are the largest group of type-II polyketide synthase. The present study involves the evaluation of the inhibitory activity of 157 actinomycete-produced angucycline compounds against MRSA CrtM enzyme (PDB ID: 3ACW and 3W7F) by docking studies. Docking analysis demonstrate that among the attempted compounds; Moromycin A (56), Saquayamycin B (58), Saquayamycin A (145) and Saprolmycin E (29) have good interactions with CrtM with higher dock scores; -14.8, -14.4, -13.7, and -13.7 kcal/mol, respectively, when compared with substrate farnesyl diphosphate (-8.3 kcal/mol) and one of current inhibitors BPH-651 (-11.5 kcal/mol). However further studies, molecular dynamic simulations and in vitro investigations are required to achieve a conclusion.

Key Words: Actinomycetes, Angucycline Antibiotics, MRSA, Staphyloxanthin, CrtM, Molecular Docking, AutoDock Vina.

LIST OF TABLES

Table No.	Description	Page
Table 3.1.	Angucycline Groups Antibiotics and their references	31
Table 4.1.	Average and maximum dock scores of ligands docked to CrtM (3ACW)	44
Table 4.2.	Average of three dock score with maximum dock score of ligands docked to (3W7F) protein with MG	48
Table 4.3.	Average of three dock score with maximum dock score of ligands docked to (3W7F) protein without MG.	56

LIST OF FIGURES

Figure No.	Description	Page
Figure 2.1.	Biosynthetic pathways. Staphyloxanthin biosynthesis in <i>S. aureus</i>	10
Figure 2.2.	Tetrangomycin structure	16
Figure 2.3.	The planar peptide group	22
Figure 2.4.	Major types of non-bonded interactions in protein-ligand complexes	25
Figure 2.5.	Aryl-aryl interactions in protein structure and protein-ligand complexes. A. Edge-to-face geometry, B. Parallel stacking geometry	26
Figure 4.1.	Superimpose structure of docked inhibitor with its original BPH-651 coordinates in the complex with CrtM	39
Figure 4.2.	Superimpose structure of docked substrate with its original farnesyl diphosphate coordinates in the complex with CrtM	40
Figure 4.3.	Superimpose structure of docked inhibitor BPH-830 with original farnesyl diphosphate coordinates in the complex with CrtM	41
Figure 4.4.	Structures of (a) The tetracyclic benz[a]anthracene frame; (b) Moromycin A (56); (c) Saquayamycin B (58); (d) Saquayamycin A (145); and (e) Saprolmycin E (29)	42
Figure 4.5.	Group 1 ligands (28, 29, 155) docked to (3ACW) protein	45
Figure 4.6.	Group 2 ligands (47, 49, 84, 88, 102) docked to (3ACW) protein	45
Figure 4.7.	Group 3 ligands (92, 93, 95, 98) docked to (3ACW) protein	46
Figure 4.8.	Group 4 ligands (37, 94) docked to (3ACW) protein	46
Figure 4.9.	Group 5 ligands (11, 54, 115, 118, 119) docked to (3ACW) protein	47
Figure 4.10.	Group 6 ligands (33, 40, 87, 133, 154) docked to (3ACW) protein	47
Figure 4.11.	Group 1 ligands (2, 13, 59, 98) docked to (3W7F) protein with MG.	50
Figure 4.12.	Group 2 ligands (19, 28, 29, 30, 38) docked to (3W7F) protein with MG.	51
Figure 4.13.	Group 3 ligands (41, 42, 69, 124) docked to (3W7F) protein with MG.	51
Figure 4.14.	Group 4 ligands (47, 49, 56, 58) docked to (3W7F) protein with MG.	52
Figure 4.15.	Group 5 ligands (65, 93, 123, 124, 134) docked to (3W7F) protein with MG.	52
Figure 4.16.	Group 6 ligands (84, 92, 145, 146, 147, 148, 151) docked to (3W7F) protein with MG.	53

Figure 4.17.	Group 7 ligands (6, 7, 11, 80, 81, 102, 116, 119, 120) docked to (3W7F) protein with MG.	53
Figure 4.18.	Group 8 ligands (31, 32, 33, 51, 62, 137) docked to (3W7F) protein with MG. ..	54
Figure 4.19.	Group 9 ligands (55, 107, 108, 109, 110) docked to (3W7F) protein with MG. ...	54
Figure 4.20.	Group 10 ligands (4, 34, 136, 154) docked to (3W7F) protein with MG.	55
Figure 4.21.	Group 11 ligands (86, 111, 115, 118) docked to (3W7F) protein with MG.	55
Figure 4.22.	Group 1 ligands (28, 29, 94, 141) docked to (3W7F) protein without MG.	58
Figure 4.23.	Group 2 ligands (38, 56, 134, 146, 147, 148, 151, 155) docked to (3W7F) protein without MG.	58
Figure 4.24.	Group 3 ligands (46, 47, 49) docked to (3W7F) protein without MG.	59
Figure 4.25.	Group 4 ligands (143_PI_085, 144_PI_087) docked to (3W7F) protein without MG.	59
Figure 4.26.	Group 5 ligands (34, 42) docked to (3W7F) protein without MG.	60
Figure 4.27.	Group 6 ligands (6, 7, 11, 80, 81, 88, 116, 119, 120) docked to (3W7F) protein without MG.	60
Figure 4.28.	Group 7 ligands (31, 32, 33, 51, 62, 82) docked to (3W7F) protein without MG.	61
Figure 4.29.	Group 8 ligands (107, 108, 109, 110) docked to (3W7F) protein without MG....	61
Figure 4.30.	Group 9 ligands (111, 115, 118) docked to (3W7F) protein without MG.....	62
Figure 4.31.	Group 10 ligands (137, 156, 157) docked to(3W7F) protein without MG.....	62

LIST OF APPENDIX

Appendix No.	Description	Page
APPENDIX I	Docking score and interacted residue of protein (3ACW) with the Angucycline Group Antibiotics (ligands), and the arrangements are according to dock scores (kcal/mol)	75
APPENDIX II	Docking score and interacted residue of protein (3W7F with MG) with the Angucycline Group Antibiotics (ligands), and the arrangements are according to dock scores (kcal/mol)	90
APPENDIX III	Docking score and interacted residue of protein (3W7F without MG) with the Angucycline Group Antibiotics (ligands), and the arrangements are according to dock scores (kcal/mol)	105
APPENDIX IV	The 2D Interactions Between Docked Angucycline Group Antibiotics (Ligands) with Proteins	119

ABBREVIATION

2D	: Two-Dimension
3D	: Three-Dimension
Å	: Angstrom
Ala	: Alanine
Arg	: Arginine
Asn	: Asparagine
Asp	: Aspartic Acid
AV.	: Average
CrtM	: Dehydrosqualine Synthase Enzyme
Cys	: Cysteine
Gln	: Glutamine
Glu	: Glutamic Acid
Gly	: Glycine
His	: Histidine
Ile	: Isoleucine
ITC	: Isothermal Titration Calorimetry
Leu	: Leucine
Lys	: Lysine
Max.	: Maximum
Met	: Methionine
MG.	: Magnesium
MGL Tools	: Molecular Graphics Laboratory Tools
MRSA	: Methicillin Resistant <i>Staphylococcus aureus</i>
MSSA	: Methicillin Sensitive <i>Staphylococcus aureus</i>
ORSA	: Oxacillin Resistant <i>Staphylococcus aureus</i>

PDB	: Protein Data Bank
PDBQT	: Protein Data Bank, Partial Charge (Q), & Atom Type (T)
Phe	: Phenylalanine
Pro	: Proline
QSAR	: Quantitative Structural Activity Relationship
RMSD	: Root Mean Square Deviation
<i>S. aureus</i>	: <i>Staphylococcus aureus</i>
Ser	: Serine
TCSs	: Two-component Systems
T.B.	: Tuberculosis
Thr	: Threonine
Trp	: Tryptophan
Tyr	: Tyrosine
Val	: Valine

1. INTRODUCTION

The discovery of penicillin in the 1940s revolved the fight against bacterial colony mainly *Staphylococcus aureus* infections. Shortly after its introduction isolates of penicillin-resistant *Staphylococcus aureus* were identified. Methicillin was first used in 1959 to treat these penicillin-resistant strains of *S. aureus* (Wilkinson 2010), and methicillin-resistant *S. aureus* (MRSA) were reported in the early 1960's first and are now viewed as a major hospital acquired pathogen worldwide (Batabyal *et al.* 2012). This infection is especially troublesome in hospitals, prisons and nursing homes, where patients with open wounds, invasive devices, and weakened immune systems are at greater risk of infection compared with the general public (Batabyal *et al.* 2012).

The swift rise of antimicrobial resistance among pathogens has led to a renewed interest to search for novel antimicrobial agents. The history of new drug discovery processes shows that novel skeletons have, in the majority of cases come from natural sources. They have been the source of, or inspiration for the development of chemical entities introduced as pharmaceutical. The evolution of microbial natural product collections and expansion of high-throughput screening methods have brought over researchers to use the natural product libraries in drug discoveries. Actinomycetes continue to be a productive and successful focus for natural products research, with many novel compounds with prominent pharmacological valuable (Adegboye and Babalola 2013).

Microbial natural products nowadays are the origin of most of the antibiotics in the market. There is an alarming scarcity of new antibiotics currently under development in the pharmaceutical industry. Microbial natural products still remain the most promising source of novel antibiotics, although new methods are required to improve the efficiency of the discovery process (Singh *et al.* 2014). The golden carotenoid pigment staphyloxanthin by *S. aureus* acts as a virulence factor, mainly by acting as a bacterial antioxidant which helps to the microbe avoid the reactive oxygen species which the host immune system uses to kill pathogens (Liu *et al.* 2008).

1. INTRODUCTION

In earlier times, secondary metabolites were defined as substances with a low molecular weight, which were not products of the primary metabolic pathway of the producing organism. As a matter of fact, it was thought that these products did not have a roll in the microbial primary functions or growth. Therefore, it was thought that production of secondary metabolites did not represent any advantage for the producing microorganism (Tabarez 2005).

Nowadays it is considered that cell investment in secondary metabolite production is almost the confirmation of a function that should give the organisms certain advantage against other members of the microbial community. In fact, secondary metabolites are accepted to be essential for the producing cell as inhibitors of other organisms that compete for the same food supply or as regulators of cellular differentiation processes. In addition, it is reported that they are indeed products of biosynthetic pathways, which have evolved to give these types of advantages (Tabarez 2005).

Actinomycetes are enthralling resource among microorganisms due to their capability to produce novel bioactive secondary metabolites with antimicrobial activities. They have proven to be effective antimicrobial agents, especially against pathogenic organisms (Adegboye and Babalola 2013). In 1940, Selman A. Waksman isolated an effective T.B. antibiotic, actinomycin and for this he got success in 1944, with the discovery of spectromycin (Demain 1998). Scientists have discovered that actinomycetes have an enormous potential to produce valuable natural products (Raczkowski 2010). Hence, they produce useful secondary metabolites of high commercial value and continue to be routinely screened for new bioactive compounds. These searches have reaches a success and approximately they provide two thirds of naturally occurring antibiotics, including many of medical importance, mainly producing over 70 % of the natural product scaffolds found in clinically used anti-infective agents (Bhat *et al.* 2013), (Gomez-Escribano *et al.* 2015). Consequently they remain essential source of new chemical diversity and main part of drug discovery. Their ingenuity and immense industrial value is extremely noteworthy (Adegboye and Babalola 2013).

There are a long time consuming and costly way in drug discovery process, computational methods attempts have been made to increase the efficiency of random screening to select a typical subset of compounds from a compound collection. This usually entails grouping (clustering) compounds with similar structure, and then choosing a few members from each cluster for screening (Silverman and Holladay 2014). Computational approaches have successfully been applied to narrow the time and cost involved in the process, with the quick increase in computational power, *in silico* methods became commonly used in the fields of structural molecular biology and structure-based drug design. Molecular docking is one of these computational techniques (Ryska 2011). Molecular docking is a computer-based, high-throughput screening method for identifying compounds of a certain structure or size (Ascencio 2010), Molecular docking may be defined as a problem of lock-and-key, where one is interested in finding the suitable orientation of a key (ligand) that will open the lock (protein) (Ryska 2011).

Knowledge about one or more known ligands or about the structure of the target itself may be used to narrow a large screening collection to a smaller set of compounds that may be more likely to hit the target. Virtual screening is the most common computational method for selection of the compounds, which involves the rapid *in silico* (by computer) assessment of large libraries of chemical structures to identify those structures that most likely bind to a drug target, such as enzyme or protein receptor. The goal is to identify new scaffolds, chiefly ones that may be in the existing collection. In computer-based analysis two components are needed: (1) a database of structures in a form that can be computationally analyzed for structural attributes and (2) a hypothesis or model of the structural attributes that are important for activity, for example, the hypothesis that structural similarities to a known active ligand should yield similarly active compounds or a hypothesis of the shape and charge density of a binding pocket that defines what features a complementary ligand structure should have (Silverman and Holladay 2014).

Furthermore, one of main tools for virtual screening procedures is docking, where a library of several compounds is “docked” against one drug target and returns the best hit. The procedure of virtual screening through docking has become essential when it is needed to test a database of thousands (or even millions) of compounds

1. INTRODUCTION

against one or more targets in a short period of time. This search would be impossible to be reproduced experimentally at a so small economic and time cost. For this reason docking has been found to be a beneficial step in Quantitative Structural Activity Relationship (QSAR) studies, where statistical analysis is applied to thousands of drug candidates (Novotarskyi, 2013).

Docking method predicts favored orientation of one molecule to the second when they bind to form a stable complex. In the field of drug design, first molecule is usually protein (macromolecule) and the second one is ligand (small organic molecule) which is potential drug candidate. Information of favored orientation of ligand and protein can be used to predict binding affinity, and this discriminating high-affinity drug candidate from the low-affinity compounds (Ryska 2011).

Usually scoring function used in docking programs in order to recreate the chemical potentials which predict the conformation of binding. Need superficial physics-based (Coulombs energy and van der Waals forces) experimentally weighted to account for the difference in energy and free energy. Usually protein (receptor) have to be prepared by adding hydrogens and charges select site and eliminate of water and cofactors included (Pétursson 2014). Improvements in structure determination methods along with quick advances in molecular visualization tools have led to the rise of structure-aided drug design or rational drug design as an integral part of the drug discovery and development process (Dias 2011). Using of computer based analysis of molecular interaction for example protein-ligand binding becomes more necessary with the availability and expanding of molecular biological data. For this purpose, docking algorithms included a reasonably accurate model of energy and the flexibility of molecules (Pétursson 2014).

Moreover docking calculations simulate the interactions between the protein's binding site and the ligand, and these interactions give a qualitative score, and therefore the outcomes may be compared to those of biochemical assays (Tunca 2012). As a consequence, hundreds of thousands of compounds can be screened by using in silico methods. In addition these computer programs can generate the crystal structure and NMR solution structures of the target or related proteins, as well as calculating atomic homology models. Candidate drug-binding pockets can be identified as well (Ascencio 2010).

2. LITERATURE REVIEW

2.1. Current Status of MRSA

Methicillin-resistant *Staphylococcus aureus* (MRSA) is a Gram positive non-motile, non-facultative coccus whose infections in humans. Best environmental conditions for growth are temperatures between 15°C and 45°C. High concentrations of sodium chloride do not change the growth, even when concentrations reach up to 15% (Stuczen 2013). Consequently the universal occurrence of MRSA has become problem for public health (Howe *et al.* 2004), (Moghadam *et al.* 2015). It is also known as multidrug-resistant *S. aureus* and oxacillin-resistant *S. aureus* (ORSA). *S. aureus* was first identified in the late 19th century and has since been recognised as part of the natural flora of humans. It frequents the face, hands and perineum, with the most common site being the nares (nostril) (Stuczen 2013). *S. aureus* species developing, resistance to beta-lactam antibiotics are known as MRSA, they include the penicillins like methicillin, dicloxacillin, nafcillin, oxacillin, etc. and the cephalosporins (Batabyal *et al.* 2012), (Rashid *et al.* 2015). This organism is part of the natural microbiota of humans, from which clones of epidemic drug-resistant *S. aureus* have emerged (Hsu *et al.* 2015). MRSA is reported as the leading cause of wound infections in most parts of the world (Stuczen 2013). Hospital acquired infections and infections in the community (Onelum *et al.* 2015). Between 30% and 60% of the healthy population carry *S. aureus*, of which between 10% and 20% are chronically colonized (ongoing, persistent population of *S. aureus* on or in the body but in the absence of infection) (Stuczen 2013). The appearance and spread of bacterial pathogens that have become modified for existence in hospitals poses a major threat to global health systems (Hsu *et al.* 2015).

The high prevalence of MRSA colonization in diabetic foot ulcers is a consequence of antibiotic overuse and the selection of broad rather than narrow spectrum agents (Stuczen 2013).

There is another classification which known as methicillin-sensitive *S. aureus* or MSSA if the strain unable to resist these antibiotics. The development of such resistance does not cause more intrinsically virulent for the organism than strains of *S. aureus* that

have no antibiotic resistance, but resistance does make MRSA infection more difficult to treat with typical types of antibiotics and thus more dangerous (Batabyal *et al.* 2012). The glycopeptides, particularly vancomycin, have been the mainstays of therapy for MRSA, and the emergence of resistance to these agents is of great concern (Howe *et al.* 2004).

People not involved in health care backgrounds are often less aware of this silent and lethal epidemic. This lack of awareness lies in significance of hazard, perils of hospital acquired MRSA infection, and potential risk to overall health care system (Rashid *et al.* 2015). Healthy people may carry MRSA asymptotically for long periods of time but patients with compromised immune system are at a significant greater risk of symptomatic infections (Onelum *et al.* 2015). A significant public behavioral alteration is needed in order to control this global risk as well as a well-informed public. (Rashid *et al.* 2015) The range of infections due to MRSA are manifold and are linked with worse outcome in addition to extended hospital stay, higher cost of treatment and increased mortality (Onelum *et al.* 2015).

The concept of “anti-infectious drugs” includes not only compounds that inhibit the growth of pathogenic microorganisms statically or kill them (so called chemotherapeutics or antibiotics) and vaccines but also compounds that control microbial adaptation/survival or pathogenicity, potentiate the activities of known antibiotics, or enhance the host immune system against microbial infection (Koyama *et al.* 2013). Production of β – lactamase enzyme in the affected area is the main cause of microbial resistance (Rashid *et al.* 2015). For example, β -lactamase inhibitors such as clavulanic acid, sulbactam, and tazobactam themselves show very weak or no antimicrobial (non-antibiotic) activity, but these compounds dramatically potentiate the antimicrobial activity of β -lactam antibiotics against β -lactamase-producing bacteria (Koyama *et al.* 2013). The choice of antibiotics as active treatment is reduced after the maturity of infection. Such methods are possibly show harmful side effects to the patient and expensive. Recent evidence supports that domestic animals like cat, dog and hen can transmit MRSA to their owners (Rashid *et al.* 2015).

In recent years, active anti-infectious compounds against MRSA have been widely searched for. Several compounds have been found to have new mechanisms of action against MRSA and are expected to be potential leads for the treatment of infection (Koyama *et al.* 2013). *S. aureus* is an extremely transmittable bacterial species found in the ecosystem. The microorganism invades the skin and enters deeper tissues. As in septicemia, it multiplies to cause a localized or systemic response (Rashid *et al.* 2015). MRSA produce a number of virulence factors which cause suppurative infections and toxinosis. Surface proteins allow bacterial attachment to the extracellular matrix of the host, specifically the proteins laminin and fibronectin, found in epithelial and endothelial tissue. Toxins produced by the bacteria damage host cell membranes and allow cell invasion. Alpha toxin is produced as a monomer that binds to the membrane of the susceptible cell. Sub-units then combine to form heptameric rings with a central pore, through which the cellular contents leak (Stuczen 2013). MRSA infections cause a huge number of deaths every year worldwide (Koyama *et al.* 2013). The patient becomes infected with the growth of its population. People who are weaker, older and sicker have weaker immune system and may get infected easily. It is also described that people may bring this infection without having any noticeable indications (Rashid *et al.* 2015).

2.2. Treatment approaches for MRSA

Microbiologists are observing an exponential growth in infectious human diseases through *S. aureus*, which is precisely known as Methicillin Sensitive *S. aureus* (MSSA). In this case, bacterial skin infections are because of the strains of Methicillin Resistant *S. aureus* (MRSA) (Rashid *et al.* 2015). According to drug bank only three antibiotics namely arbekacin, meticillin and linezolid, are approved for the treatment of MRSA. Drug targets are inadequate and there is an urgent need to find out the novel drug targets. Apart from drug target, there is also a need for good ligand preparation that would act as effective inhibitors to the novel targets without affecting human proteome. Any contaminated surfaces is the source of several kind of infections, MRSA is one of them (Balaji SR *et al.* 2014). As observed in the last decade the microbe has latency to severely resist antibiotics. The human race is facing significant morbidity caused by these lethal infections (Rashid *et al.* 2015).

Extermination (Eradication) or inhibition of staphylococcal colonization is still considered as main strategy to prevent infection and transmission of these strains. Basis behind such a strategy is that the most staphylococcal infections are caused by endogenous strains; so, existence of *S. aureus* is a major risk factor for consequent infections (Moghadam *et al.* 2015)

Increasing multiple resistances of *S. aureus* to antibiotics makes the development of new treatment routes for serious infections a matter of urgent concern (Kuroda *et al.* 2007).

MRSA is one of typical resistant strains, and research for its prevention and treatment was carried out. The idea (concept) that MRSA infection presents different signs for treatment and diagnoses of colonization is controversial, but the differential diagnosis between bacterial infection and bacterial colonization is essential to avoid unnecessary use of anti-MRSA drugs (Shigemura *et al.* 2013).

The last-resort antibiotic for the treatment of MRSA infections was vancomycin, but MRSA resistance to vancomycin has been reported too. This suggests that MRSA will likely obtain more resistance to vancomycin in the near future. Therefore, it is increasingly essential to discover new antibiotics or to devise new actions that are effective against MRSA infections (Koyama *et al.* 2013). As a consequence, clinicians across the world faced clinical challenge in controlling MRSA (Rashid *et al.* 2015).

The main component of the bacterial cell wall is peptidoglycan which is an attractive target for the development of anti-infectious agents. It forms a huge macromolecule that surrounds the cell as a single, flexible meshwork and is closely involved in cell division. The structure determines the cell shape and maintains cell integrity by protecting it against the high internal osmotic pressure. Important antibiotics have been clinically used including β -lactams and glycopeptides that target cell wall peptidoglycan synthesis (Koyama *et al.* 2013).

2.3. Targeted pathway

It is well known that MRSA produces a yellow pigment called staphyloxanthin (STX). Recently, several research groups described that STX is a virulent factor acting as an antioxidant, with its various conjugated double bonds enabling detoxification of host immune system-generated reactive oxygen species (Sakai *et al.* 2012). The first committed step in staphyloxanthin biosynthesis by *S. aureus* is dehydrosqualene synthase (CrtM) enzyme (Pelz *et al.* 2005), which catalyzes the condensation of two farnesyl diphosphates to produce the C30 species, presqualene diphosphate (Liu *et al.* 2008), (Song *et al.* 2008), which then undergoes skeletal rearrangement and further loss of diphosphate to produce dehydrosqualene (Song *et al.* 2008). Successive dehydrogenations yield 4,4'-diaponeurosporene (Pelz *et al.* 2005), which is then further oxidized, glycosylated, and esterified to give the carotenoid, staphyloxanthin (Figure 2.1) (Liu *et al.* 2008). Hence forgetting the biosynthesis of STX may provide an alternative way to develop new drug for preventing a treatment, staphyloxanthine remain infections since the lack of STX is susceptible to neutrophil killing.

The orange carotenoid staphyloxanthin is produced by most *S. aureus* strains. The staphyloxanthin biosynthesis genes are organized in an operon, crtOPQMN, with a B-dependent promoter upstream of acyl transferase (crtO) and a termination region downstream of dehydrosqualine desaturase (crtN). The functions of the five encoded enzymes are predicted on the basis of their sequence similarity to known enzymes and by product analysis of gene deletion mutants (Pelz *et al.* 2005).

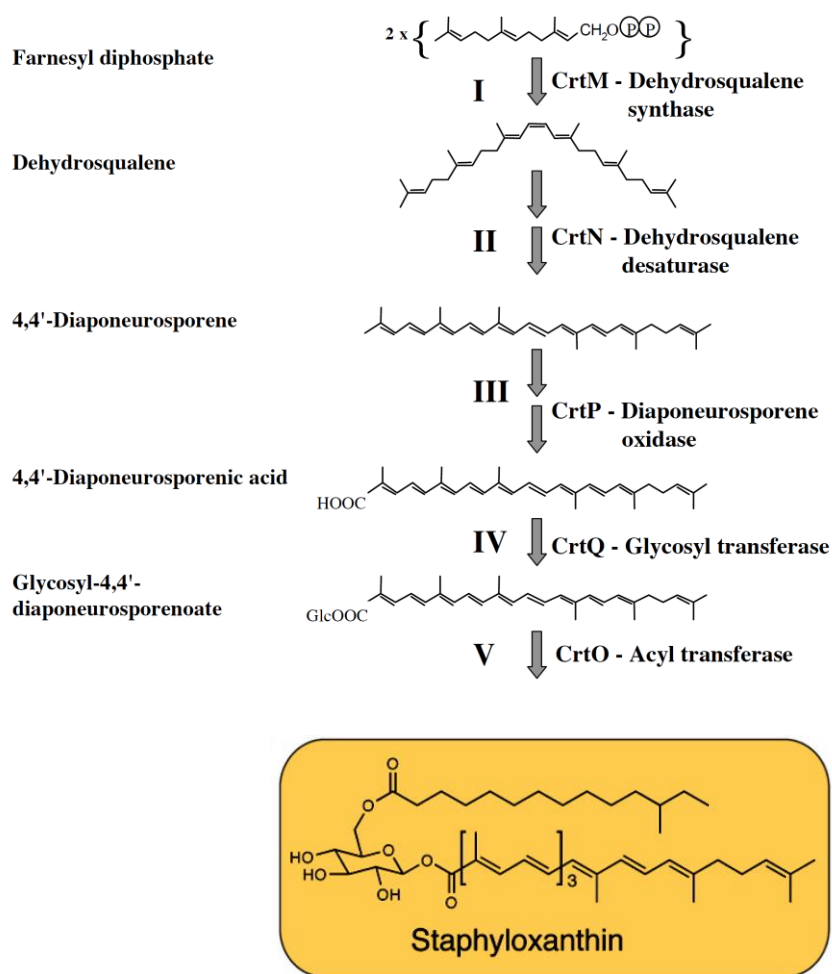


Figure 2.1. Biosynthetic pathways. Staphyloxanthin biosynthesis in *S. aureus* (Pelz et al. 2005), (Song et al. 2008).

The dehydrosqualene desaturase (CrtN) dehydrogenates dehydrosqualene to form the yellow, main intermediate 4,4-diaponeurosporene. Diaponeurosporene oxidase (CrtP), very likely a mixed function oxidase, oxidizes the terminal methyl group of 4,4-diaponeurosporene to form 4,4-diaponeurosporenic acid. Glycosyl transferase (CrtQ), a glycosyltransferase, esterifies glucose at the C1 position with the carboxyl group of 4,4-diaponeurosporenic acid to yield glycosyl 4,4-diaponeurosporenoate; this compound was the major product in the clone expressing crtPQMN. In the final step, the acyltransferase (CrtO) esterifies glucose at the C6 position with the carboxyl group of 12-methyltetradecanoic acid to yield staphyloxanthin (Pelz et al. 2005). Hence targeting are of these step is STX biosynthesis may provide a way to develop the anti-MRSA drugs.

2.4. Secondary Metabolites

Secondary metabolites are metabolic products that are not critical for vegetative growth of the producing organisms but they are considered differentiation compounds conferring adaptive roles, for example, by functioning as resistance compounds or signaling molecules in ecological interactions. They are produced at the end of the exponential phase of growth and their syntheses seriously depend on the growth conditions. Production is usually when growth is limited by the exhaustion of fundamental nutrients such as carbon or nitrogen. They are structurally different and most of them are endowed with biological activities, such as antimicrobial agents, toxins, pesticides, ionophores, bioregulators, and quorum signalling. These bioactive metabolites are extremely used as antimicrobial agents for the treatment of different diseases (Adegboye and Babalola 2013). One third of the 22,500 known microbial metabolites are the secondary metabolites of actinomycetes, mainly *Streptomyces* species (Miyaoka *et al.* 2014). Microbial secondary metabolites have been in the first appearance in the discovery of novel antimicrobial agents for pharmaceutical industry, and nowadays all indications suggests that novel compounds with potential therapeutic applications are still waiting to be discovered from secondary metabolites mainly those produced by actinomycetes. Actinomycetes are abundant producers of secondary metabolites with biological activities (Adegboye and Babalola 2013).

Secondary metabolites are provided by the producer organism with survival advantages in various ways, such as improving nutrient availability, acting as a metabolic defense mechanism, protecting against environmental stressors, enhancing competitive interactions with other organisms (Breitling *et al.* 2013). Secondary metabolites are organic compounds that often play an important role in defense systems of different organisms and are not directly involved in the normal growth, development or reproduction of an organism. (Davati and Najafi 2013).

Secondary metabolites usually include various chemical moieties, such as polyketide backbones, amino acid derivatives and sugars. Biosynthesis of secondary metabolite is catalyzed by a number of enzymes, usually encoded by genes. These genes occur nearby to one another in cluster. The gene cluster contains all the needed genes for the synthesis of a particular secondary metabolite. This includes: the genes that encode the biosynthetic enzymes, regulatory proteins, genes for resistance to the toxic

action of secondary metabolites and genes for secretion of the metabolites. Enzymes such as synthase (PKS) and non-ribosomal peptide synthetase (NRPS) are take part in the synthesis of secondary metabolites. Other enzymes responsible for the synthesis of other constitutive compounds, such as sugars, are often encoded by genes nearby to the gene cluster. Through processes such as elongation, synthesis, glycosylation, alkylation and oxidation, structurally different and complex metabolites are produced. The complete process of production and transportation of secondary metabolites are severely regulated by transcriptional regulators and transporters. The genes encoding for tailoring enzymes, transcriptional regulators and transporters are often located nearby to PKS and NRPS genes. The gene cluster size responsible for the synthesis of each secondary metabolite is usually between 10 -100 kb. (Adegboye and Babalola 2013).

The secondary metabolites frequently have unusual structures and their formation is regulated by nutrients, feedback control, growth rate, enzyme induction, and enzyme inactivation. These events generate signals which affect a cascade of regulatory actions resulting in chemical differentiation (secondary metabolism) and morphological differentiation (morphogenesis). The signal is often a low molecular weight inducer which acts by negative control, by binding to and inactivating a regulatory protein which normally prevents secondary metabolism and morphogenesis during rapid growth and nutrient sufficiency (Demain 1998). Regulation is affected by unique low molecular mass compounds, sigma factors, transfer RNA, and gene products formed during post-exponential growth. The synthases of secondary metabolism are often coded by clustered genes on chromosomal DNA and infrequently on plasmid DNA (Davati and Najafi 2013).

2.4.1. Function and Importance of Secondary Metabolites

Several hypotheses exist about the origin and function of secondary metabolites. The most accepted considers secondary metabolites as waste products that under the pressure of natural selection have evolved as messenger molecules which must endure long enough to shuttle between the various components of the microbial community. This fact, would explain the secondary metabolites tendency to be small organic molecules, as a natural consequence of their functions.

The presence of complex biosynthetic pathways for production of complex antibiotics suggest that they must have an important role in microbial survival, either as inhibitors of other competing organisms or as regulatory effectors during some stage of the cell differentiation process, since sensitive organisms need to evolve only a single enzyme of any several reaction types to inactivate most antibiotics (Tabarez 2005).

Nowadays, several arguments support the hypothesis that secondary metabolites improve the survival of the producer in competition with other living species. These arguments are as follows:

- a) Secondary metabolites act as an alternative defense mechanism, because only the organisms lacking an immune system are prolific producers of these compounds.
- b) They have sophisticated structures, complex and mechanisms of action, and energetically expensive pathways.
- c) They act in the competition between microorganisms, plant and animals.
- d) They are produced by biosynthetic genes clusters, which would only be selected if the product conferred a selective advantage. Some exactitudes of these genes clusters are the absence of non-functional genes and the presence of regulatory and resistance genes.
- e) The production of them with antibiotic activities is temporarily related with sporulation when the cells are particularly sensitivity to competitors and requiring special protection when a nutrient runs out.

Furthermore, the wide diversity of secondary metabolites suggests a broad range of functions. Nevertheless, these functions could depend on the conditions, optimal or not, surrounding the producer microorganism. Finally, due to their crucial importance the study and exploitation of secondary metabolites continue to progress despite the lack of agreement regarding why microbes produce such chemical diversity of antimicrobial compounds (Tabarez 2005).

2.4.2. Genetics of Secondary Metabolites

The genes regulating and ensuring synthesis of secondary metabolites and their expression can be grouped in 5 classes: first, structural genes, that code for enzymes involved in the biosynthesis; second, regulatory genes, that determine the repression or induction of the structural genes; third, genes that determine the resistance of the generating organism; fourth, genes controlling the compound permeability and fifth, genes that control primary pathways. The genetic regulation of all above mentioned genes is highly complicated because many environmental and microbial factors affect the production of these compounds (Tabarez 2005)

Previous studies showed that the gene cluster responsible for the production of secondary metabolites is not found in all bacteria and even in those surviving it is not regularly distributed among them. For example *Streptomyces coelicolor* has more than 20 gene clusters for the synthesis of secondary metabolites while *Streptomyces avermitilis* has 30 gene clusters. Genome mining for new candidate secondary metabolic pathways built on clustering and co-expression has proved to be a highly successful method in microbes. This useful to expect the types of antibiotic one might expect to find after extraction and purification. With the increasing number of genome nucleotide sequence information in the Gen Bank and coming of next generation sequencing it will be possible to search for secondary metabolite gene cluster candidate in a wide range of actinomycetes (Adegboye and Babalola 2013).

Functionally related genes are on the chromosome suggesting that at least part of their evolution has occurred as a unit. Evolutionally, two types of gene clusters that produce secondary metabolites can be described: first, a gene cluster might contain many genes, some of which give rise to chemical transformations of substrates and many other that do not. This type of cluster would be indicative of a natural product which has not been selected for and which is not functional. Second, a gene cluster might contain only genes that code for enzymes of a particular biosynthetic pathway, resistance genes and regulatory factors, and no “junk” genes. This type of gene cluster indicate a natural product that has been selected for and which has had a beneficial function for the producer organism (Tabarez 2005).

2.5. Angucycline Compounds

The angucyclines are a large group of natural products (Song *et al.* 2015). They are a large family of actinomycete-derived polyketide antibiotics with a four-ring skeleton that constitutes the aglycone part. Angucyclines structure consist of a benz[a]anthracene core (Ren *et al.* 2011), (Abdelmuhsen *et al.* 2014), which is commonly C- or O-glycosylated with sugar (single deoxyoligosaccharide) chains of various lengths with up to six carbohydrates, as in landomycin A, the carbohydrate composition of landomycin is a repeated sequence of β -D-olivose-(4-1)- β -D-olivose-(3-1)- α -L-rhodinose attached to a phenolic oxygen of the angucycline core. The biological activities were mainly found to be depending on the length of the saccharide chain (Helaly *et al.* 2013). They have been shown to exhibit different activities of medical interest (Rohr and Thiericke, 1992) including antitumor (Kirschning *et al.* 2000), (Kharat *et al.* 2009) and antibacterial activities (Kawasaki *et al.* 2010a), (Kormanec *et al.* 2014) as well as platelet aggregation inhibitory effects (Kirschning *et al.* 2000). The first reported compounds of this class were tetrangomycin and tetrangulol (Song *et al.* 2015).

The angucycline family of antibiotics is a large and ever-growing group of secondary metabolites of microbial origin comprising more than 100 members (Ren *et al.* 2011). The first member of this group of secondary metabolites was tetrangomycin (Figure 2.2) (Kalyon *et al.* 2013). These compounds are found widely in nature and are mainly produced by soil-dwelling *Streptomyces* bacteria. The biosynthesis of angucyclines diverges from other type II aromatic polyketides by the action of the specific cyclase that closes the fourth ring of the polyketide into an angular orientation producing UWM6 (Kallio *et al.* 2008), (Patrikainen *et al.* 2012). Aromatic polyketides include a large class of natural secondary metabolites produced by certain fungi, bacteria and plants. Many of these compounds have medically important properties, and several are in clinical use (Koskiniemi *et al.* 2007).

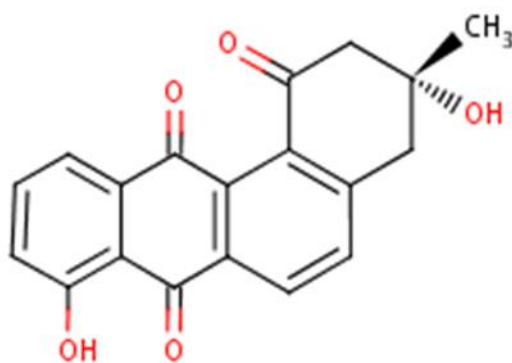


Figure 2.2. Tetrangomycin structure (Kalyon *et al.* 2013)

Aromatic polyketides are complex natural compounds well established for their biological activities and applications in medicine. They are produced by a number of different organisms, but especially *Streptomyces* bacteria have been a rich source of natural products. Angucyclines comprise a distinct group of aromatic polyketides (Rohr and Thiericke 1992), and first member of group was discovered in 1966 (Kawasaki *et al.* 2010a), but still only a few hundred structures have been described. New genetic screening studies have indicated that angucycline gene clusters are much more abundant than expected depending on traditional cultivation methods (Metsä-Ketela *et al.* 2002), which proposes that many potentially important angucyclines are yet to be discovered (Kallio *et al.* 2008).

Many of the new compounds, mainly described most recently, are from marine-derived actinomycetes, and represent new structures arising from new biosynthetic pathways. This suggests that even if marine actinomycetes are derived from terrestrial sources, they have been exposed to a new environment long enough to have evolved exceptional biosynthetic pathways of their own. The discovery of new actinomycete species producing novel chemical compounds suggests that they are metabolically active in the marine environment. Furthermore, it has become more obvious that there is great diversity of new actinomycete species within the marine environment with evidence that stems from phylogenetic analysis, metabolic requirements, morphological characteristics, and new secondary metabolite production (Raczkowski 2010).

2.6. Actinomycetes, especially genus *Streptomyces* (Angucyline producers)

Actinomycetes belong to the phylum Actinobacteria that consist of Gram positive, aerobic bacteria, were first thought to be fungi for their mycelium producing capabilities, for which the suffix “mycetes” is used (Raczkowski 2010). They are filamentous, characterized by a complex life cycle (Sharma 2014) and produce extensive branching vegetative (substrate) mycelium and aerial mycelium bearing chains of arthrospores. The substrate mycelium and spores can be pigmented, but also diffusible pigments are produced. On agar plates, they form lichenoid, leathery or butyrous colonies (Rintala 2013).

The high densities of actinomycetes in soils revealed their importance, which are one of the most significant decomposers for recalcitrant polymers such as cellulose. In addition to cellulose, *Streptomyces spp.* have shown to degrade chitin, keratin, pectin, and other hemicelluloses. These compounds can also be used as elements for their growth on artificial media. Although some pesticides inhibit their proliferation, *Nocardiosis spp.*, are known to degrade chlordane insecticides. With the ability to produce numerous antifungal compounds, actinomycetes are suspected to be associated with plant roots and hence they are believed to protect roots by using these compounds to inhibit the growth of fungal pathogens (Raczkowski 2010).

Actinomycetes produce perhaps the most diverse and most unique, unprecedented, sometimes very complicated compounds exhibiting excellent antimicrobial potency and usually low toxicity. The metabolic diversity of the actinomycetes is due to their extremely big genome, which has hundreds of transcription factors that control gene expression, allowing them to respond to specific needs (Singh *et al.* 2014). The GC-content of the DNA is 69-78 %. L-diaminopimelic acid is the characteristic compound present in the cell wall peptidoglycan of streptomycetes. The streptomycetes are able to use a wide range of organic compounds as a carbon source, with complex biological materials, such as cellulose and lignin, and can also use an inorganic nitrogen source (Rintala 2013).

Actinomycetes have provided important bioactive compounds of great commercial value and continue to be routinely screened for new bioactive substances. It

is generally accepted that the streptomycetes have a specific capacity to produce a large variety of different bioactive compounds with a wide range of activity (Singh *et al.* 2014). There are over 500 species of *Streptomyces* bacteria described by Euzéby (2008) (Raja and Prbakarana 2011). They undergo a complex process of morphological and physiological differentiation that leads to the production of exospores and specialized metabolites possessing a wide range of biological activities. While the function of many of these molecules in the natural environment is not always evident, they are believed to provide a competitive advantage to the producing organism (Gomez-Escribano *et al.* 2015).

Among the 140 defined Actinomycetes genera, only a few are responsible for the more than 10,000 bioactive compounds in clinical use (Adegboye and Babalola 2013), more than half of the 10,000 documented bioactive compounds, have offered over 50 years of interest to industry and academia (Anderson and Wellington 2001). It is representing 45% of known bioactive microbial metabolites were isolated from various actinomycetes species (Sharma 2014), and 7600 compounds are produced by *Streptomyces* species (74% of all actinomycetales), while the rare actinomycetes represent 26%, altogether 2500 compounds (Raja and Prbakarana 2011), (Singh *et al.* 2014).

The composition of cell wall in actinomycetes differs greatly among different groups and is of considerable taxonomic value. Four major cell wall types are distinguished in these filamentous bacteria depends on three features of peptidoglycan composition and structure, which are: diaminopimelic acid isomer on tetrapeptide side chain position 3, sugar content of peptidoglycan, and the presence of glycine in interpeptide bridges (Adegboye and Babalola 2012).

Microorganisms included in the genus *Streptomyces* that inhabit soil niches, thus facing ever changing environmental conditions and nutrient scarcity. Along evolution, this challenging environment has pushed the genus *Streptomyces* towards complex adaptive responses. Among them, two-component systems (TCSs) are the most important transduction signal mechanism in bacteria, allowing the translation of these rapid environmental or nutritional changes into a regulatory readout. Typically, TCSs

comprise a membrane-bound histidine kinase, which senses specific environmental stimuli, and a cognate regulator, which mediates the cellular response, mainly through the transcriptional regulation of target genes (Rodríguez *et al.* 2013).

It is now evident that as new molecular approaches are used to unravel the microbial community of unique marine environments, new species of actinomycetes have continued to reappear. However, these new species of actinomycetes indigenous to the marine habitat, or they derived from terrestrial run-off that are in fact a source of dormant actinomycete spores that can survive for many years (Raczkowski 2010). Bacteria belonging to the genus *Streptomyces* harbor a high number of TCSs in comparison with other bacterial genera, probably due to the changing environment that these organisms must inhabit (Rodríguez *et al.* 2013).

Approaches for overproduction of microbial products can be based on microbial response (elicitors, quorum sensing), genetic, ribosome, and metabolic engineering. Also, molecular genetic improvement methods include amplification of secondary metabolites biosynthetic genes, inactivation of competing pathways, disruption or amplification of regulatory genes, manipulation of secretory mechanisms, expression of a convenient heterologous protein and combinatorial biosynthesis (Davati and Najafi 2013). In 1995 more undoubted data, through the use of 16S rRNA genus-specific probe with hybridization experiments, suggested that salt marsh *Streptomyces* sp. strains were indigenous to the marine environment by being important proportion of the microbiota of the salt marsh environment through an entire annual cycle. Scientists revealed that *Streptomyces* sp. occurred dominantly in the salt marsh community and that dormant spores were not the source of their abundance (Raczkowski 2010).

2.7. Protein (Receptor) Structure and Function

2.7.1. Introduction to Proteins

The highly specific protein complex formation is an important process in biology and an integral constituent to all major biochemical pathways. The structures of these protein complexes can provide detailed understanding into the mechanisms of function, starting from disease pathogenesis, to drug action, to promoting our

understanding of innate biochemical processes. Using of the computational tools of molecular modeling, is the beginning to develop methods to predict high-resolution structures for protein complexes and assume how their functions arise from their structures (Chaudhury 2010). Multiple weak interactions stabilize protein structure, hydrophobic interactions are the major contributors to stabilizing the spherical form of most soluble proteins, ionic interactions and hydrogen bonds are optimized in the thermodynamically most stable structures (Nelson and Cox 2008).

One of the five major biological macromolecules is proteins, and they are responsible for a various biochemical processes from structure, to signaling, to catalyzing vital biochemical reactions. They are polymers of amino acids that are encoded by genes.

In the process of translation, a mRNA transcript from the nucleus is used to create the protein chain in the ribosome. After translation, this amino acid polymer (polypeptide chain) adopts the lowest free-energy conformation in solution, through a protein folding process. Most proteins fold into a very specific shape or structure, and their function is directly outcome from this structure. A typical folding energy for a protein is -10 kcal/mol, which is means that well over 99% of the protein molecules in solution adopt this lowest-free energy conformation (Chaudhury 2010).

Like protein folding, almost all protein functions come from this basic thermodynamic principle: in a given environment the protein will adopt the lowest energy conformation (Chaudhury 2010).

In almost all cases, this lowest energy conformation is highly specific structure that biologically-evolved. For example, in protein-protein binding, which are free in solution, the lowest energy conformation will be a highly specific complex between the two proteins. The energy of a given conformation is a function of numerous molecular forces that act on that conformation, these are electrostatics, van der Waals interactions, hydrogen-bonding, and solvation energies (Chaudhury 2010).

2.7.2. Protein Structure

The monomers of a polypeptide chain, are known as residues, form a polymer through peptide bonds. The polypeptide chain itself (backbone) is made up totally of an N-C α bond, a C α -C bond, and a C-N bond, repeated for each amino acid. The protein structure and function specificity arises from the sequence of its amino acids. Although all polypeptide chain atoms for all residues are identical, each residue in the backbone is one of 20 amino acids. Amino acids differ from one another in their side-chains, defined as those non-backbone atoms bonded to the C α atom (Chaudhury 2010). The 20 common amino acids practically never occur in equal amounts in a protein. Some of them may occur only once or occur in large number or not at all in a given type of protein. Differences in amino acids composition and sequence leads to differences in protein function. Some differences in sequence are possible for a specific protein, with little or no effect on function. There are hundreds of individual bonds in a covalent backbone of a typical protein, free rotation is possible around many of these bonds, and as a consequence the protein can assume a huge conformation. However, each protein has a structural function or specific chemical activity, strongly suggesting that each has a unique three-dimensional structure (Nelson and Cox 2008).

The energy of a certain protein conformation is a result of molecular forces that act on and between all non-bonded atoms in that conformation. In protein biophysics the van der Waals energies, steric repulsions, electrostatic energies, solvation energies, and hydrogen bonding energies are the most significant energy contributions that favor one conformation over another. These component energy functions of the atomic positions and atom types of the atoms in the protein structure (Chaudhury 2010).

The C α of nearby amino acid residues are separated by three covalent bonds, arranged as C α -C-N-C α . X-ray diffraction studies of crystals of amino acids and of dipeptides and tripeptides showed that the peptide C-N bond is a bit shorter than the C-N bond in a simple amine and that the atoms linked with the peptide bond are coplanar.

This means the existence of a partial sharing of two pairs of electrons or resonance between the carbonyl oxygen and the amide nitrogen (Figure 2.3.a). The oxygen with a partial negative charge and the nitrogen with a partial positive charge,

setting up a small electric dipole. The six atoms of the peptide group located in a single plane, with the oxygen atom of the carbonyl group trans to hydrogen atom of the amide nitrogen. Pauling and Corey concluded from these findings that the peptide C-N bonds, cannot rotate freely because of their partial double-bond character. Rotation is permitted about the N-C $_{\alpha}$ and the C $_{\alpha}$ -C bonds. The backbone of a polypeptide chain accordingly can be pictured as a series of rigid planes, with repeated planes sharing a common point of rotation at C $_{\alpha}$ (Figure 2.3.b). So the range of possible conformations for a polypeptide chain limited by the rigid peptide bonds (Nelson and Cox 2008).

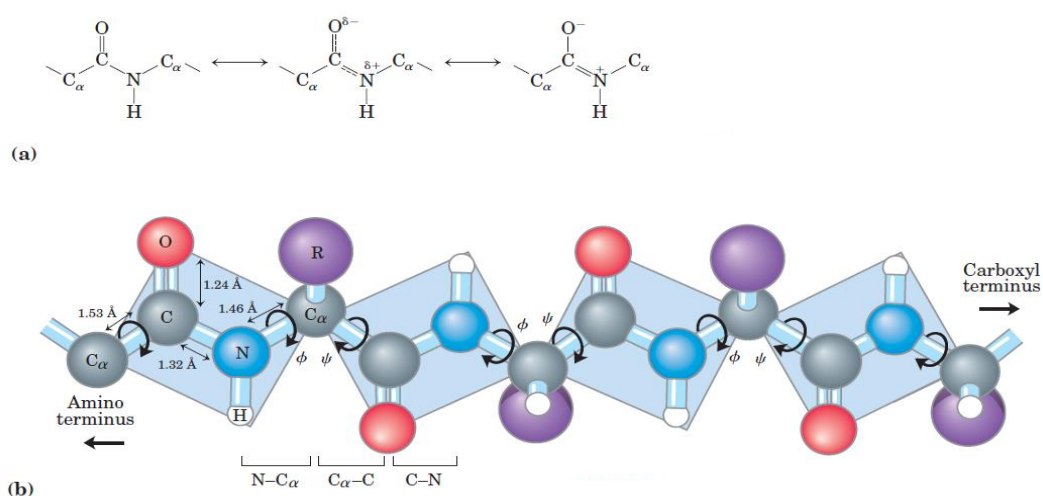


Figure 2.3. The planar peptide group. (a) Each peptide bond has some double-bond character due to resonance and hence cannot rotate. (b) Three bonds separate sequential α carbons in a polypeptide chain. The N-C $_{\alpha}$ and C $_{\alpha}$ -C bonds can rotate, described by dihedral angles designated ϕ and ψ , respectively. The peptide C-N bond is not free to rotate. Other single bonds in the backbone may also be rotationally hindered, depending on the size and charge of the R groups (Nelson and Cox 2008).

2.7.3. Protein Data Bank

The Protein Data Bank (PDB; www.rcsb.org) (RCSB: Research Collaborator for Structural Bioinformatics) is an archive or library of experimentally determined three dimensional structures of biological macromolecules. The information contained in this archive includes atomic coordinates, crystallographic structure factors and NMR experimental data. In addition to coordinates, each deposition also consist of the names of molecules, primary, secondary structure information, and sequence database

references, where appropriate, and ligand and biological assembly information, details information about data collection and structure solution, and bibliographic citations.

All the Protein Data Bank items are stored in a special file format - PDB. Every PDB file is demonstrated in a number of lines. Each line in the PDB item file consists of 80 columns and is self-identifying. There is a record name in the first six columns of every line. Record names are listed and explained in detail in the PDB format guide. Collection of record types which consists of one or more lines is another way to describe a PDB file (Novotarskyi 2013).

2.8. Ligands

Many proteins functions involve the reversible binding of other molecules. Ligand is a molecule bound reversibly by a protein, and may be any kind of molecule, including another protein. Binding site is the location that ligand binds on the protein, which is matching the ligand in size, shape, charge, and hydrophobic or hydrophilic character. Additionally, the interaction is specific: the protein can favor among the thousands of different molecules in its environment and selectively bind only one or a few. A certain protein may have different binding sites for several different ligands (Nelson and Cox 2008).

A ligand binds at a site on the protein called the binding site, which is complementary to the ligand in size, shape, charge, and hydrophobic or hydrophilic character. Furthermore, the interaction is specific: the protein can discriminate among the thousands of different molecules in its environment and selectively bind only one or a few. A given protein may have separate binding sites for several different ligands. These specific molecular interactions are crucial in maintaining the high degree of order in a living system. This discussion excludes the binding of water, which may interact weakly and nonspecifically with many parts of a protein (Nelson and Cox 2008).

2.9. Protein-Ligand Interaction

Interactions of protein-ligand have a central role in all processes in living systems. A complete understanding of protein-ligand interactions is of great interest as

it provides opportunities for therapeutic intervention and understanding function. Molecular recognition is, however, a complex interaction of several factors like intermolecular interactions of protein, ligand and the surrounding solvent, conformational differences of binding partners and the thermodynamics of molecular association. Experimental and computational techniques have been developed over the past few decades that shed light on the role of these factors (Haider 2010).

Interactions between ligands and proteins may be regulated, usually through specific interactions with one or more additional ligands. These other ligands may cause conformational changes in the protein that affect the binding of the first ligand (Nelson and Cox 2008).

2.9.1. Non-covalent binding

The non-covalent binding of ligand (small-molecule) to proteins (receptors) is mediated by a various inter-atomic interactions. Generally, these include electrostatic and van der Waal interactions (Figure 2.4) the affinity of protein-ligand binding also greatly relies on contributions from other factors such as entropy, flexibility of receptor structure, desolvation, and the structural water molecules in the binding site (Haider 2010).

2.9.2. Electrostatic Interaction

Electrostatic interactions is a result of forces that are generally accepted to be the most important intermolecular driving factors for ligand-protein binding. Electrostatic phenomena in biomolecular systems are very complex because of the long-range nature of electrostatic forces among large numbers of interacting atoms. The existence of charged groups in proteins further complicates the situation (Tunca 2012)

Electrostatic complementary between the protein and the ligand at the binding site is very important for complex formation via electrostatic interactions such as hydrogen bonding, salt bridges, and metal interactions (Figure 2.4).

Hydrogen bonding is very important directional interaction in biological macromolecules, known for giving stability to protein structure and selectivity to

protein-ligand interactions. In general, hydrogen bonding takes place between two electronegative atoms, one of which (donor) has a covalently bound hydrogen atom and the other (acceptor) has a long pair of electrons. The strong electrostatic inter-attraction arises from attractive interaction between partial positive charge on the hydrogen atom and partial negative charge on the acceptor atom. Theoretical and experimental studies have approved an additional covalent component to hydrogen bonds also which is based on the interaction between empty (σ^*) anti-bonding orbital of the hydrogen atom and highest occupied orbital of the acceptor (Haider 2010).

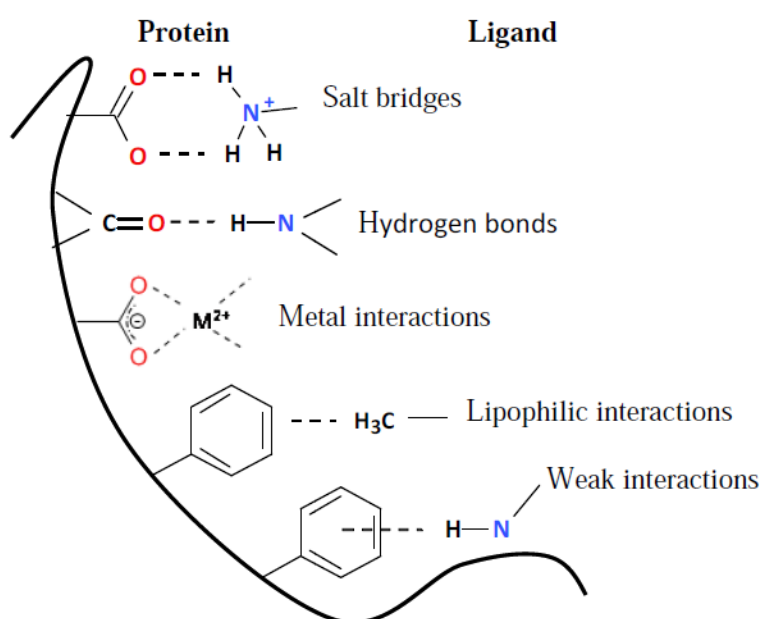


Figure 2.4. Major types of non-bonded interactions in protein-ligand complexes (Haider 2010)

2.9.3. Hydrophobic Interaction

Hydrophobic interactions include contacts between non-polar parts of the molecule (Figure 2.5). In protein-ligand complexes non-polar parts at the interacting surface are embedded or buried upon binding. This causes displacement of water molecules and this mean increasing the entropy. These types of interaction are therefore entropy-driven and have been shown to play central role in ligand binding. The burial of non-polar surface area and binding affinity relationship is well established and amounts to an affinity gain of 30 cal/mol for 1 Å^2 of buried lipophilic surface area. This indicates that adjusting non-polar contacts of ligand atoms in hydrophobic protein pockets results

in strength binding. For example, Peters et al. demonstrated that improving interactions of aromatic rings in hydrophobic pocket of dipeptidyl peptidase IV resulted in 10^5 fold increase in affinity (Haider 2010).

In protein binding sites aromatic residues such as His, Phe, Trp and Tyr are frequently involved in aryl-aryl interactions. They are known to interact with each other mostly via one of two geometries: parallel displaced stacking and T-shaped edge to face interaction. Quantum mechanical studies on model system like benzene dimers have shown that these two geometries are isoenergetic. In protein structures, however, parallel displaced geometry has been more recurrently observed. Aliphatic-aromatic interactions including aromatic rings and alkyl groups are also commonly occurring interactions at non-polar interfaces. Similar to aromatic-aromatic interactions, preferable interaction geometries include parallel displaced and edge-to-face interactions. The strength of aliphatic-aromatic interactions, specially, CH- π interactions vary with increasing acidity of CH groups (Haider 2010).

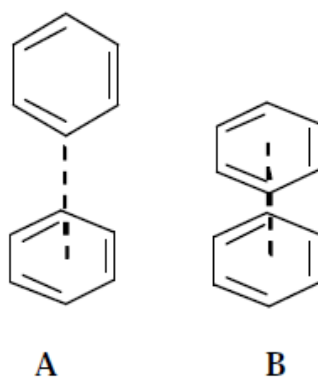


Figure 2.5. Aryl-aryl interactions in protein structure and protein-ligand complexes. A. Edge-to-face geometry, B. Parallel stacking geometry (Haider 2010)

2.9.4. Binding affinity and binding site

The binding affinity is used to describe how tightly or strongly a ligand bind to its target and is controlled by non-covalent interactions such as electrostatic and van der Waals forces, including solvation and desolvation contributions. These interactions are

important for structural and energetic recognition between ligand and target. Although covalent bonds are stronger than non-covalent interactions, small interactions accumulate to make important participations to stabilize ligand binding (Tunca 2012).

The binding site can be defined as a set of residues belonging to the protein that lies within the zone of 4.5\AA from any atom of the corresponding ligand (Anand et al. 2014). For a strong ligand binding to a target, some requirements should be fulfilled: High level of steric complementarity should be between the ligand and the target protein. Ligand and the target protein surface properties should chemically complement to each other. In the meantime lipophilic parts of the protein are mostly in contact with the lipophilic parts of the ligand and polar groups are usually matching accordingly to form hydrogen bonds or ionic interactions, surface properties of both sides should match. The ligand should be in an energetically favorable conformation for stability (Tunca 2012).

2.9.5. Affinity prediction problem

The information about 3D structures of the targets and/or the ligands and the correct prediction of the binding modes and the target-ligand complex structures constitute the basis for receptor-based drug design, however understanding the protein-ligand interactions on the molecular level and the precise prediction of these interactions determine the success of a receptor-based drug project. Even though different docking algorithms are able to generate experimentally observed binding modes of ligands to protein, it is still a challenge to recognize and pick them in huge libraries and give accurate scores to rank them. The efficiency of a computational drug design method relies on the true prediction of binding affinity (Tunca 2012).

2.10. Molecular Docking

Over the past 25 years, advances in structural biology have greatly changed the technique in which drugs are designed and discovered. Nowadays, rather than treating protein targets as simple black boxes, now pharmaceutical scientists are treating them as complex molecular entities that have well-defined structures with active sites that can be activated or deactivated rationally with small molecule ligands (Dias 2011).

Molecular docking is a computer simulation procedure to predict the conformation of a receptor-ligand complex, where the receptor is usually a protein and the ligand is either a small molecule or another protein. It can also be defined as a simulation process where a ligand position is estimated in a predicted or predefined binding site (Novotarskyi, 2013).

Docking studies, in general have two main aims: true structural modeling and correct binding affinity predictions. Therefore, most docking algorithms concentrate on ligand and sometimes on flexibility of protein, and explore the suitable space available to the ligand in the binding site. According to how ligand flexibility is treated, mainly search algorithms categorized in: random or stochastic method, systematic methods and simulation methods. In docking program the research algorithm can apply only a single method or a combination of different methods (Tunca 2012).

2.10.1. AutoDock Vina

AutoDock Vina (Trott and Olson 2010) is the updated member of AutoDock suite introduced in 2010. It has been developed by Dr. Oleg Trott in the Molecular Graphics Lab at The Scripps Research Institute. It differs from earlier versions in many regards, one of them the accessibility. The three-dimensional structures of molecules with polar hydrogens only, as AutoDock Vina still uses United Atom model, and a box definition is required for search space. Partial charges, pre-calculated interaction energy grids or solvation parameters are not necessary for the simulation (Reska 2011). AutoDock Vina does not provide the user with a choice of search algorithm, it uses Iterated Local search global optimizer instead. Steps consisting of mutation and subsequent local optimization are performed in this algorithm, with each step being accepted according to the Metropolis criterion. (Reska 2011)

Changes in conformation of protein may be indirect, reflecting molecular vibrations and small movements of amino acid residues throughout the protein. A protein flexing in this way is sometimes said to “breathe.” This may also be quite dramatic, with major fragments of the protein structure moving as much as several nanometers. Specific conformational changes are normally essential to a protein’s function. The binding of protein and ligand is repeatedly coupled to a conformational

change in the protein that makes the binding site to the ligand more complementary, allowing tighter binding. Ligands and proteins interactions may be regulated, usually through specific interactions with one or more extra ligands. These extra ligands may cause conformational changes in the protein acts on the binding of the first ligand (Nelson and Cox 2008).

The contact surfaces relatively small between the ligand and the protein, much smaller than in protein-protein docking. In this case, the main challenge is the ligand's flexibility as it can fit to the protein's surface in various way. Computational methods are then used to predict ligands binding affinity and conformation. This method is used to obtain leads for medical development as drug-like molecules are screened with this method (Pétursson 2014). Protein interactions with ligands, or with other proteins, or surfaces are controlled by a complex arrangement of intermolecular processes. Such interactions depend on the specificity in the binding site and the non-specific forces outside the binding pocket. The aim of various biophysical studies is to identify the molecular forces that control biological interactions and to use this known information for rationally or intelligence manipulate of the protein structure or function by modifying the protein, the interacting ligand, or both (Dias 2011).

3. MATERIALS AND METHODS

The crystal structures of dehydrosqualene synthase enzyme (CrtM) which catalysis the first committed step in staphyloxanthin biosynthesis, (PDB-ID: 3W7F, 3ACW) the former involving magnesium were downloaded from Protein Data Bank (PDB) (<http://www.rcsb.org>), one involves an inhibitor and the other (3W7F) involves magnesium and without magnesium. All the non-standard amino acid residues were removed from that structures and they were separately prepared as receptors and ligands using Discovery Studio 4.1 (<http://accerys.com>), (Maldonado-Rojas and Olivero-Verbel 2012).

MGL Tools 1.5.6 (<http://mgltools.scripps.edu>) was employed to prepare protein and ligand structures for molecular docking, after opening the protein the polar hydrogens added to the structures and the structures saved in (PDBQT) file format.

In present study a total of 157 ligand structures were extracted from literatures and articles comprises the angucycline antibiotics (Table 3.1) and these 2D structures are drawn by ChemDraw Pro 12.0 (www.cambridgesoft.com) and they were converted to 3D structures using Discovery Studio 4.1 (<http://accerys.com>) they are shown in Appendix IV for the lack of the space.

Table 3.1. Angucycline Groups Antibiotics and their references

#	Angucycline Antibiotics	References
1	Marangucycline_A	(Song et al. 2015)
2	Marangucycline_B	(Song et al. 2015)
3	Actinosporin_A	(Abdelmohsen et al. 2014), (Dashti et al. 2014)
4	Actinosporin_B	(Abdelmohsen et al. 2014), (Dashti et al. 2014)
5	Hatomarubigin_A	(Kawasaki et al. 2010a, b), (Izawa et al. 2013)
6	Hatomarubigin_B	(Kawasaki et al. 2010a, b), (Izawa et al. 2013)
7	Hatomarubigin_C	(Abdelfattah et al. 2003), (Kawasaki et al. 2010a, b), (Izawa et al. 2013)
8	Hatomarubigin_D	(Kawasaki et al. 2010a, b), (Izawa et al. 2013)
9	Hatomarubigin_E	(Kawasaki et al. 2010b), (Izawa et al. 2013)
10	Hatomarubigin_F	(Izawa et al. 2013)

3. MATERIALS AND METHODS

Table 3.1. Angucycline Groups Antibiotics and their references (continue)

#	Angucycline Antibiotics	References
11	Rubiginone_B2	(Kawasaki et al. 2010a), (Izawa et al. 2013)
12	Warkmycin	(Helaly et al. 2013)
13	Waldiomycin	(Igarashi et al. 2013)
14	Langkocycline_A1	(Kalyon et al. 2013)
15	Langkocycline_A2	(Kalyon et al. 2013)
16	Langkocycline_A3	(Kalyon et al. 2013)
17	Langkocycline_B1	(Kalyon et al. 2013)
18	Langkocycline_B2	(Kalyon et al. 2013)
19	Fradimycin_C	(Ganesan et al. 2013)
20	UWM6	(Kallio et al. 2008), (Kharel & Rohr 2012), (Patrikainen et al. 2012)
21	Jadomycin_B	(Fan et al. 2012a, b)
22	Jadomycin_A	(Fan et al. 2012a)
23	L_digitoxosyl_dehydro-rabelomycin	(Fan et al. 2012a)
24	Dehydrorabelomycin	(Fan et al. 2012a), (Patrikainen et al. 2012)
25	Saprolmycin_A	(Nakagawa et al. 2012)
26	Saprolmycin_B	(Nakagawa et al. 2012)
27	Saprolmycin_C	(Nakagawa et al. 2012)
28	Saprolmycin_D	(Nakagawa et al. 2012)
29	Saprolmycin_E	(Nakagawa et al. 2012)
30	Saquayamycin_A	(Nakagawa et al. 2012)
31	Gaudimycin_A	(Kallio et al. 2008)
32	Gaudimycin_B	(Kallio et al. 2008)
33	Gaudimycin_C	(Kallio et al. 2008), (Patrikainen et al. 2012)
34	Aquayamycin	(Faust et al. 2000), (Igarashi et al. 2013)
35	Landomycin_A	(Kirschning et al. 2000), (Kharel & Rohr 2012)
36	Landomycin_E	(Zhu et al. 2005), (Kawasaki et al. 2010a), (Kharel & Rohr 2012)
37	Landomycin_Z	(Kharel & Rohr 2012)
38	Grincamycin	(Huang et al. 2012)
39	Grincamycin_F	(Huang et al. 2012)
40	JBIR_90	(Ueda et al. 2011)
41	JBIR_116	(Ueda et al. 2011)

Table 3.1. Angucycline Groups Antibiotics and their references (continue)

#	Angucycline Antibiotics	References
42	JBIR_91	(Ueda et al. 2011)
43	JBIR_92	(Ueda et al. 2011)
44	JBIR_93	(Ueda et al. 2011)
45	Urdamycin_A	(Rohr & Thiericke 1992), (Faust et al. 2000), (Ueda et al. 2011)
46	Urdamycin_B	(Ueda et al. 2011)
47	N05WA963A	(Ren et al. 2011)
48	N05WA963B	(Ren et al. 2011)
49	N05WA963D	(Ren et al. 2011)
50	JBIR_88	(Motohashi et al. 2010)
51	Rabelomycin	(Rohr & Thiericke 1992), (Faust et al. 2000), (Metsa-ketela et al. 2002), (Metsa-ketela et al. 2003), (Metsa-ketela et al. 2004), (Kallio et al. 2008), (Kharel et al. 2010), (Kharel & Rohr 2012), (Igarashi et al. 2013)
52	Mayamycin	(Schneemann et al. 2010)
53	8_O_methyltetrangomycin	(Maruna et al. 2010)
54	8_O_methyltetrangulol	(Maruna et al. 2010)
55	8_O_methyl_7_deoxo_7_hydroxytetrangomycin	(Maruna et al. 2010)
56	Moromycin_A	(Abdelfattah et al. 2008)
57	Moromycin_B	(Abdelfattah et al. 2008)
58	Saquayamycin_B	(Abdelfattah et al. 2008)
59	Sch_47554	(Basnet et al. 2006)
60	Sch_47555	(Basnet et al. 2006)
61	BE_23254	(Dey & Mal 2005)
62	Tetrangomycin	(Rohr & Thiericke 1992), (Zhu et al. 2005)
63	Tetrangulol	(Rohr & Thiericke 1992)
64	Ochromocinone	(Rohr & Thiericke 1992), (Kaliappan & Ravikumar 2007)
65	PI_080 (=PI-6621)	(Rohr & Thiericke 1992)
66	Urdamycin_D	(Rohr & Thiericke 1992)
67	Urdamycin_H	(Rohr & Thiericke 1992)
68	8_O_methylrabelomycin	(Rohr & Thiericke 1992)
69	Urdamycin_C	(Rohr & Thiericke 1992)
70	Urdamycin_E	(Rohr & Thiericke 1992)

3. MATERIALS AND METHODS

Table 3.1. Angucycline Groups Antibiotics and their references (continue)

#	Angucycline Antibiotics	References
71	Urdamycin_F	(Rohr & Thiericke 1992)
72	Urdamycin_G	(Rohr & Thiericke 1992)
73	Sakyomicin_A	(Rohr & Thiericke 1992), (Igarashi et al. 2013)
74	Sakyomicin_B	(Rohr & Thiericke 1992)
75	Sakyomicin_C	(Rohr & Thiericke 1992)
76	Sakyomicin_D	(Rohr & Thiericke 1992)
77	Sakyomicin_E	(Rohr & Thiericke 1992)
78	BE_7585A	(Rohr & Thiericke 1992)
79	8_O_methylurdamycin_A	(Henkel et al. 1988)
80	Seitomycin	(Abdelfattah et al. 2003)
81	Tetragulol methyl ether	(Abdelfattah et al. 2003)
82	SM_196B	(Rohr & Thiericke 1992), (Abdelfattah et al. 2003)
83	Saquayamycin_A1	(Alvi et al. 2000)
84	A_7884	(Alvi et al. 1999)
85	Aquamycin	(Alvi et al. 1999)
86	SM_196 A	(Rohr & Thiericke 1992)
87	C104_Aglycon	(Rohr & Thiericke 1992)
88	C104	(Rohr & Thiericke 1992)
89	3_Deoxyrabelomycin	(Rohr & Thiericke 1992)
90	6_Deoxy_8_O_methyl-rabelomycin	(Rohr & Thiericke 1992)
91	Aggreticin (OM-4842)	(Rohr & Thiericke 1992)
92	BA_12100B	(Rohr & Thiericke 1992)
93	BA_12100C	(Rohr & Thiericke 1992)
94	BA_12100D	(Rohr & Thiericke 1992)
95	BA_12100E	(Rohr & Thiericke 1992)
96	BA_12100Z1	(Rohr & Thiericke 1992)
97	BA_12100Z2	(Rohr & Thiericke 1992)
98	BA_12100Z3	(Rohr & Thiericke 1992)
99	BA_12100MY_1	(Rohr & Thiericke 1992)
100	Benzanthrin_A	(Rohr & Thiericke 1992)
101	Benzanthrin_B	(Rohr & Thiericke 1992)

Table 3.1. Angucycline Groups Antibiotics and their references (continue)

#	Angucycline Antibiotics	References
102	Capoamycin	(Rohr & Thiericke 1992)
103	De_O_acylcapoamycin	(Rohr & Thiericke 1992)
104	Elmycin_A	(Rohr & Thiericke 1992)
105	Elmycin_B	(Rohr & Thiericke 1992)
106	Elmycin_C	(Rohr & Thiericke 1992)
107	Elmycin_D	(Rohr & Thiericke 1992)
108	Elmycin_E	(Rohr & Thiericke 1992)
109	X_14881_B	(Rohr & Thiericke 1992)
110	X_14881_D	(Rohr & Thiericke 1992)
111	Emycin_A	(Rohr & Thiericke 1992)
112	SF_2315_A	(Rohr & Thiericke 1992)
113	SS_228Y	(Rohr & Thiericke 1992)
114	Fridamycin_C	(Rohr & Thiericke 1992)
115	Fujianmycin_A	(Rohr & Thiericke 1992)
116	Fujianmycin_B	(Rohr & Thiericke 1992)
117	MM_47755	(Rohr & Thiericke 1992)
118	PD116779	(Rohr & Thiericke 1992)
119	X_14881_C	(Rohr & Thiericke 1992)
120	X_14881_E	(Rohr & Thiericke 1992)
121	X_14881_A	(Rohr & Thiericke 1992)
122	Landomycin_A	(Rohr & Thiericke 1992)
123	Landomycin_B	(Rohr & Thiericke 1992)
124	Landomycin_C	(Rohr & Thiericke 1992)
125	Landomycin_D	(Rohr & Thiericke 1992)
126	PD116740	(Rohr & Thiericke 1992)
127	TAN_1085	(Rohr & Thiericke 1992)
128	Landomycin_F	(Kharel & Rohr 2012)
129	Homorabelomycin	(Kharel & Rohr 2012)
130	Prejadomycin	(Kharel & Rohr 2012)
131	Homoprejadomycin	(Kharel & Rohr 2012)
132	11_HydroxyTetrangomycin	(Kharel & Rohr 2012)

3. MATERIALS AND METHODS

Table 3.1. Angucycline Groups Antibiotics and their references (continue)

#	Angucycline Antibiotics	References
133	Homo_UWM6	(Kharel & Rohr 2012)
134	PI_1894B (Vineomycin A) (OS-4742A1)	(Rohr & Thiericke 1992)
135	Yoronomycin	(Rohr & Thiericke 1992)
136	Urdamycin_I	(Faust et al. 2000)
137	Urdamycin_J	(Faust et al. 2000)
138	Urdamycin_K	(Faust et al. 2000)
139	Kerriamycin_A	(Rohr & Thiericke 1992)
140	Kerriamycin_B	(Rohr & Thiericke 1992)
141	Kerriamycin_C	(Rohr & Thiericke 1992)
142	PI_083	(Rohr & Thiericke 1992)
143	PI_085	(Rohr & Thiericke 1992)
144	PI_087	(Rohr & Thiericke 1992)
145	Saquayamycin_A	(Rohr & Thiericke 1992)
146	Saquayamycin_B	(Rohr & Thiericke 1992)
147	Saquayamycin_C	(Rohr & Thiericke 1992)
148	Saquayamycin_D	(Rohr & Thiericke 1992)
149	Saquayamycin_K1	(Rohr & Thiericke 1992)
150	Saquayamycin_K2	(Rohr & Thiericke 1992)
151	Saquayamycin_K4	(Rohr & Thiericke 1992)
152	Saquayamycin_K7	(Rohr & Thiericke 1992)
153	Saquayamycin_Z	(Antal et al. 2005)
154	Retymicin	(Antal et al. 2005)
155	Vineomycin_A1	(Nakagawa & Omura 1996)
156	Azicemicin_A	(Ogasawara and Liu 2009)
157	Azicemicin_B	(Ogasawara and Liu 2009)

The docking site for the ligands on the protein was defined by establishing a cube, where is fairly simple, it involves manipulating a coloured box into the area, which includes a docking stage. The volume can be as large or as small as required. But there is an exponential increase in computation time as the volume of the box increased.

In our docking experiments we used dimensions $38 \times 38 \times 38 \text{ \AA}$, covering the ligand binding site with a grid point spacing of 1.0 \AA , and center grid boxes of 55.653, 12.678 and 52.021 in X, Y and Z dimensions, respectively.

Once the docking area has been defined the coordinates of the grid box in a configuration file must be written in order in the text document file which is required to supply the AutoDock Vina 1.1.2 (Trott and Olson 2010) via the command line. The configuration file also specifies the ligand molecule, the protein to dock into and the exhaustiveness of the search which can be set between 1 and 8, where 8 is the most exhaustive search. This was the present used in our docking experiments (Stevenson 2012).

AutoDock Vina (Trott and Olson 2010) was run on the Windows 8.1 operating system and all ligands (angucycline antibiotics compounds) were docked to the protein with and without magnesium. Three runs were performed for every single angucycline compound.

4. RESULTS AND DISCUSSION

First of all to validate approach of docking, inhibitor and substrate were docked and they were compared with their original x-ray structures as shown in (Figure 4.1), it demonstrate that both ligands are docked into the active site of dehydrosqualine synthase enzyme (CrtM) (PDB ID: 3ACW) in a similar manner to the original coordinates with very low RMSDs, or zero RMSD in case of substrate and farnesyl diphosphate with CrtM (PDB ID: 3W7F) as shown in (Figure 4.2).

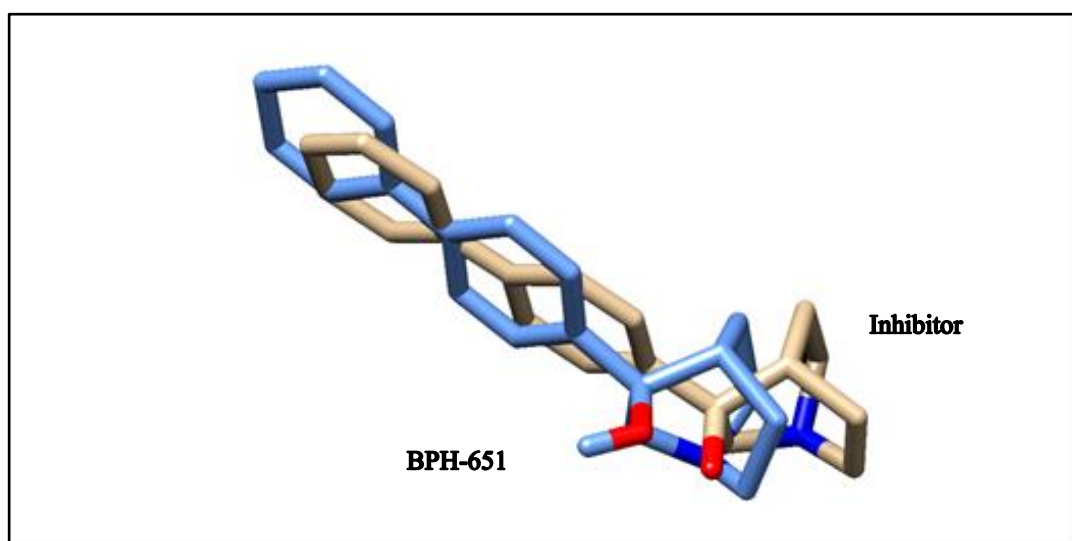


Figure 4.1. Superimpose structure of docked inhibitor with its original BPH-651 coordinates in the complex with CrtM

The active site of inhibitor BPH-651 with dock score -11.5 kcal/mol involves (Ala134, Ala157, Asn168, Gln165, Gly138, Gly161, Ile241, Leu141, Leu145, Leu160, Leu164, Phe22, Phe26, Val137) for that of farnesyl-farnesyl diphosphate the dock score -8.3 kcal/mol and involved residues are (Ala134, Ala157, Arg171, Arg265, Arg45, Asn168, Gln165, Gly138, Gly161, His18, Leu141, Leu145, Leu160, Leu164, Lys20, Phe22, Phe233, Ser19, Ser21, Tyr248, Tyr41, MG304) for CrtM. It seems that Arg45, Arg171, Arg265 are the main residues for the substrate (Appendices I, II, and III).

After seeing the success of docking protocol all ligands were successfully docked into the proteins. They were all docked into the active site where inhibitor and farnesyl diphosphate (substrate) are located.

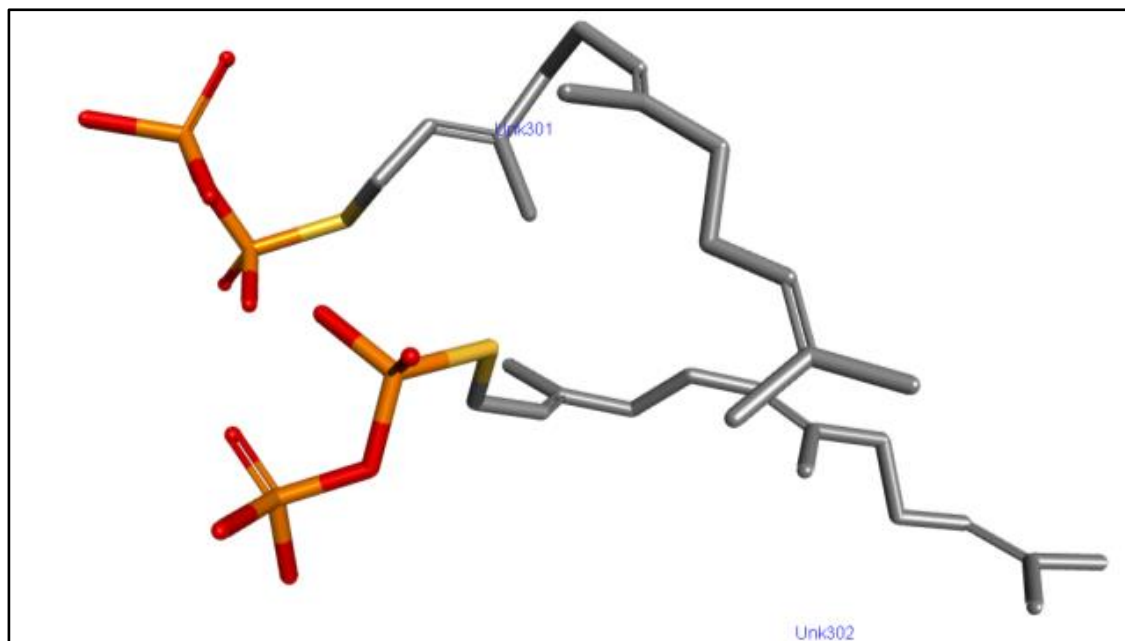


Figure 4.2. Superimpose structure of docked substrate with its original farnesyl diphosphate coordinates in the complex with CrtM

The dock scores of the inhibitor (830) and the substrate (farnesyl-farnesyl diphosphate) of CrtM (PDB ID: 2ZY1) was -9.1 and -11.5 kcal/mol respectively when docked to the active site of CrtM (3W7F), the inhibitor interact with residues (Ala134, Arg171, Arg265, Asn168, Gly161, His18, Leu141, Leu164, Lys20, Phe22, Ser19, Ser21, Tyr248, Tyr41, Val137, Val37) as illustrated in (Figure 4.3). Higher dock score of substrate to CrtM is derived from the number and type of interactions (Ala134, Ala157, Arg171, Arg265, Arg45, Asn168, Asp172, Asp48, Cys44, Gln165, Gly138, Gly161, His18, Leu141, Leu145, Leu160, Leu164, Lys20, Met15, Phe22, Phe233, Ser19, Ser21, Tyr248, Tyr41).

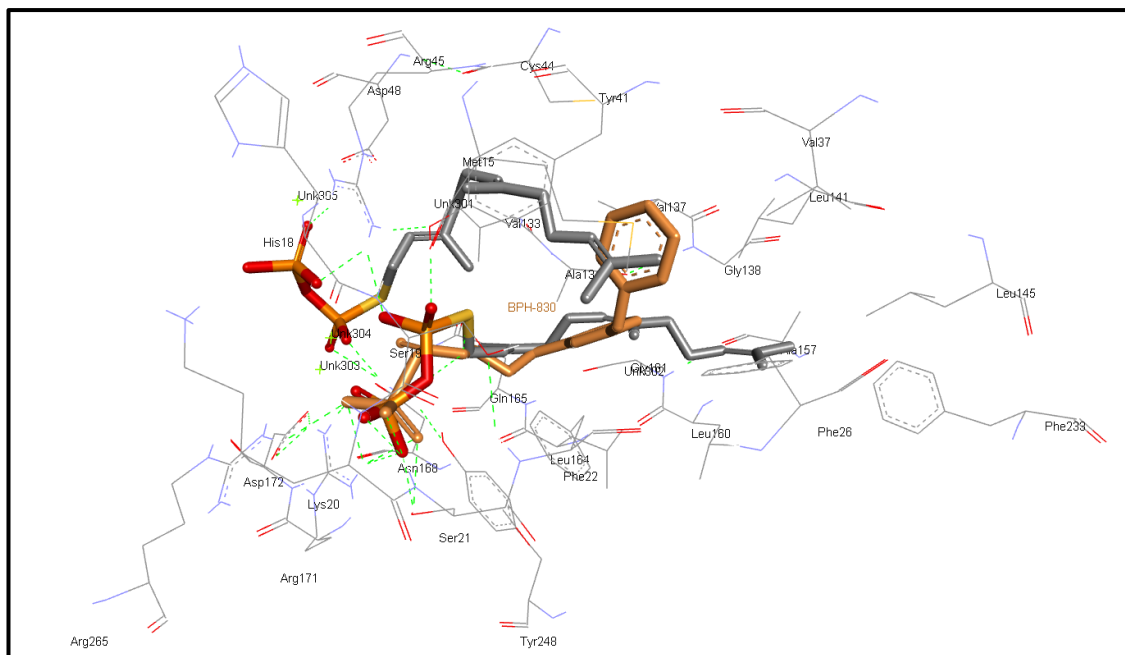
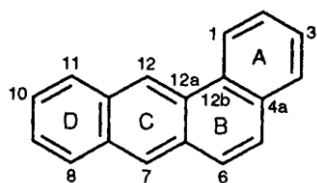
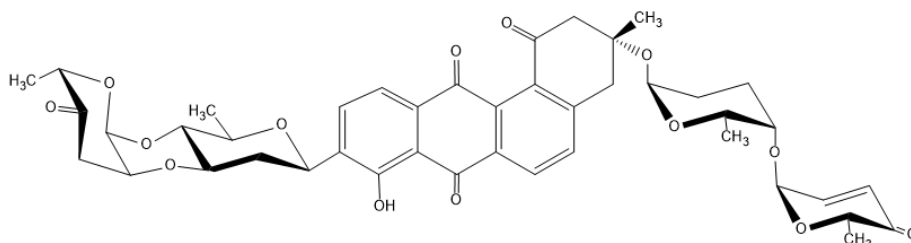


Figure 4.3. Superimpose structure of docked inhibitor BPH-830 with original farnesyl diphosphate coordinates in the complex with CrtM

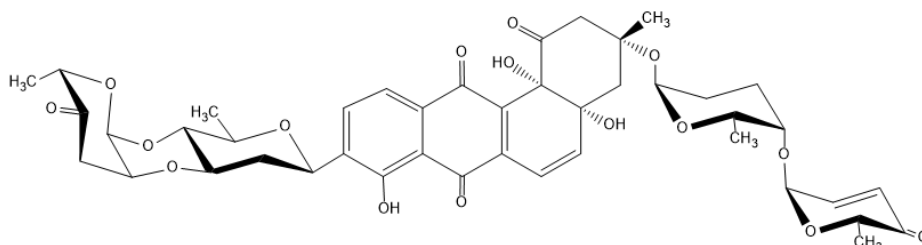
Docking analysis demonstrate that among the attempted compounds; moromycin A (56), saquayamycin B (58), saquayamycin A (145) and saprolmycin E (29) have good interactions with CrtM with higher dock scores; -14.8, -14.4, -13.7, and -13.7 kcal/mol, respectively, when compared with substrate farnesyl diphosphate -11.5 kcal/mol as shown in (Figure 4.4). It has been observed in (56) and (58) the dock scores were -14.8 and -14.4 kcal/mol respectively, the dock scores are similar as well as the structure composition almost same and there are four different sugar subunits, but in (58) there are OH group between A and B rings on Carbons (4a and 12b). But in (29) and (145) we get same dock scores -13.7 kcal/mol when we compared between them we found that there are three sugar subunits connected to D ring and the fourth sugar connected to the A ring.



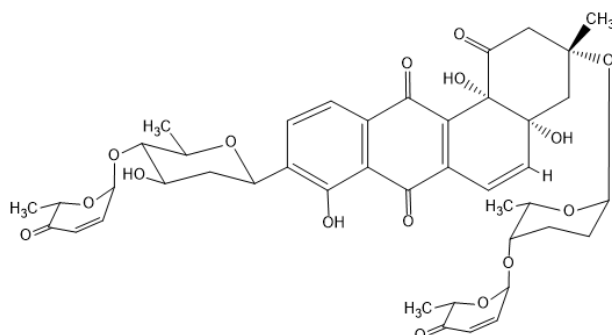
(a) The tetracyclic benz[a]anthracene frame



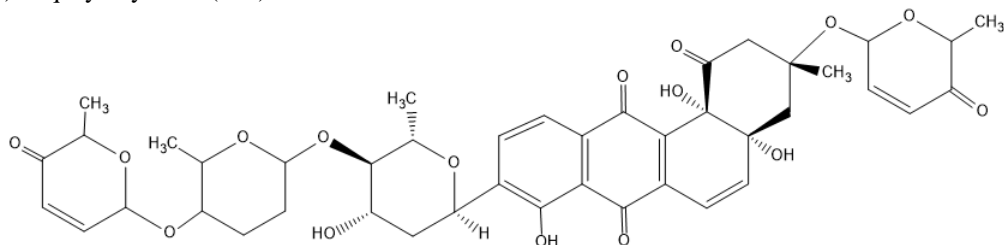
(b) Moromycin A (56) -14.8 kcal/mol



(c) Saquayamycin B (58) -14.4 kcal/mol



(d) Saquayamycin A (145) -13.7 kcal/mol



(e) Saprolymycin E (29) -13.7 kcal/mol

Figure 4.4. Structures of (a) The tetracyclic benz[a]anthracene frame; (b) Moromycin A (56); (c) Saquayamycin B (58); (d) Saquayamycin A (145); and (e) Saprolymycin E (29)

The ligands were grouped in terms of their structure similarity, their locations in the active site of the proteins. The structures with high dock scores were explicitly shown in (Table 4.1, 4.2, 4.3) and (Figures 4.5-4.10, 4.11-4.21, 4.22-4.32) which distributed on 6, 11, 10 groups for CrtM (3ACW) and 3W7F with and without magnesium respectively.

So generally all (157) structures were successfully docked into the proteins and they were all docked into the pocket where original inhibitor and substrate were sockets, and few of them not docked, the residues involved in the interactions of the ligands with proteins are demonstrated in appendices I, II, and III.

The docking scores for the ligands presented in (Table 4.1) and (Figures 4.5 – 4.10) among the ligands, a group of them have higher docking score than the original inhibitor -11.5 kcal/mol such as in Saprolmycin_D (28), N05WA963A (47), N05WA963D (49), BA_12100E (95), BA_12100Z3 (98), which their dock score was -12.9, -12.9, -11.7, -11.7, -11.6 kcal/mol respectively.

4. RESULTS AND DISCUSSION

Table 4.1: Average and maximum dock scores of ligands docked to CrtM (3ACW)

#	Angucycline Group Antibiotics	AV. Kcal/mol	Max. Dock	Interacted Residues No.	Group
0	Ligand of Protein	-11.5	-11.5	14	-
29	Saprolmycin_E	-11.4		13	G1
28	Saprolmycin_D	-12.9	-12.9	12	G1
155	Vineomycin_A1	-11.5	-11.6	15	G1
47	N05WA963A	-12.9	-13.0	14	G2
49	N05WA963D	-11.7	-12.7	15	G2
84	A_7884	-10.7	-11.7	15	G2
88	C104	-10.5	-10.5	12	G2
102	Capoamycin	-10.4	-10.5	12	G2
95	BA_12100E	-11.7	-12.1	13	G3
98	BA_12100Z3	-11.6	-11.8	15	G3
92	BA_12100B	-10.8	-11.7	14	G3
93	BA_12100C	-11.2	-11.7	14	G3
37	Landomycin_Z	-10.2	-11.0	14	G4
94	BA_12100D	-10.2	-10.3	13	G4
11	Rubiginone_B2	-8.5	-8.9	15	G5
54	8_O_methyltetrangulol	-8.6	-8.6	13	G5
118	PD116779	-8.1	-8.3	14	G5
119	X_14881_C	-7.9	-8.2	14	G5
115	Fujianmycin_A	-7.9	-8.0	13	G5
40	JBIR_90	-9.3	-9.7	15	G6
87	C104_Aglycon	-9	-9.6	15	G6
33	Gaudimycin_C	-8.4	-8.5	13	G6
154	Retymicin	-8.2	-8.5	11	G6
133	Homo_UWM6	-8.1	-8.2	11	G6

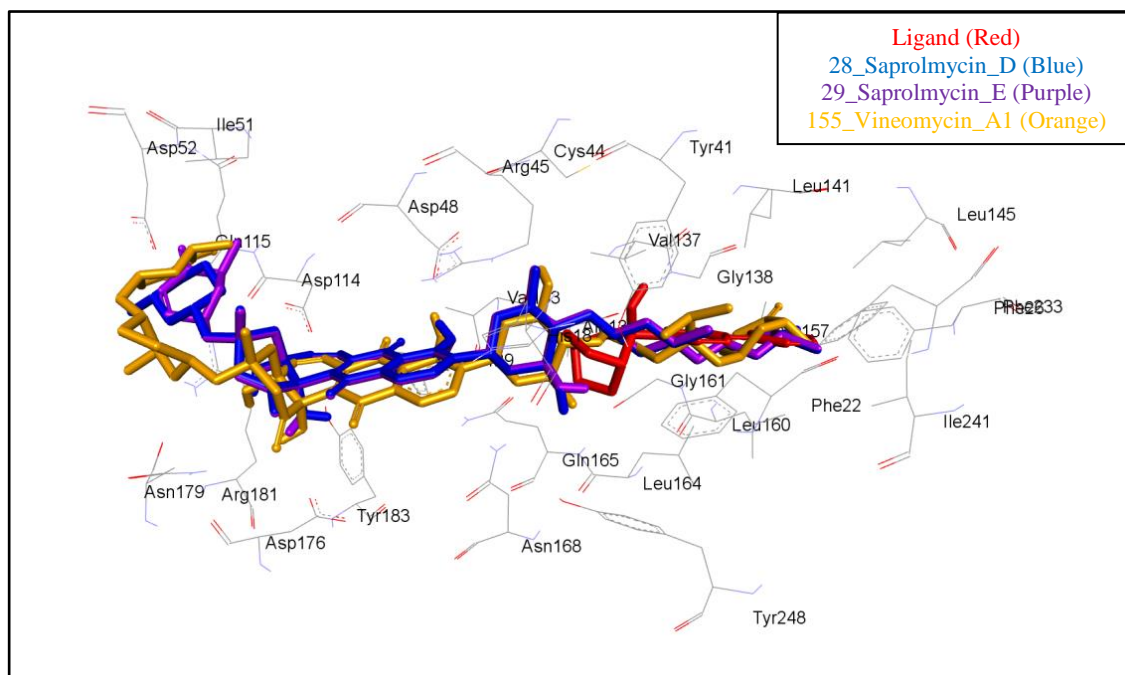


Figure 4.5. Group 1 ligands (28, 29, 155) docked to (3ACW) protein

Saprolmycin_E (29) is the best candidate with score of -13.0 kcal/mol and Vineomycin_A1 (155) is the worst of group 1 with score of -11.6 kcal/mol.

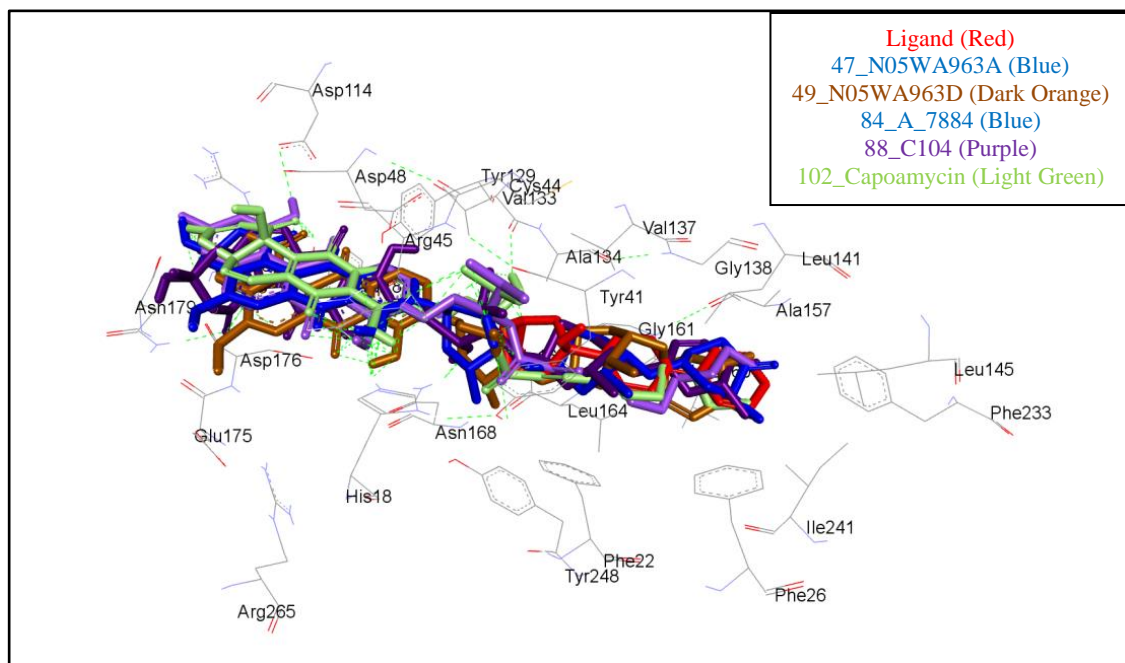


Figure 4.6. Group 2 ligands (47, 49, 84, 88, 102) docked to (3ACW) protein

N05WA963A (47) is the best candidate with score of -13.0 kcal/mol and Capoamycin (102) is the worst of group 2 with score of -10.5 kcal/mol.

4. RESULTS AND DISCUSSION

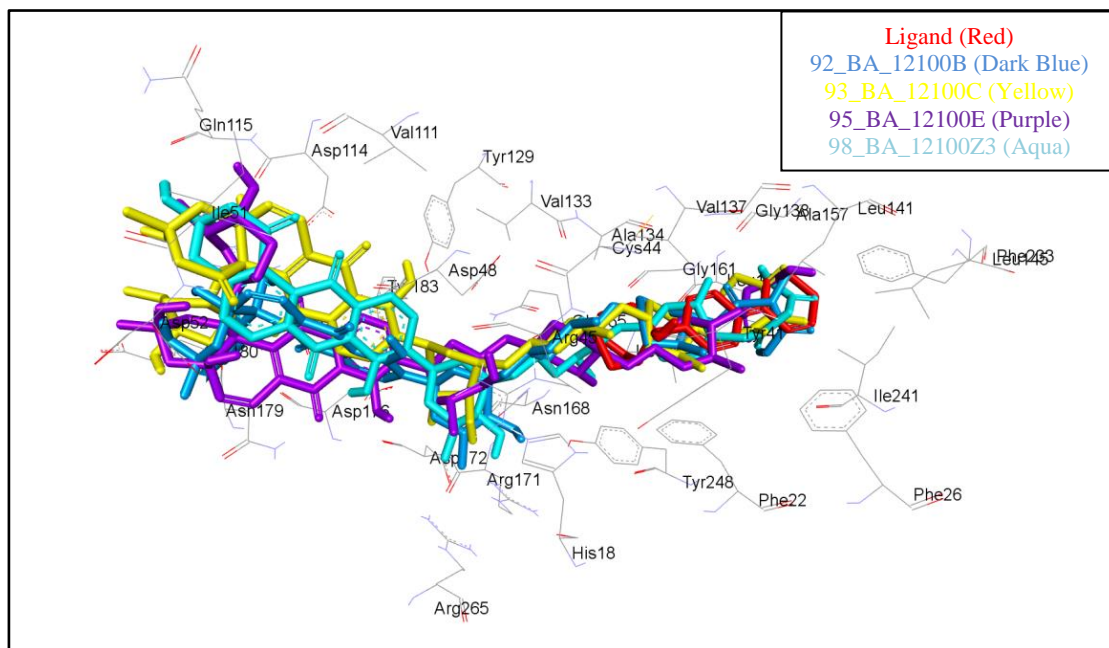


Figure 4.7. Group 3 ligands (92, 93, 95, 98) docked to (3ACW) protein

BA_12100E (95) is the best candidate with score of -12.1 kcal/mol and BA_12100B (92) is the worst of group 3 with score of -11.7 kcal/mol.

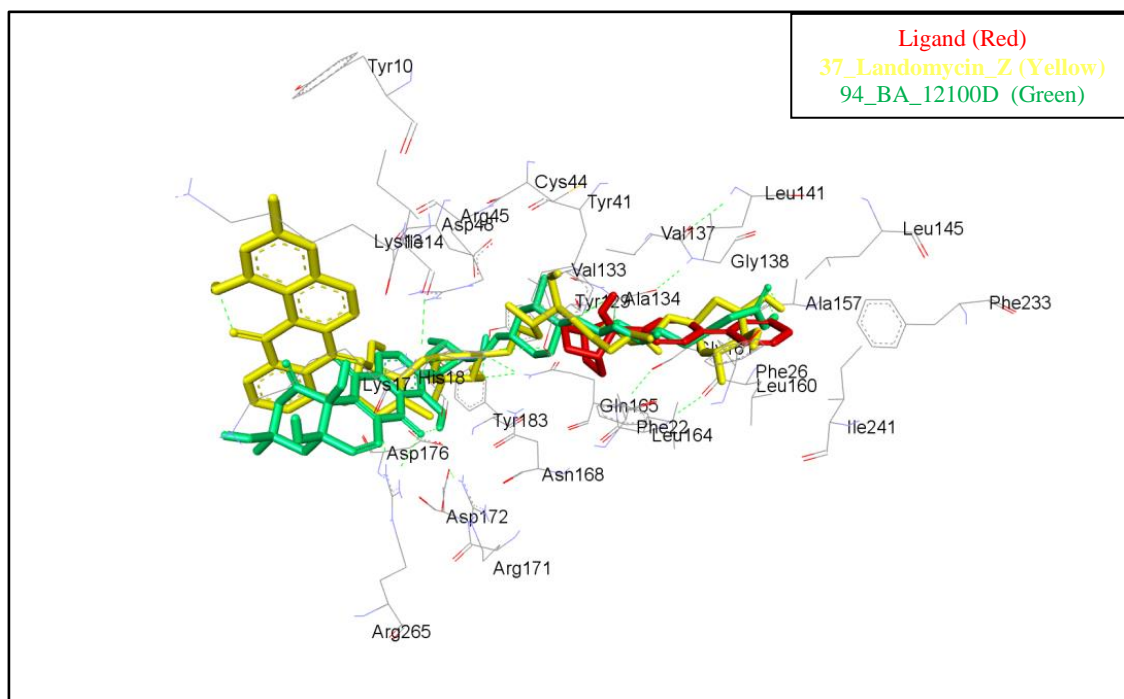


Figure 4.8. Group 4 ligands (37, 94) docked to (3ACW) protein

Landomycin_Z (37) is the best candidate with score of -11.0 kcal/mol and BA_12100D (94) is the worst of group 4 with score of -10.3 kcal/mol.

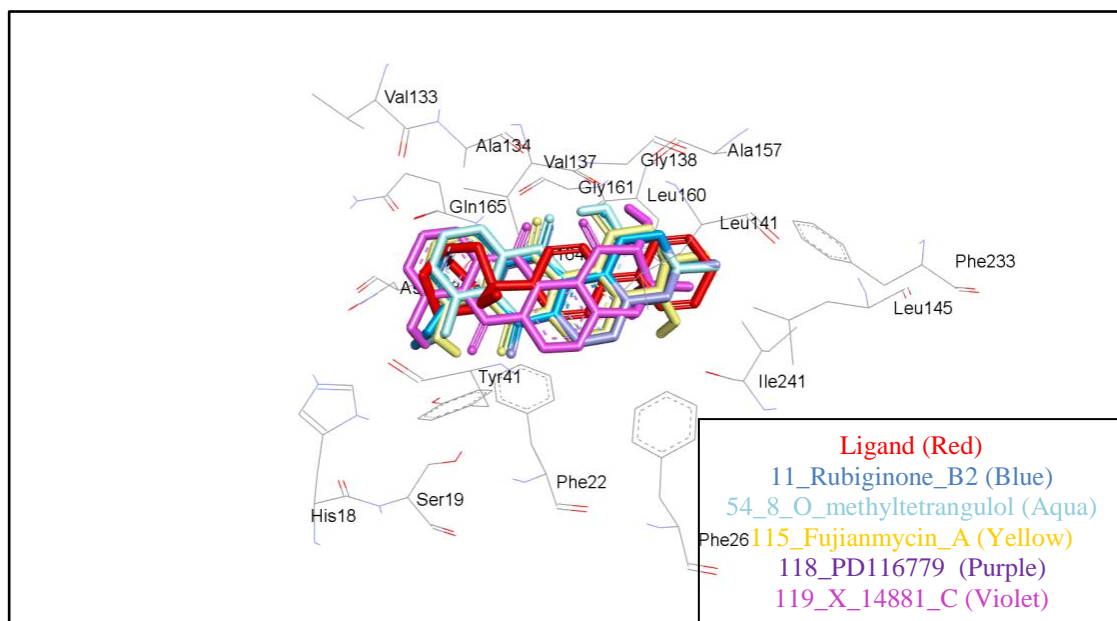


Figure 4.9. Group 5 ligands (11, 54, 115, 118, 119) docked to (3ACW) protein

Rubiginone_B2 (11) is the best candidate with score of -8.9 kcal/mol and Fujianmycin_A (115) is the worst of group 5 with score of -8.0 kcal/mol.

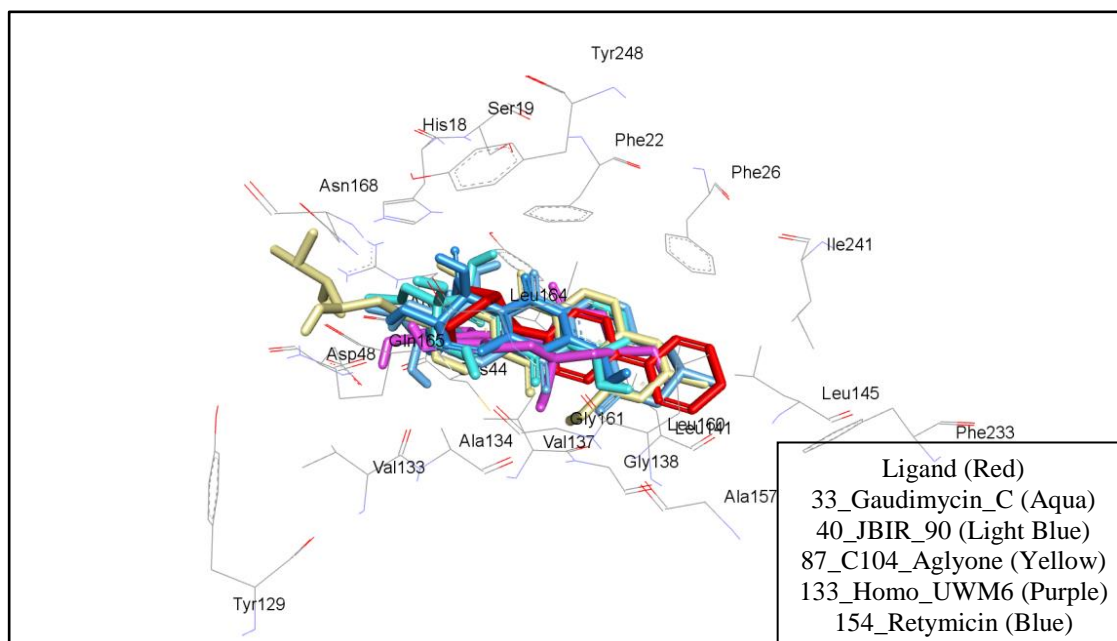


Figure 4.10. Group 6 ligands (33, 40, 87, 133, 154) docked to (3ACW) protein

JBIR_90 (40) is the best candidate with score of -9.7 kcal/mol and Homo_UWM6 (133) is the worst of group 6 with score of -8.2 kcal/mol.

4. RESULTS AND DISCUSSION

The docking scores for the ligands presented in (Table 4.2) and (Figures 4.11 – 4.21) the ligands were grouped in terms of their structure similarity, their locations in the active site of the proteins, all ligands have higher docking score than the original inhibitor -8.3 kcal/mol and the dock scores are in the range of -14.8 to -8.6 kcal/mol for the protein with magnesium for more detail see Appendix II.

Table 4.2. Average of three dock score with maximum dock score of ligands docked to (3W7F) protein with MG.

#	Angucycline Group Antibiotics	AV. kcal/mol	Max. Dock	Interacted Residues No.	Group
0	Ligand of Protein	-8.3	-8.3	22	-
2	Marangucycline_B	-13.4	-13.5	21	G1
98	BA_12100Z3	-11.7	-11.8	19	G1
59	Sch_47554	-11.6	-11.7	18	G1
13	Waldiomycin	-11.4	-11.5	18	G1
29	Saprolmycin_E	-13.5	-13.7	16	G2
28	Saprolmycin_D	-13.5	-13.5	20	G2
38	Grincamycin	-12.5	-12.5	22	G2
30	Saquayamycin_A	-11.2	-11.4	23	G2
19	Fradimycin_C	-11.1	-11.1	17	G2
42	JBIR_91	-12.5	-12.5	19	G3
69	Urdamycin_C	-11.5	-12.2	17	G3
124	Landomycin_C	-11.8	-12.0	26	G3
41	JBIR_116	-10.2	-10.2	19	G3
56	Moromycin_A	-14.8	-14.8	17	G4
58	Saquayamycin_B	-13.7	-14.4	21	G4
47	N05WA963A	-12.5	-12.5	20	G4
49	N05WA963D	-12.2	-12.2	20	G4
134	PI_1894B (Vineomycin A) (OS-4742A1)	-12.7	-12.7	24	G5
65	PI_080 (=PI-6621)	-12.0	-12.1	22	G5
124	Landomycin_C	-11.8	-12.0	26	G5
123	Landomycin_B	-11.3	-11.4	22	G5
93	BA_12100C	-11.0	-11.0	23	G5

Table 4.2. Average of three dock score with maximum dock score of ligands docked to (3W7F) protein with MG. (continue)

#	Angucycline Group Antibiotics	AV. kcal/mol	Max. Dock	Interacted Residues No.	Group
146	Saquayamycin_B	-13.8	-13.8	21	G6
145	Saquayamycin_A	-13.7	-13.7	21	G6
148	Saquayamycin_D	-13.6	-13.6	19	G6
151	Saquayamycin_K4	-12.4	-12.4	19	G6
92	BA_12100B	-11.4	-12.3	19	G6
84	A_7884	-12.0	-12.0	18	G6
147	Saquayamycin_C	-11.8	-11.8	17	G6
6	Hatomarubigin_B	-9.8	-9.8	15	G7
7	Hatomarubigin_C	-9.7	-9.7	16	G7
81	Tetrangulol methyl ether	-9.7	-9.7	11	G7
11	Rubiginone_B2	-9.5	-9.6	14	G7
119	X_14881_C	-9.6	-9.6	14	G7
120	X_14881_E	-9.6	-9.6	12	G7
116	Fujianmycin_B	-9.5	-9.5	14	G7
102	Capoamycin	-9.4	-9.4	17	G7
80	Seitomyacin	-8.9	-9.0	13	G7
31	Gaudimycin_A	-9.8	-9.8	14	G8
33	Gaudimycin_C	-9.5	-9.7	15	G8
32	Gaudimycin_B	-9.6	-9.6	16	G8
137	Urdamycin_J	-9.6	-9.6	15	G8
51	Rabelomycin	-9.3	-9.3	16	G8
62	Tetrangomycin	-9.3	-9.3	16	G8
108	Elmycin_E	-9.6	-9.6	12	G9
110	X_14881_D	-9.6	-9.6	13	G9
55	8_O_methyl_7_deoxo_7_hydroxytetrangomycin	-9.3	-9.3	14	G9
107	Elmycin_D	-9.1	-9.1	11	G9
109	X_14881_B	-9.1	-9.1	13	G9

4. RESULTS AND DISCUSSION

Table 4.2. Average of three dock score with maximum dock score of ligands docked to (3W7F) protein with MG. (continue)

#	Angucycline Group Antibiotics	AV. kcal/mol	Max. Dock	Interacted Residues No.	Group
34	Aquayamycin	-10.3	-10.3	17	G10
4	Actinosporin_B	-8.7	-9.3	18	G10
154	Retymicin	-9.3	-9.3	12	G10
136	Urdamycin_I	-9.2	-9.2	15	G10
86	SM_196 A	-9.8	-9.8	12	G11
115	Fujianmycin_A	-9.5	-9.6	10	G11
118	PD116779	-9.4	-9.5	11	G11
111	Emycin_A	-9.3	-9.4	11	G11

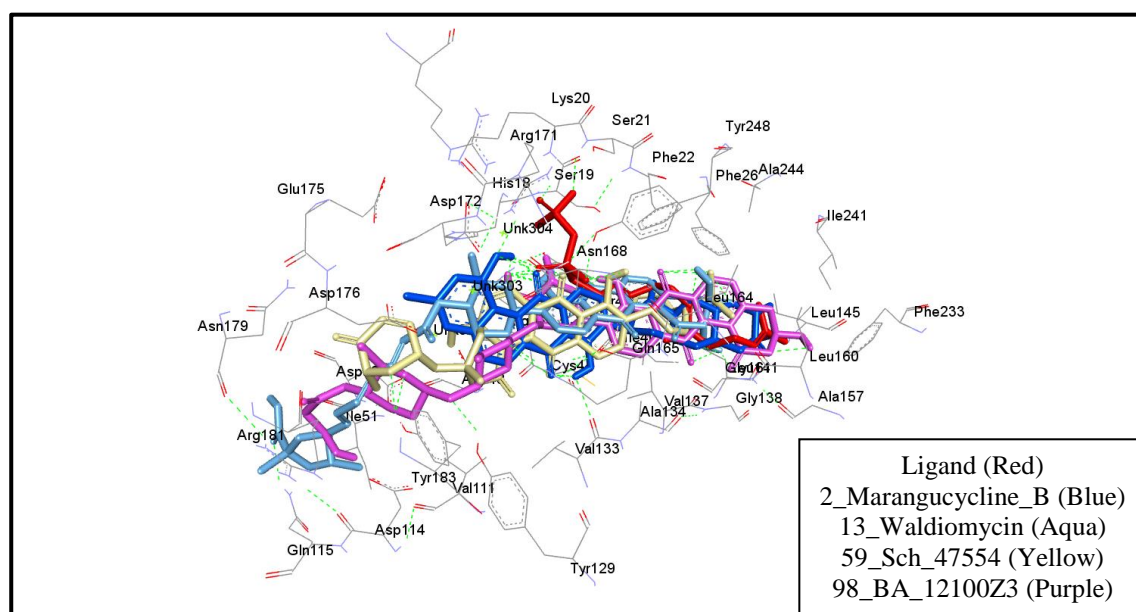


Figure 4.11. Group 1 ligands (2, 13, 59, 98) docked to (3W7F) protein with MG.

Marangucycline_B (2) is the best candidate with score of -13.5 kcal/mol and Waldiomycin (13) is the worst of group 1 with score of -11.5 kcal/mol.

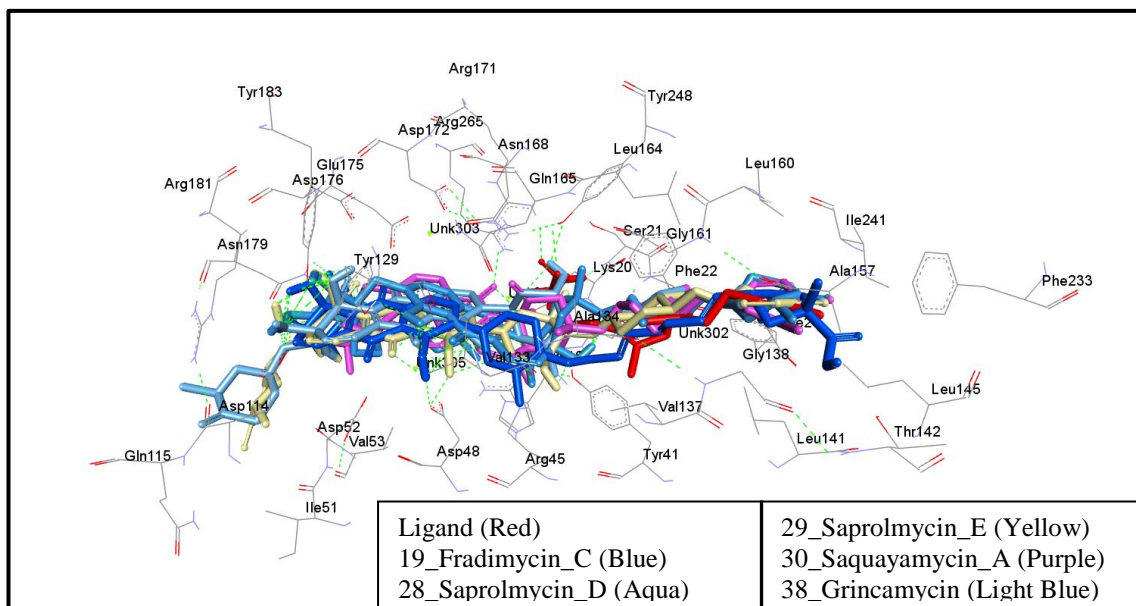


Figure 4.12. Group 2 ligands (19, 28, 29, 30, 38) docked to (3W7F) protein with MG.

Saprolmycin_E (29) is the best candidate with score of -13.7 kcal/mol and Fradimycin_C (19) is the worst of group 2 with score of -11.1 kcal/mol.

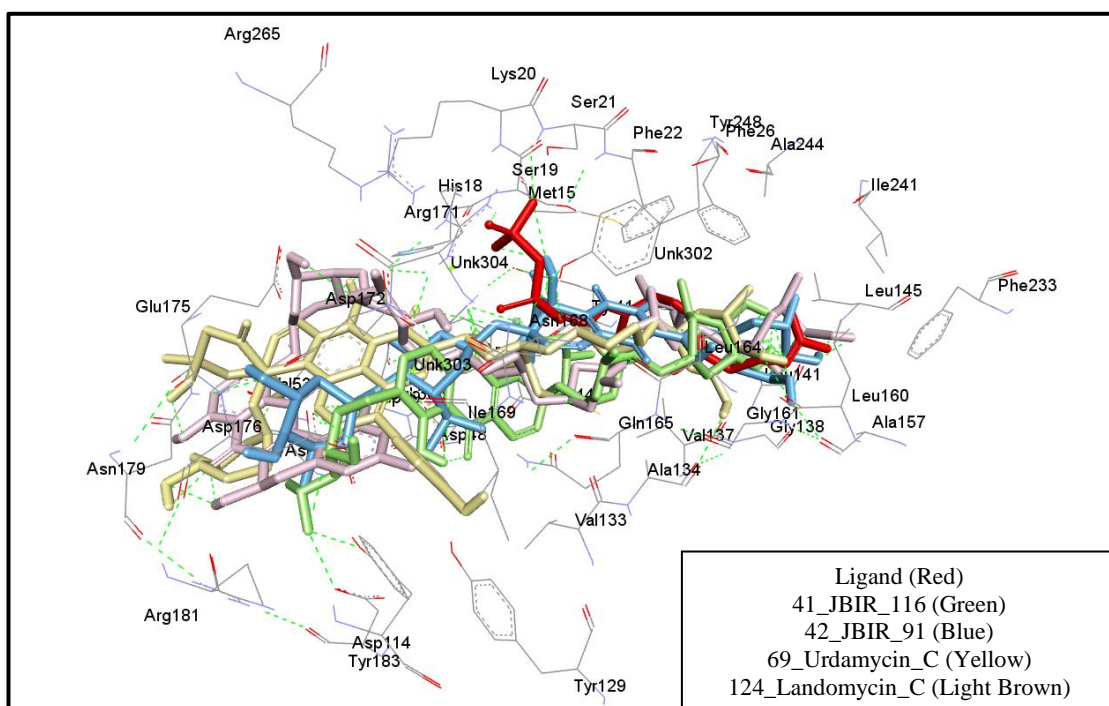


Figure 4.13. Group 3 ligands (41, 42, 69, 124) docked to (3W7F) protein with MG.

JBIR_91 (42) is the best candidate with score of -12.5 kcal/mol and JBIR_116 (41) is the worst of group 3 with score of -10.2 kcal/mol.

4. RESULTS AND DISCUSSION

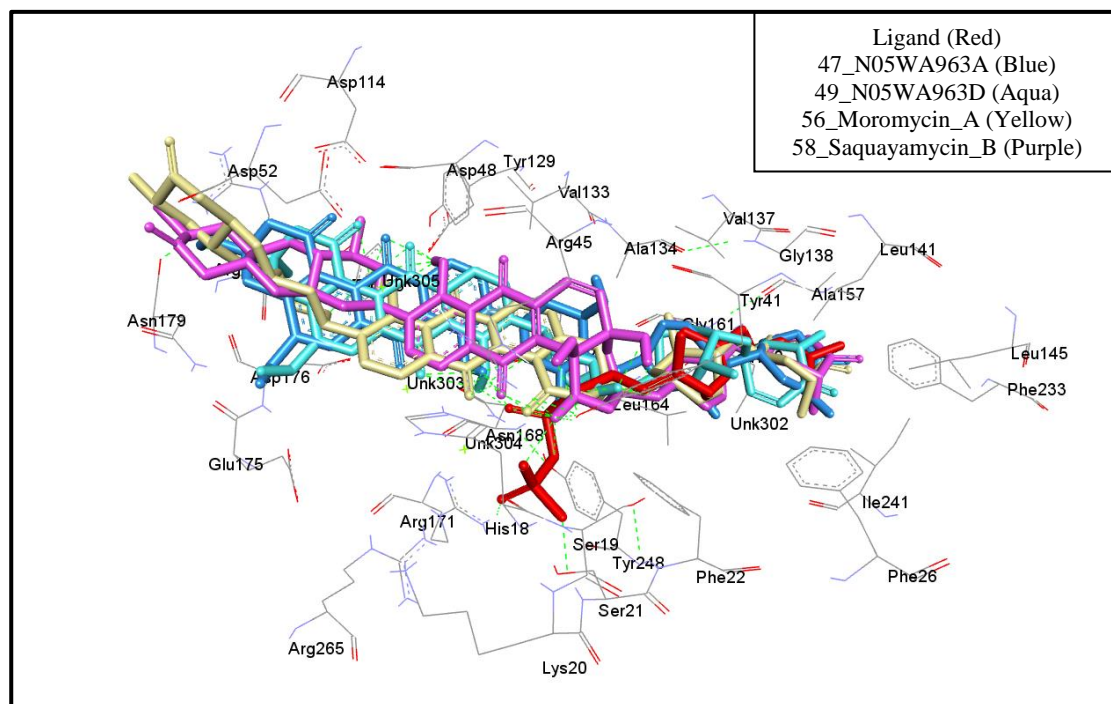


Figure 4.14. Group 4 ligands (47, 49, 56, 58) docked to (3W7F) protein with MG.

Moromycin_A (56) is the best candidate with score of -14.8 kcal/mol and N05WA963D (49) is the worst of group 4 with score of -12.2 kcal/mol.

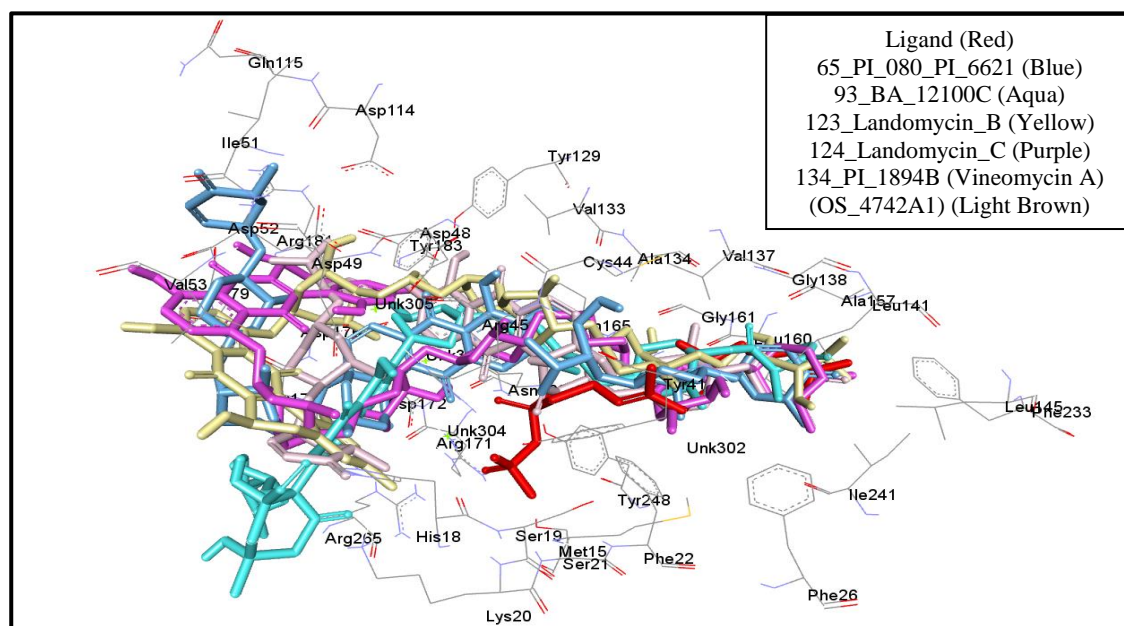


Figure 4.15. Group 5 ligands (65, 93, 123, 124, 134) docked to (3W7F) protein with MG.

PI_1894B (Vineomycin A) (OS-4742A1) (134) is the best candidate with score of -12.7 kcal/mol and BA_12100C (93) is the worst of group 5 with score of -11.0 kcal/mol.

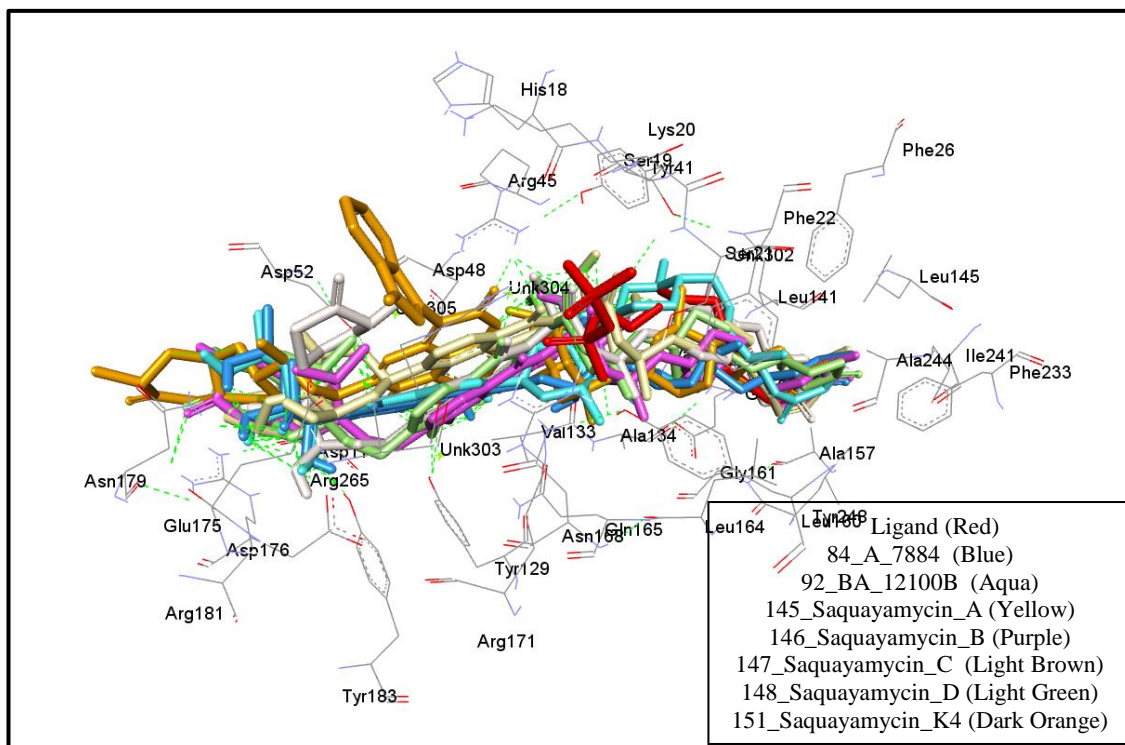


Figure 4.16. Group 6 ligands (84, 92, 145, 146, 147, 148, 151) docked to (3W7F) protein with MG.

Saquayamycin_B (146) is the best candidate with score of -13.8 kcal/mol and Saquayamycin_C (147) is the worst of group 6 with score of -11.8 kcal/mol.

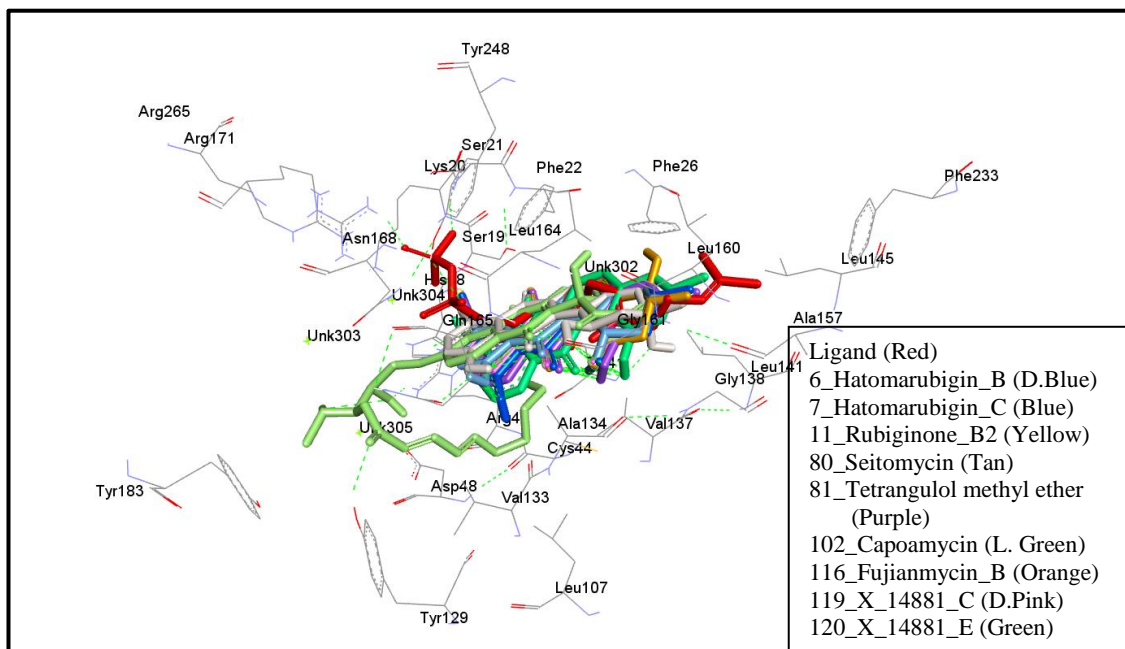


Figure 4.17. Group 7 ligands (6, 7, 11, 80, 81, 102, 116, 119, 120) docked to (3W7F) protein with MG.

Hatamarubigin_B (6) is the best candidate with score of -9.8 kcal/mol and Seitomycin (80) is the worst of group 7 with score of -9.0 kcal/mol.

4. RESULTS AND DISCUSSION

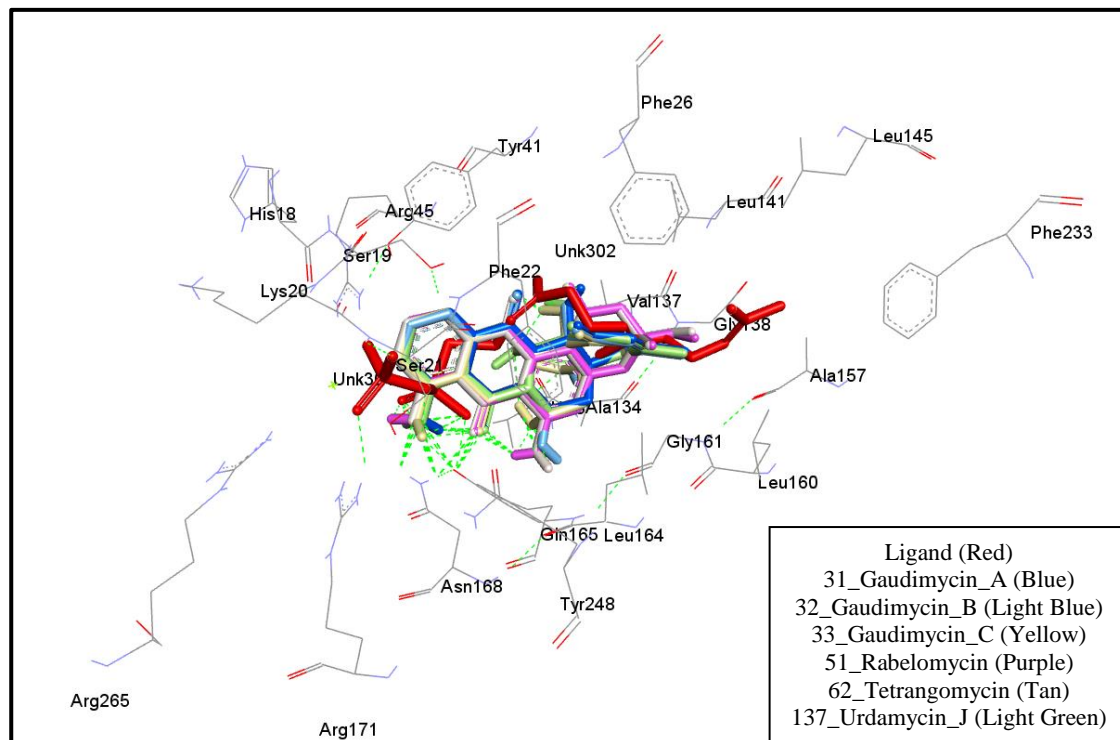


Figure 4.18. Group 8 ligands (31, 32, 33, 51, 62, 137) docked to (3W7F) protein with MG.

Gaudimycin_A (31) is the best candidate with score of -9.8 kcal/mol and Tetrangomycin (62) is the worst of group 8 with score of -9.3 kcal/mol.

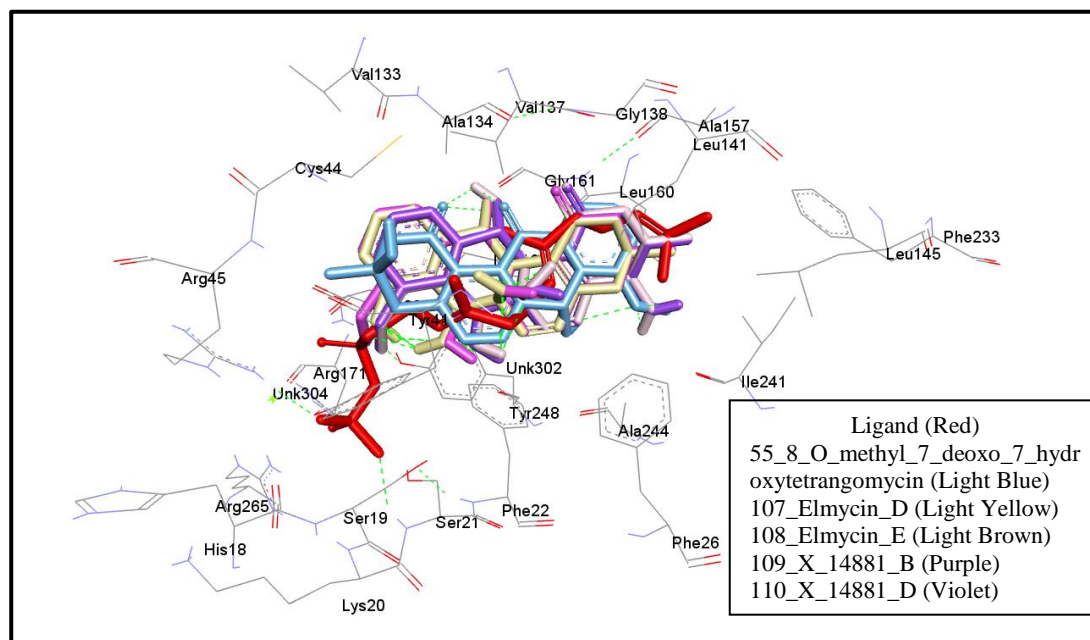


Figure 4.19. Group 9 ligands (55, 107, 108, 109, 110) docked to (3W7F) protein with MG.

Elmycin_E (108) is the best candidate with score of -9.6 kcal/mol and X_14881_B (109) is the worst of group 9 with score of -9.1 kcal/mol.

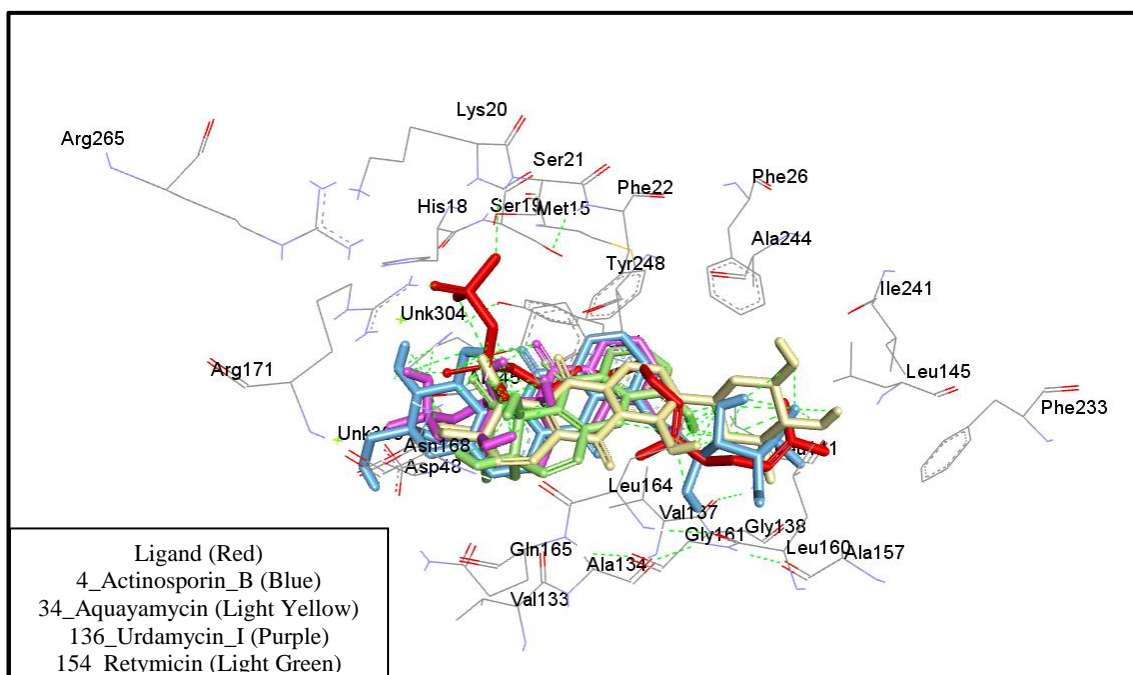


Figure 4.20. Group 10 ligands (4, 34, 136, 154) docked to (3W7F) protein with MG.

Aquayamycin (34) is the best candidate with score of -10.3 kcal/mol and Urdamycin_I (136) is the worst of group 10 with score of -9.2 kcal/mol.

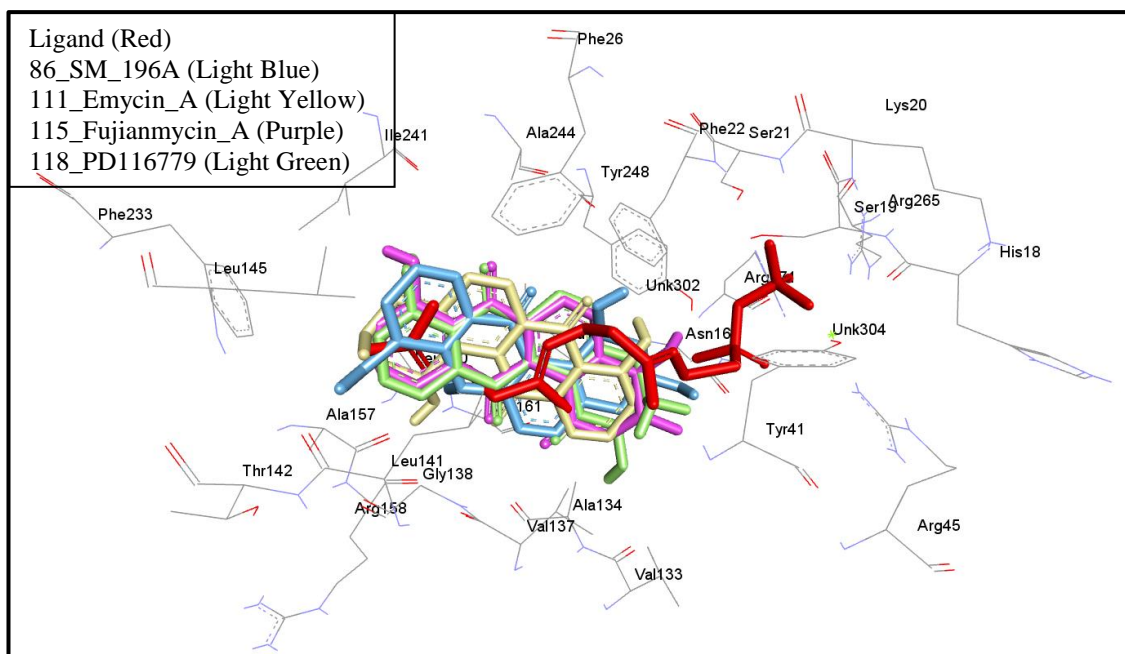


Figure 4.21. Group 11 ligands (86, 111, 115, 118) docked to (3W7F) protein with MG.

SM_196 A (86) is the best candidate with score of -9.8 kcal/mol and Emycin_A (111) is the worst of group 11 with score of -9.4 kcal/mol.

4. RESULTS AND DISCUSSION

The docking scores for the ligands presented in (Table 4.3) and (Figures 4.22 – 4.32) the ligands were grouped in terms of their structure similarity, their locations in the active site of the proteins, all of the ligands have higher docking score than the original inhibitor -8.1 kcal/mol and the dock scores are in the range of -14.3 to -8.7 kcal/mol for the protein without magnesium for more detail see Appendix III.

Table 4.3. Average of three dock score with maximum dock score of ligands docked to (3W7F) protein without MG.

#	Angucycline Group Antibiotics	AV. kcal/mol	Max. Dock	Interacted Residues No.	Group
0	Ligand of Protein	-8.1	-8.3	20	G1
29	Saprolmycin_E	-13.2	-13.3	19	G1
28	Saprolmycin_D	-13.2	-13.2	18	G1
141	Kerriamycin_C	-11.9	-11.9	21	G1
94	BA_12100D	-11.6	-11.7	19	G1
151	Saquayamycin_K4	-14.3	-15.0	24	G2
56	Moromycin_A	-14.4	-14.4	18	G2
146	Saquayamycin_B	-13.6	-13.6	17	G2
148	Saquayamycin_D	-13.5	-13.6	20	G2
134	PI_1894B (Vineomycin A) (OS-4742A1)	-12.6	-12.6	24	G2
155	Vineomycin_A1	-11.9	-12.3	17	G2
38	Grincamycin	-12.2	-12.2	24	G2
147	Saquayamycin_C	-11.7	-12.1	16	G2
46	Urdamycin_B	-12.8	-12.8	17	G3
47	N05WA963A	-12.6	-12.6	14	G3
49	N05WA963D	-12.3	-12.4	13	G3
143	PI_085	-12.6	-12.7	18	G4
144	PI_087	-12.4	-12.5	19	G4
42	JBIR_91	-12.6	-12.7	20	G5
34	Aquayamycin	-10	-10.1	20	G5

Table 4.3. Average of three dock score with maximum dock score of ligands docked to (3W7F) protein without MG. (continue)

#	Angucycline Group Antibiotics	AV. kcal/mol	Max. Dock	Interacted Residues No.	Group
6	Hatomarubigin_B	-9.7	-9.7	15	G6
88	C104	-9.6	-9.7	14	G6
7	Hatomarubigin_C	-9.6	-9.6	16	G6
81	Tetrangulol methyl ether	-9.6	-9.6	11	G6
11	Rubiginone_B2	-9.5	-9.5	14	G6
119	X_14881_C	-9.5	-9.5	14	G6
120	X_14881_E	-9.5	-9.5	12	G6
116	Fujianmycin_B	-9.4	-9.4	14	G6
80	Seitomycin	-8.8	-8.8	15	G6
31	Gaudimycin_A	-9.7	-9.7	15	G7
33	Gaudimycin_C	-9.7	-9.7	14	G7
32	Gaudimycin_B	-9.6	-9.6	15	G7
51	Rabelomycin	-9.3	-9.3	15	G7
62	Tetrangomycin	-9.3	-9.3	15	G7
82	SM_196B	-9.2	-9.2	16	G7
108	Elmycin_E	-9.5	-9.5	14	G8
110	X_14881_D	-9.5	-9.5	13	G8
107	Elmycin_D	-9.1	-9.1	13	G8
109	X_14881_B	-9.1	-9.1	15	G8
115	Fujianmycin_A	-9.6	-9.6	12	G9
111	Emycin_A	-9.4	-9.5	13	G9
118	PD116779	-9.4	-9.4	14	G9
137	Urdamycin_J	-9.5	-9.5	13	G10
156	Azicemicin_A	-9.1	-9.1	18	G10
157	Azicemicin_B	-9.0	-9.0	22	G10

4. RESULTS AND DISCUSSION

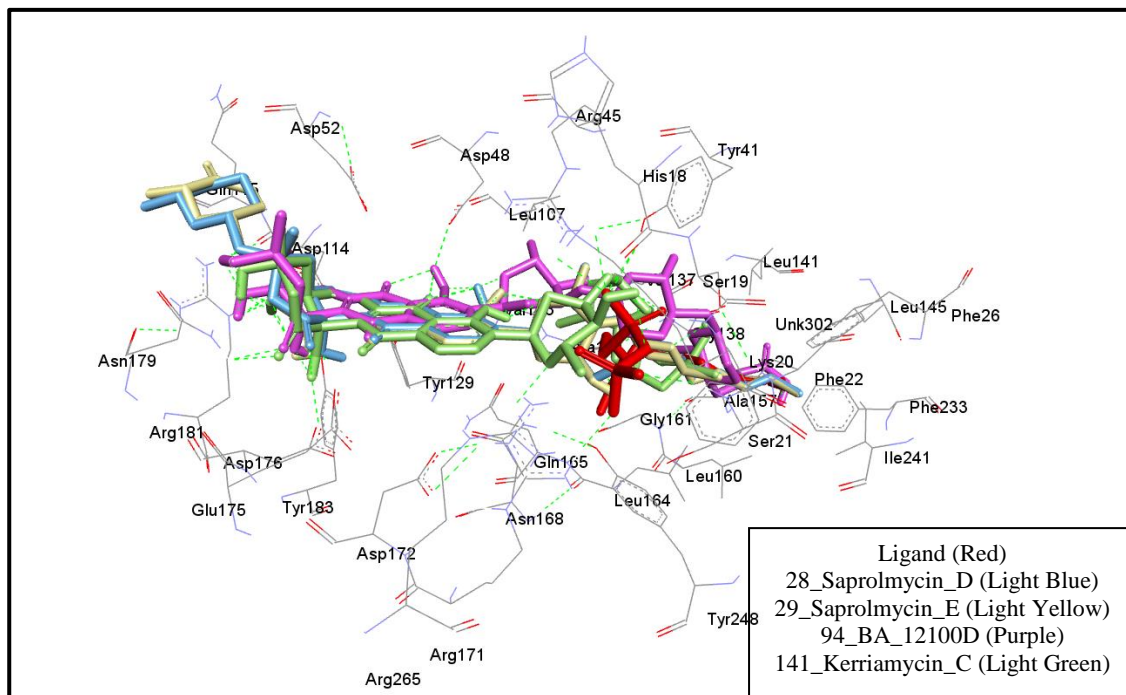


Figure 4.22. Group 1 ligands (28, 29, 94, 141) docked to (3W7F) protein without MG.

Saprolmycin_E (29) is the best candidate with score of -13.3 kcal/mol and BA_12100D (94) is the worst of group 1 with score of -11.7 kcal/mol.

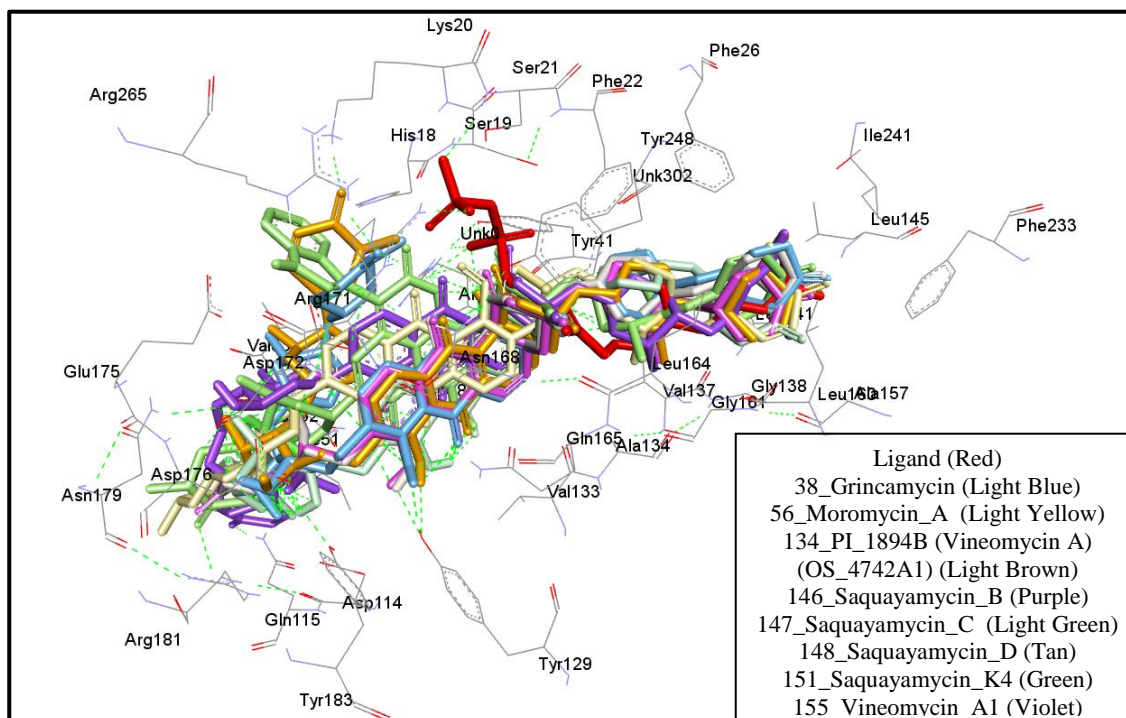


Figure 4.23. Group 2 ligands (38, 56, 134, 146, 147, 148, 151, 155) docked to (3W7F) protein without MG.

Saquayamycin_K4 (151) is the best candidate with score of -15.0 kcal/mol and Saquayamycin_C (147) is the worst of group 2 with score of -12.1 kcal/mol.

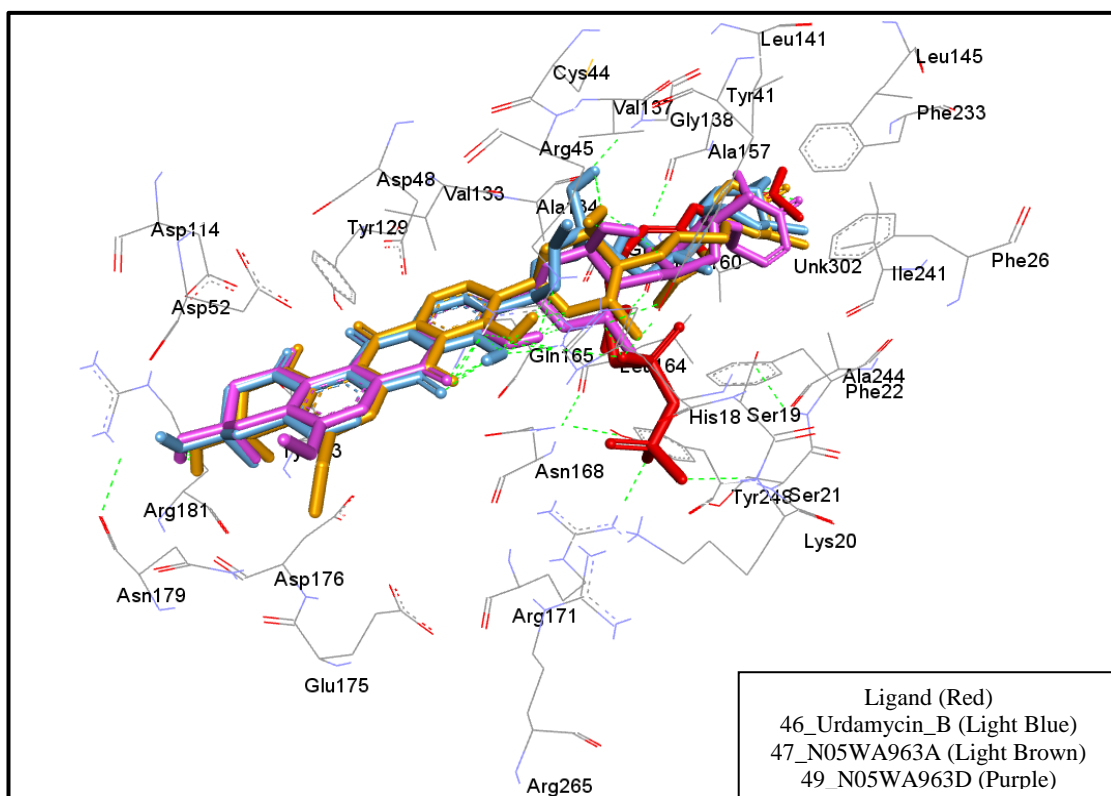


Figure 4.24. Group 3 ligands (46, 47, 49) docked to (3W7F) protein without MG.

Urdamycin_B (46) is the best candidate with score of -12.8 kcal/mol and N05WA963D (49) is the worst of group 3 with score of -12.4 kcal/mol.

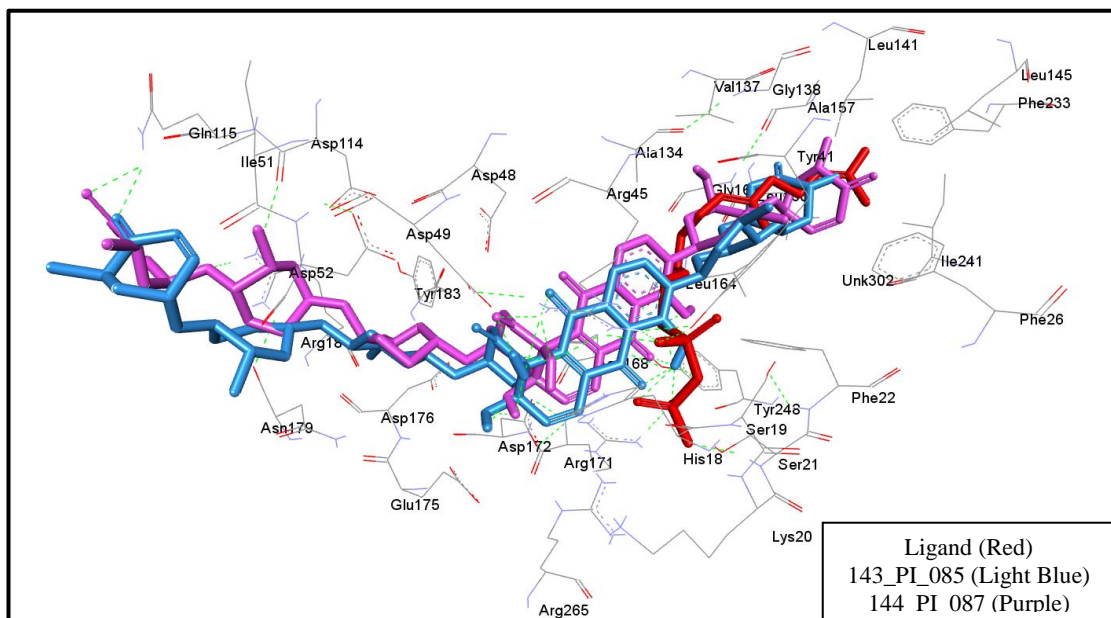


Figure 4.25. Group 4 ligands (143_PI_085, 144_PI_087) docked to (3W7F) protein without MG.

PI_085 (143) is the best candidate with score of -12.7 kcal/mol and PI_087 (144) is the worst of group 4 with score of -12.5 kcal/mol.

4. RESULTS AND DISCUSSION

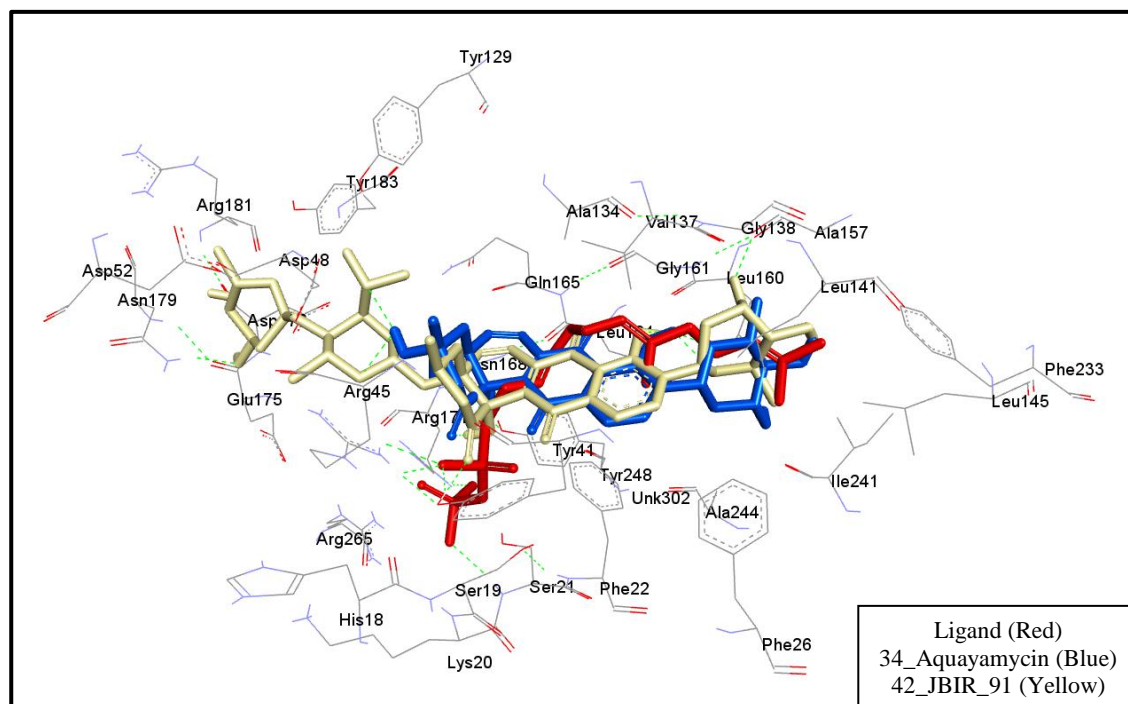


Figure 4.26. Group 5 ligands (34, 42) docked to (3W7F) protein without MG.

JBIR_91 (42) is the best candidate with score of -12.7 kcal/mol and Aquayamycin (34) is the worst of group 5 with score of -10.1 kcal/mol.

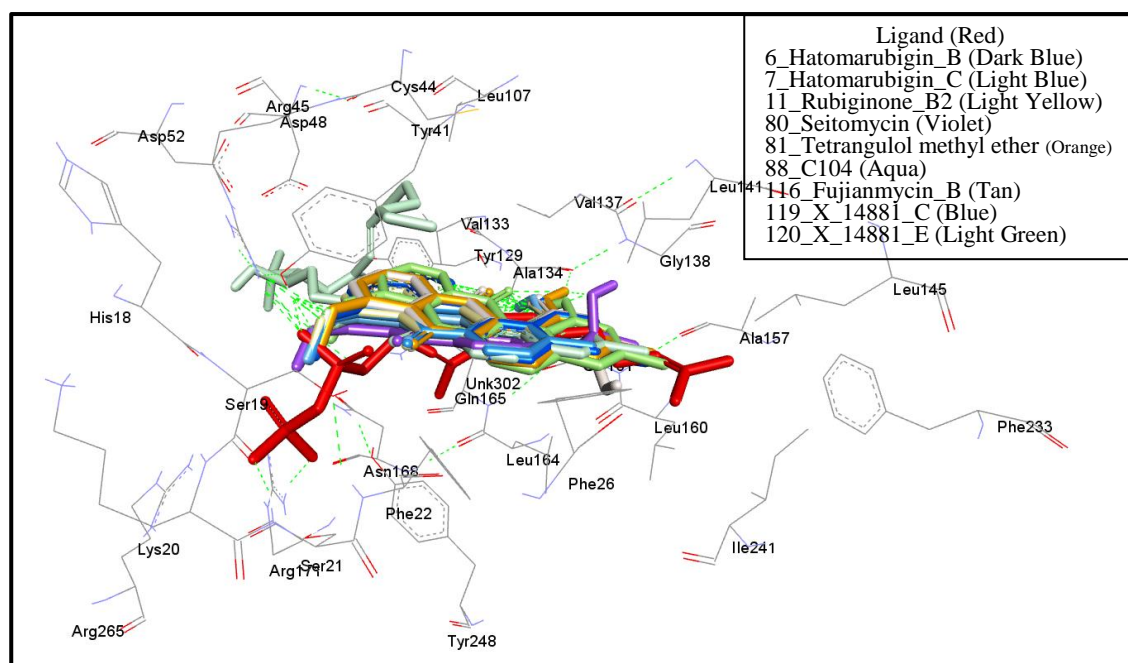


Figure 4.27. Group 6 ligands (6, 7, 11, 80, 81, 88, 116, 119, 120) docked to (3W7F) protein without MG.

Hatomarubigin_B (6) is the best candidate with score of -9.7 kcal/mol and Seitomycin (80) is the worst of group 6 with score of -8.8 kcal/mol.

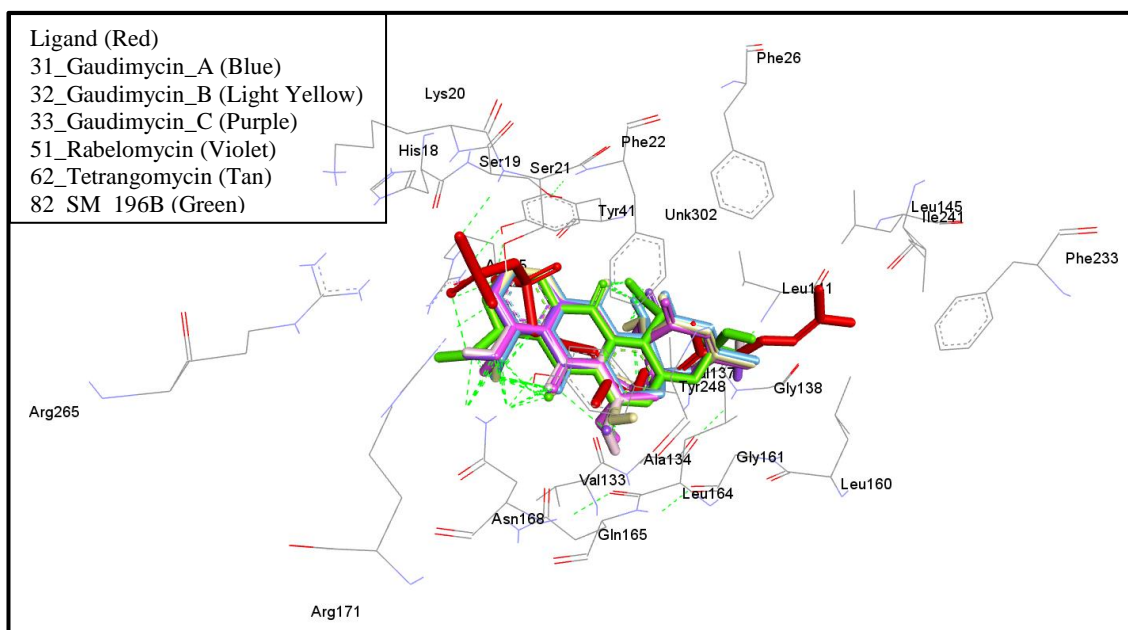


Figure 4.28. Group 7 ligands (31, 32, 33, 51, 62, 82) docked to (3W7F) protein without MG.

Gaudimycin_A (31) is the best candidate with score of -9.7 kcal/mol and SM_196B (82) is the worst of group 7 with score of -9.2 kcal/mol.

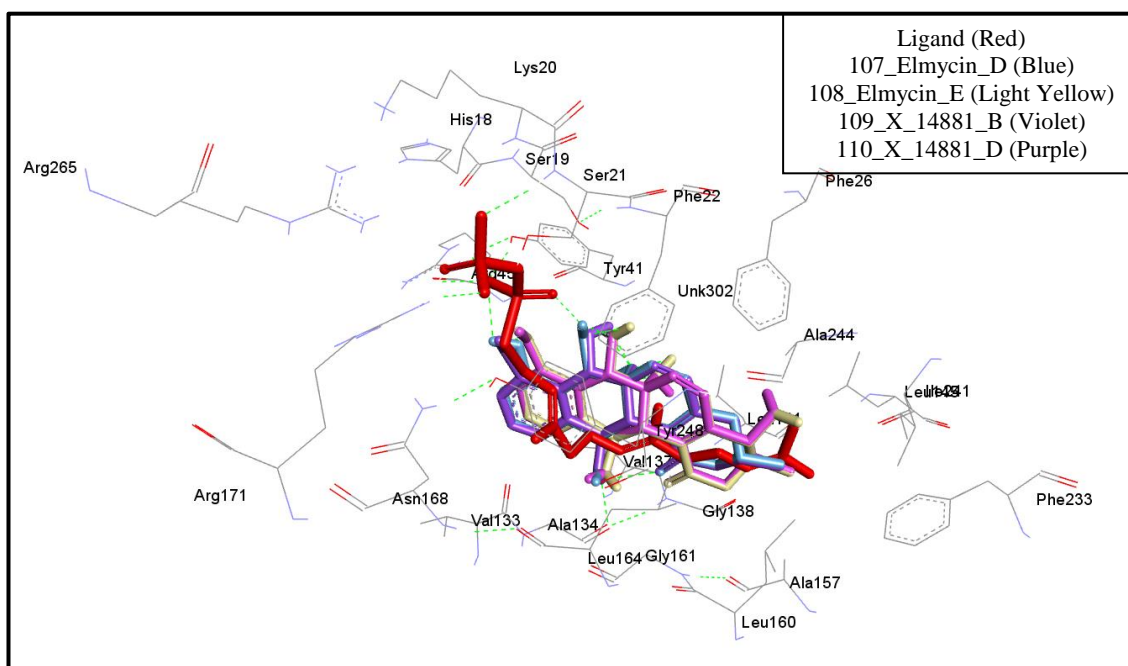


Figure 4.29. Group 8 ligands (107, 108, 109, 110) docked to (3W7F) protein without MG.

Elmycin_E (108) is the best candidate with score of -9.5 kcal/mol and X_14881_B (109) is the worst of group 8 with score of -9.1 kcal/mol.

4. RESULTS AND DISCUSSION

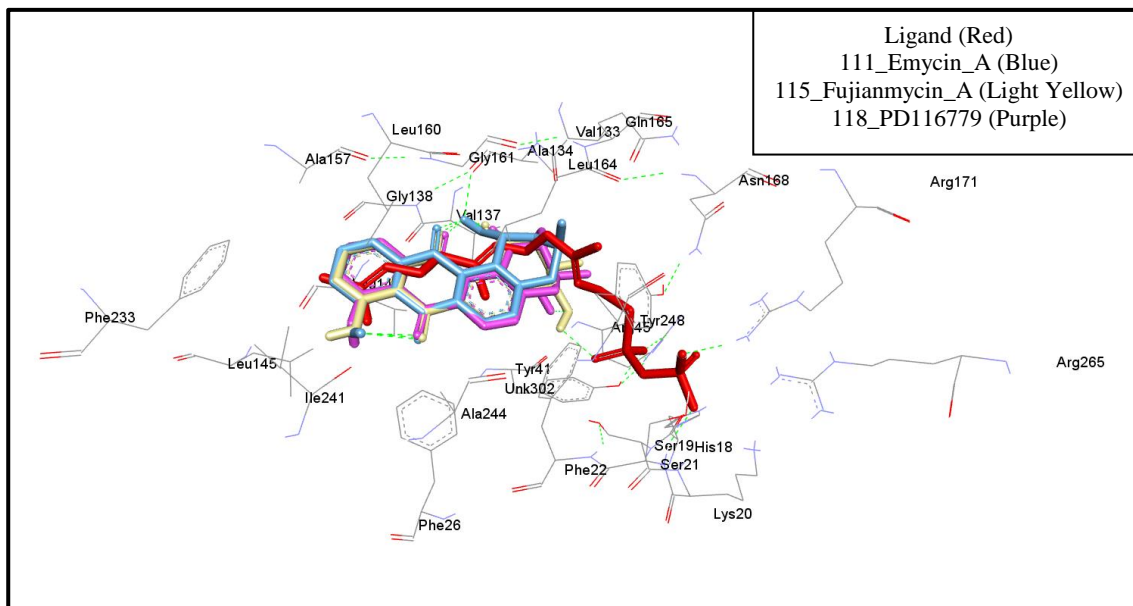


Figure 4.30. Group 9 ligands (111, 115, 118) docked to (3W7F) protein without MG.

Fujianmycin_A (115) is the best candidate with score of -9.6 kcal/mol and PD116779 (118) is the worst of group 9 with score of -9.4 kcal/mol.

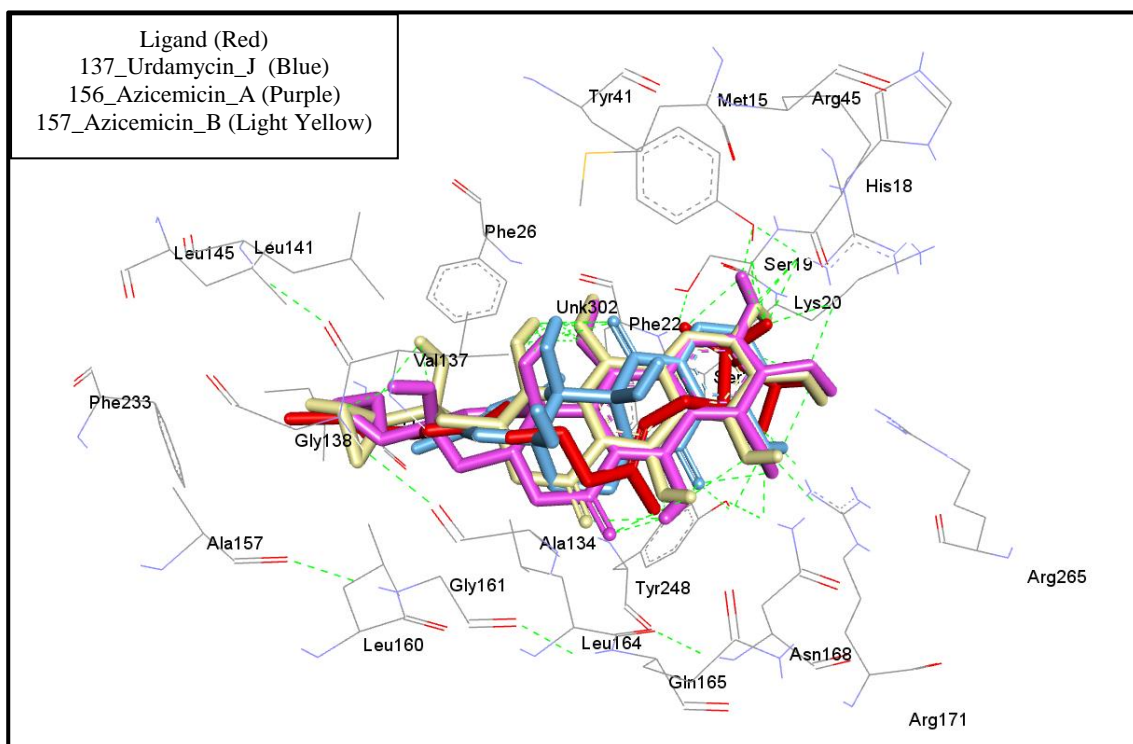


Figure 4.31. Group 10 ligands (137, 156, 157) docked to (3W7F) protein without MG.

Urdamycin_J (137) is the best candidate with score of -9.5 kcal/mol and Azicemicin_B (157) is the worst of group 10 with score of -9.0 kcal/mol.

Sakai *et al.* (2012) reported that the first member of angucycline compound tetrangomycin was found to inhibit the staphyloxanthin production by MRSA in the paper disk assay. However, the potential therapeutic target of tetrangomycin in the biosynthesis of staphyloxanthin was not clear, as reported by Ozakin *et al.* (2016) in order to investigate the potential molecular targets for tetrangomycin the molecular docking was performed using dehydrosqualene synthase enzyme (PDB ID: 2ZY1) which is catalyzed the first committed step of staphyloxanthin biosynthesis. Total dock scores were -49.55 for inhibitor 830 and -33.71 for tetrangomycin. The means of, the dock score of inhibitor 830 was higher than the dock score of tetrangomycin. Also in our study we get the same result regarding the tetrangomycin. At the same time when it is performed molecular docking using Autodock Vina for all angucycline compounds (157), it has been observed that we get higher dock scores as shown in the Appendices I, II, III, and IV, and this is because of most of the angucycline structures have large chemical structure that help them in interacting with residues that are presents in active site, and because most of interaction are electrostatic and hydrophobic as a result gives higher dock scores.

Molecular docking study was done by Kahlon *et al.* (2010) to find the binding site of SQS inhibitors within the dehydrosqualene synthase of *S. aureus* to inhibit its function. They used lapaquistat acetate and squalestatin analogs reported as squalene synthase (SQS) inhibitors in human and animal models but not studied in context to CrtM of *S. aureus*. From the frequency of residue occurrence in the formation of hydrogen bonding with inhibitors studied, it was found that the Arg45, Asp48, Asp52, Asp172, Asn168, Gln165, His18, Tyr129 are interacted comparatively with higher frequency. In our study the interacted residues of CrtM involved in the interactions with the angucycline compounds was (Ala134, Ala157, Arg171, Arg265, Arg45, Asn168, Gln165, Gly138, Gly161, His18, Leu141, Leu145, Leu160, Leu164, Lys20, Phe22, Phe233, Ser19, Ser21, Tyr248, Tyr41) It seems that Arg45, Arg171, Arg265 are the main residues for the substrate.

5 CONCLUSIONS

Docking results produced valuable information not just regarding potential use of a large class of actinomycetes based compounds as candidates against MRSA.

Docking analysis demonstrate that among the attempted compounds; Moromycin_A (56), Saquayamycin_B (58), Saquayamycin_B (146), Saquayamycin_A (145), Saprolmycin_E (29), Saquayamycin_D (148), Saprolmycin_D (28), Marangucycline_B (2), PI_1894B (Vineomycin A) (134), JBIR_91 (42), Grincamycin (38), Saquayamycin_K4 (151) are best candidates with scores of -14.8, -14.4, -13.8, -13.7, -13.7, -13.6, -13.5, -13.5, -12.7, -12.5, -12.5, 12.4 kcal/mol respectively.

However, *in vitro* tests are required to have further study regarding their activities.

It is also necessary to carry out Molecular Dynamic Simulations to have a precise information regarding the stability and dynamic behaviors of the complexes, since molecular dynamic calculations produce data to calculate binding free energies of the complexes, which are comparable with those obtained by isothermal titration calorimetry (ITC) values, which is one of the best method, determining experimental binding free energies between a receptor and a ligand.

5. CONCLUSION

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APPENDIX I
Docking score and interacted residue of protein (3ACW)
with the Angucycline Group Antibiotics (ligands), and the arrangements are according to dock scores (kcal/mol)

#	Compound Name Docked to Protein (3ACW)	Residues that interacts with the original ligand & Residues that interacts with the ligands					Run1	Run2	Run3	AV. Dock	Ligand of Protein	Saquayamycin_B	N05WA963A	Saprolmycin_E	Saprolmycin_D	Moromycin_A	N05WA963D	PI_085	Grincamycin_F	Saquayamycin_D
		Residues that NOT interacts with the original ligand																		
0	3ACW with its ligand From PDB						-11.5	-12.1	-12.9	-13.0	-12.9	-13.0	-13.0	-12.9	-12.1	-12.7	-12.7	-12.7	-12.6	-12.4
58							-11.5	-12.0	-12.9	-10.6	-10.6	-12.9	-12.7	-12.7	-12.7	-9.9	-10.5	-12.6	-12.3	
47							-11.5	-13.3	-13.0	-10.6	-10.6	-12.9	-12.9	-12.9	-12.7	-12.6	-11.2	-11.4	-12.3	
29							-11.5	-12.5	-12.9	-11.4	-11.4	-12.9	-12.5	-12.5	-11.7	-11.5	-12.2	-12.3		
28							-11.5	-13.3	-13.0	-13.0	-13.0	-12.9	-12.7	-12.7	-12.7	-12.7	-12.7	-12.7	-12.6	
56							14	14	10	14	13	12	9	9	15	9	15	15	8	
49							Ala134	Ala134	Ala134	Ala134	Ala157	Ala157	Ala157	Ala157	Ala157	Ala157	Ala157	Ala157	Ala157	
143							Asn168	Asn168	Asn168	Asn168	Asn168	Asn168	Asn168	Asn168	Asn168	Asn168	Asn168	Asn168	Asn168	
39							Gln165	Gln165	Gln165	Gln165	Gln165	Gln165	Gln165	Gln165	Gln165	Gln165	Gln165	Gln165	Gln165	
148							Gly138	Gly138	Gly138	Gly138	Gly138	Gly138	Gly138	Gly138	Gly138	Gly138	Gly138	Gly138	Gly138	
							Gly161	Gly161	Gly161	Gly161	Gly161	Gly161	Gly161	Gly161	Gly161	Gly161	Gly161	Gly161	Gly161	
							His18	His18	His18	His18	His18	His18	His18	His18	His18	His18	His18	His18	His18	
							Ile241	Ile241	Ile241	Ile241	Ile241	Ile241	Ile241	Ile241	Ile241	Ile241	Ile241	Ile241	Ile241	
							Leu141	Leu141	Leu141	Leu141	Leu141	Leu141	Leu141	Leu141	Leu141	Leu141	Leu141	Leu141	Leu141	
							Leu145	Leu145	Leu145	Leu145	Leu145	Leu145	Leu145	Leu145	Leu145	Leu145	Leu145	Leu145	Leu145	
							Leu160	Leu160	Leu160	Leu160	Leu160	Leu160	Leu160	Leu160	Leu160	Leu160	Leu160	Leu160	Leu160	
							Leu164	Leu164	Leu164	Leu164	Leu164	Leu164	Leu164	Leu164	Leu164	Leu164	Leu164	Leu164	Leu164	
							Phe22	Phe22	Phe22	Phe22	Phe22	Phe22	Phe22	Phe22	Phe22	Phe22	Phe22	Phe22	Phe22	
							Phe26	Phe26	Phe26	Phe26	Phe26	Phe26	Phe26	Phe26	Phe26	Phe26	Phe26	Phe26	Phe26	
							Val137	Val137	Val137	Val137	Val137	Val137	Val137	Val137	Val137	Val137	Val137	Val137	Val137	
							Unk1001	Unk1001	Unk1001	Unk1001	Unk1001	Unk1001	Unk1001	Unk1001	Unk1001	Unk1001	Unk1001	Unk1001	Unk1001	
							HOH474	Phe233	Arg45	Arg181	Ala157	Arg181	Arg181	Arg181	Arg181	Arg181	Arg181	Arg181	Ser19	Arg181
								Tyr41	Arg45	Arg181	Arg45	Arg181	Arg45	Arg181	Arg45	Arg181	Arg45	Tyr129	Arg45	
								Val133	Asn179	Arg45	Asn179	Arg45	Asn179	Arg45	Asn179	Arg45	Asn179	Tyr183	Asn179	
									Asp176	Asp176	Asp176	Asp176	Asp176	Asp176	Asp176	Asp176	Asp176	Tyr248	Asp176	
									Asp114	Asp114	Asp114	Asp114	Asp114	Asp114	Asp114	Asp114	Asp114	Tyr248	Asp114	
									Asp48	Asp48	Asp48	Asp48	Asp48	Asp48	Asp48	Asp48	Asp48	Val133	Asp48	
									Gln115	Gln115	Gln115	Gln115	Gln115	Gln115	Gln115	Gln115	Gln115	Gln115	Asp52	
									Gly55	Gly55	Gly55	Gly55	Gly55	Gly55	Gly55	Gly55	Gly55	Gly55	Val171	
									Ile51	Ile51	Ile51	Ile51	Ile51	Ile51	Ile51	Ile51	Ile51	Ile51	Gln115	
									Tyr129	Tyr129	Tyr129	Tyr129	Tyr129	Tyr129	Tyr129	Tyr129	Tyr129	Tyr129	Gly161	
									Tyr183	Tyr183	Tyr183	Tyr183	Tyr183	Tyr183	Tyr183	Tyr183	Tyr183	Tyr183	Arg181	
									Tyr41	Tyr41	Tyr41	Tyr41	Tyr41	Tyr41	Tyr41	Tyr41	Tyr41	Tyr41	Arg265	
									Val133	Val133	Val133	Val133	Val133	Val133	Val133	Val133	Val133	Val133	Tyr129	
																			Arg45	
																			Asn179	
																			Tyr183	
																			Asp114	
																			Tyr248	
																			Asp172	
																			Tyr41	
																			Val133	
																			Asp48	
																			Val133	
																			Asp52	
																			Cys44	
																			Gln115	

APPENDIX I
Docking score and interacted residue of protein (3ACW)
with the Angucycline Group Antibiotics (ligands), and the arrangements are according to dock scores (kcal/mol) (continue)

#	Compound Name Docked to Protein (3ACW)	Run1	Run2	Run3	AV.	Dock	Residues that interacts with the original ligand & Residues that interacts with the ligands	Residues that NOT interacts with the original ligand
146	Saquayamycin_B	-12.2	-12.1	-12.0	-12.1	-12.2	Val137	
145	Saquayamycin_A	-11.8	-12.1	-12.2	-12.0	-12.2	Val137	
65	PI_080 (=PI-6621)	-12.2	-11.6	-12.2	-12.0	-12.2	Val137	
96	BA_12100Z1	-12.1	-12.1	-11.4	-11.9	-12.1	Val137	
95	BA_12100E	-12.1	-11.1	-12.0	-11.7	-12.1	Val137	
71	Urdamycin_F	-11.9	-12.0	-11.9	-12.0	-12.0	Val137	
151	Saquayamycin_K4	-11.9	-11.9	-11.9	-11.9	-11.9	Val137	
25	Saprolmycin_A	-11.8	-11.8	-11.8	-11.8	-11.8	Val137	
98	BA_12100Z3	-11.2	-11.7	-11.8	-11.6	-11.8	Val137	
1	Marangucycline_A	-11.7	-10.4	-11.7	-11.3	-11.7	Val137	
139	Kerriamycin_A	-11.6	-10.5	-11.7	-11.3	-11.7	Val137	
		10	9	15	9	13	Val137	
		Ala134	Ala134	Ala134	Ala134	Ala134	Val137	
		Asn168	Asn168	Asn168	Asn168	Asn168	Val137	
		Gln165	Gln165	Gln165	Gln165	Gln165	Val137	
		Gly138	Gly138	Gly138	Gly138	Gly138	Val137	
		Gly161	Gly161	Gly161	Gly161	Gly161	Val137	
		His18	His18	His18	His18	His18	Val137	
		His18	His18	His18	His18	His18	Val137	
		Ile241	Ile241	Ile241	Ile241	Ile241	Val137	
		Leu141	Leu141	Leu141	Leu141	Leu141	Val137	
		Leu141	Leu141	Leu141	Leu141	Leu141	Val137	
		Leu145	Leu145	Leu145	Leu145	Leu145	Val137	
		Leu160	Leu160	Leu160	Leu160	Leu160	Val137	
		Leu164	Leu164	Leu164	Leu164	Leu164	Val137	
		Phe22	Phe22	Phe22	Phe22	Phe22	Val137	
		Phe22	Phe22	Phe22	Phe22	Phe22	Val137	
		Phe26	Phe26	Phe26	Phe26	Phe26	Val137	
		Val137	Val137	Val137	Val137	Val137	Val137	
		Arg181	Arg171	Arg181	Arg171	Arg181	Val137	
		Arg45	Arg181	Arg45	Arg181	Arg45	Val137	
		Asp114	Arg265	Arg265	Asn179	Arg265	Val137	
		Asp176	Arg45	Arg45	Asn179	Arg45	Val137	
		Asp48	Asn179	Asn179	Asp114	Asp114	Val137	
		Asp52	Asp114	Asp114	Asp48	Asp48	Val137	
		Gln115	Asp172	Asp48	Gln115	Asp52	Val137	
		Gly55	Gln115	Asp176	Asp52	Ile51	Val137	
		Ile51	Gly55	Asp48	Cys44	Ile51	Val137	
		Tyr129	Ile51	Asp52	Gln115	Tyr129	Val137	
		Tyr183	Tyr129	Tyr129	Tyr183	Tyr129	Val137	
		Tyr41	Tyr183	Tyr183	Val111	Tyr41	Val137	
		Val133	Tyr248	Tyr248	Ile51	Val111	Val137	
			Tyr41	Tyr41	Val133		Val137	
			Val133	Val111			Val137	
			Val137	Val133			Val137	

APPENDIX I
Docking score and interacted residue of protein (3ACW)
with the Angucycline Group Antibiotics (ligands), and the arrangements are according to dock scores (kcal/mol) (continue)

#	Compound Name Docked to Protein (3ACW)	Run1	Run2	Run3	AV.	Dock	Residues that interacts with the original ligand & Residues that interacts with the ligands	Residues that NOT interacts with the original ligand
2	Marangucycline_B	-11.3	-11.2	-11.2	-11.3	-11.3	Val137	
26	Saprolmycin_B	-11.2	-11.2	-11.2	-11.2	-11.2		
152	Saquayamycin_K7	-11.2	-11.2	-11.2	-11.2	-11.2	Val137	
27	Saprolmycin_C	-11.0	-11.0	-11.1	-11.0	-11.1	Val137	
37	Landomycin_Z	-11.0	-11.0	-10.2	-11.0	-11.0	Val137	
124	Landomycin_C	-9.7	-11.0	-9.7	-10.1	-11.0		Val268
134	PI_1894B (Vineomycin A) (OS-4742A1)	-10.9	-10.9	-10.8	-10.9	-10.9	Val137	
19	Fradimycin_C	-10.9	-10.9	-10.8	-10.9	-10.9	Val137	
13	Waldiomycin	-10.9	-10.9	-10.7	-10.8	-10.9	Val137	
97	BA_12100Z2	-10.9	-10.6	-10.9	-10.8	-10.9	Val137	
83	Saquayamycin_A1	-10.8	-10.8	-10.8	-10.8	-10.8	Val137	
		7	3	7	4	14		
		Asn168	Asn168	Asn168	Ala157	Ala134		
		Gln165	Gln165	Gln165	Ala157	Ala134		
					Gly138			
					Gly161			
		His18	His18	His18	His18	His18		
		Leu141	Leu141	Leu141	Leu141	Leu141		
					Leu145			
					Leu160			
		Leu164	Leu164	Leu164	Leu164	Leu164		
					Phe22			
		Phe26			Phe26			
		Val137			Phe26			
		Arg171	Arg171	Arg171	Arg171	Arg171		
		Arg181	Arg265	Arg265	Arg265	Arg265		
		Arg45	Arg45	Arg45	Arg45	Arg45		
		Asn179	Asn179	Asn179	Asn179	Asn179		
		Asp114	Asp114	Asp114	Asp114	Asp114		
		Asp172	Asp172	Asp172	Asp172	Asp172		
		Asp48	Asp48	Asp48	Asp48	Asp48		
		Tyr129	Asp52	Asp52	Asp49	Asp52		
		Tyr183	Gln175	Asp52	Asp48	Asp52		
		Tyr41	Ser19	Gln115	Lys13	Asp52		
			Ile51	Tyr129	Lys17	Gln175		
			Tyr129	Tyr183	Ile251	Asp52		
			Tyr183	Tyr248	Ile51	Gln175		
			Tyr41	Tyr41	Ile51	Tyr41		
			Val133	Val133	Lys20	Tyr129		
					Lys273	Tyr183		
					Phe267	Tyr248		
					Ser21	Tyr41		
					Tyr129			
					Tyr183			
					Val111			
					Val133			
					Val268			

APPENDIX I
Docking score and interacted residue of protein (3ACW)
with the Angucycline Group Antibiotics (ligands), and the arrangements are according to dock scores (kcal/mol) (continue)

#	Compound Name Docked to Protein (3ACW)	Run1	Run2	Run3	AV.	Dock	Residues that interacts with the original ligand & Residues that interacts with the ligands	Residues that NOT interacts with the original ligand
125	Landomycin_D	-10.0	-9.9	-9.9	-9.9	9	Val137 Val137 Val137	
3	Actinosporin_A	-9.9	-9.9	-9.9	-9.9	5	Val137 Val137 Val137	
45	Urdamycin_A	-9.9	-9.4	-9.3	-9.5	10	Val137 Val137	
78	BE_7585A	-9.9	-8.9	-9.4	-9.4	3	Val137 Val137	
141	Kerriamycin_C	-9.8	-9.6	-9.6	-9.7	11	Val137 Val137	
91	Aggreticin (OM-4842)	-9.8	-9.1	-9.8	-9.6	11	Val137 Val137	
40	JBIR_90	-9.6	-8.6	-9.7	-9.3	15	Val137 Val137	
41	JBIR_116	-8.4	-9.7	-9.7	-9.3	9	Val137 Val137	
42	JBIR_91	-9.1	-8.7	-9.7	-9.2	4	Val137 Val137	
8	Hatomarubigin_D	-9.5	-9.6	-9.6	-9.6	7	Val137 Val137	
23	L_digitoxosyl_dehydrorabelomycin	-9.1	-9.5	-9.6	-9.4	4	Val137 Val137	
							Ala134 Ala134 Ala134	
							Asn168 Asn168 Asn168	
							Gln165 Gln165 Gln165	
							Gly138 Gly138 Gly138	
							Gly161 Gly161 Gly161	
							His18 His18 His18	
							Leu141 Leu141 Leu141	
							Leu145 Leu145 Leu145	
							Leu160 Leu160 Leu160	
							Leu164 Leu164 Leu164	
							Phe22 Phe22 Phe22	
							Phe26 Phe26 Phe26	
							Val137 Val137 Val137	
							Arg171 Arg171 Arg171	
							Arg181 Arg181 Arg181	
							Arg265 Arg265 Arg265	
							Arg45 Arg45 Arg45	
							Asn179 Asn179 Asn179	
							Asp114 Asp114 Asp114	
							Asp172 Asp172 Asp172	
							Asp176 Asp176 Asp176	
							Asp48 Asp48 Asp48	
							Cys44 Cys44 Cys44	
							Gln175 Gln175 Gln175	
							Tyr248 Tyr248 Tyr248	
							Tyr41 Tyr41 Tyr41	
							Val133 Val133 Val133	
							Thr110 Thr110 Thr110	
							Tyr129 Tyr129 Tyr129	
							Tyr183 Tyr183 Tyr183	
							Tyr41 Tyr41 Tyr41	
							Val111 Val111 Val111	
							Val133 Val133 Val133	

APPENDIX I
Docking score and interacted residue of protein (3ACW)
with the Angucycline Group Antibiotics (ligands), and the arrangements are according to dock scores (kcal/mol) (continue)

#	Compound Name Docked to Protein (3ACW)	Run1	Run2	Run3	AV.	Dock	Residues that interacts with the original ligand & Residues that interacts with the ligands	Residues that NOT interacts with the original ligand
87	C104_Aglycon	-8.8	-9.4	-8.9	-9.0	-9.1	Leu141, Leu145, Leu160, Leu164, Phe22, Phe26, Val137	
149	Saquayamycin_K1	-8.6	-9.4	-9.3	-9.1	-8.9	Leu141, Leu145, Leu160, Leu164, Phe22, Phe26, Val137	
12	Warkmycin	-9.6	-8.5	-9.3	-9.1	-8.9	Leu141, Leu145, Leu160, Leu164, Phe22, Phe26, Val137	
103	De_O_acylcapoamycin	-9.0	-9.1	-9.1	-9.1	-9.1	Leu141, Leu145, Leu160, Leu164, Phe22, Phe26, Val137	
60	Sch_47555	-9.6	-9.4	-9.3	-9.1	-9.1	Leu141, Leu145, Leu160, Leu164, Phe22, Phe26, Val137	
128	Landomycin_F	-8.8	-9.4	-9.3	-9.1	-8.1	Leu141, Leu145, Leu160, Leu164, Phe22, Phe26, Val137	
73	Sakyomicin_A	-8.8	-9.4	-9.3	-9.1	-8.3	Leu141, Leu145, Leu160, Leu164, Phe22, Phe26, Val137	
11	Rubiginone_B2	-8.1	-8.6	-8.9	-8.5	-8.1	Leu141, Leu145, Leu160, Leu164, Phe22, Phe26, Val137	
34	Aquayamycin	-8.7	-8.6	-8.6	-8.6	-8.7	Leu141, Leu145, Leu160, Leu164, Phe22, Phe26, Val137	
127	TAN_1085	-8.3	-8.7	-8.3	-8.4	-8.7	Leu141, Leu145, Leu160, Leu164, Phe22, Phe26, Val137	
54	8_O_methyltetrangulol	-8.6	-8.6	-8.6	-8.6	-8.6	Leu141, Leu145, Leu160, Leu164, Phe22, Phe26, Val137	
		14	8	12	3	6	8	3
		Ala134	Ala134	Ala134			Ala134	
		Ala157					Ala157	
		Asn168	Asn168	Asn168	Asn168	Asn168	Asn168	
		Gln165	Gln165	Gln165	Gln165	Gln165	Gln165	
		Gly138					Gly138	
		Gly161	Gly161	Gly138			Gly161	
		His18	His18	Gly161	His18	His18	His18	
				His18			His18	
							Ile241	
		Leu141					Leu141	
		Leu145					Leu145	
		Leu160					Leu160	
		Leu164	Leu164	Leu164	Leu164	Leu164	Leu164	
		Phe22	Phe22	Phe22	Phe22	Phe22	Phe22	
		Phe26		Phe26			Phe26	
		Val137	Val137	Val137	Val137	Val137	Val137	
		Arg45	Arg171	Arg181	Arg181	Arg45	Arg171	
		Asp48	Arg181	Arg45	Arg45	Asp114	Phe233	
		Tyr129	Arg265	Asn179	Asn179	Arg45	Tyr41	
		Tyr41	Arg45	Asp114	Asp48	Arg45	Val133	
		Val133	Asn179	Asp176	Asp52	Cys44	Asn179	
			Asp114	Asp48	Gln115	Ile51	Asp172	
			Asp176	Asp49	Ile51	Tyr129	Asp176	
			Asp48	Asp52	Tyr129	Cys44	Tyr129	
			Asp52	Cys44	Val111	Tyr183	Val176	
			Cys44	Gln115	Val133	Tyr41	Tyr183	
			Gln175	Ile51	Tyr248		Tyr41	
			Tyr129	Tyr129			Val133	
			Tyr183	Tyr183				
			Tyr41	Tyr41				
			Val133	Val111				
			Val133	Val133				

APPENDIX I
Docking score and interacted residue of protein (3ACW)
with the Angucycline Group Antibiotics (ligands), and the arrangements are according to dock scores (kcal/mol) (continue)

#	Compound Name Docked to Protein (3ACW)	Run1	Run2	Run3	AV.	Dock	Residues that interacts with the original ligand & Residues that interacts with the ligands	Residues that NOT interacts with the original ligand
75	Sakyomicin_C	-8.2	-8.2	-8.2	-8.2	-8.2	Asn168 Asn168 Gln165 Gln165	
61	BE_23254	-8.2	-8.1	-8.2	-8.2	-8.2		
77	Sakyomicin_E	-8.2	-8.2	-8.1	-8.2	-8.2	Asn168 His18	
113	SS_228Y	-8.1	-8.2	-8.1	-8.1	-8.2	Asn168 Gln165	
133	Homo_UWM6	-8.2	-7.8	-8.2	-8.1	-8.2	Asn168 Gln165 Gly138 Gly161 Ala134	Leu141 Leu160 Leu164 Phe22 Phe26 Val137
101	Benzanthrin_B	-7.9	-8.1	-8.2	-8.1	-8.2	Asn168 Gln165 His18	Arg171
100	Benzanthrin_A	-7.8	-7.9	-8.2	-8.0	-8.2	Asn168 Gln165 His18	Arg181 Asn179 Arg265 Arg45 Asn179 Asp172 Asp176 Asp48 Gln115 Ile51 Tyr129 Val133 Ile51 Tyr129 Val133 Tyr183 Val111
14	Langkocycline_A1	-7.8	-8.2	-7.8	-7.9	-8.2	Asn168 Gln165 His18 Gly161 Ala134 Ala157	Arg181 Arg265 Arg45 Asn179 Asp172 Asp176 Asp48 Gln115 Asp49 Gln115 Tyr129 Gln115 Ile51 Ser50 Tyr129 Tyr183 Tyr183
119	X_14881_C	-7.4	-8.2	-8.2	-7.9	-8.2	Asn168 Gln165 Gly138 Gly161 Ala134 Ala157	Phe233 Tyr41 Arg181 Arg265 Arg45 Asn179 Asp172 Asp176 Asp48 Gln115 Ile169 Ile51 Tyr129 Tyr183 Val111 Val133
4	Actinosporin_B	-8.1	-8.0	-8.0	-8.0	-8.1	Asn168 Gln165 His18 His18	Arg171 Arg181 Arg265 Arg45 Asn179 Asp172 Tyr129 Asp176 Val111 Val133
114	Fridamycin_C	-8.0	-8.0	-8.0	-8.0	-8.0	Asn168 Gln165 His18	

APPENDIX I
Docking score and interacted residue of protein (3ACW)
with the Angucycline Group Antibiotics (ligands), and the arrangements are according to dock scores (kcal/mol) (continue)

#	Compound Name Docked to Protein (3ACW)	Run1	Run2	Run3	AV.	Dock	Residues that interacts with the original ligand & Residues that interacts with the ligands	Residues that NOT interacts with the original ligand
106	Elmycin_C	-7.6	-7.6	-7.8	-7.7	-7.8	Val137	Arg181, Asp114, Asp172, Asp176, Asp48, Ile51, Tyr129, Tyr183, Val111
111	Emycin_A	-7.7	-7.7	-7.7	-7.7	-7.7	Val137	
136	Urdamycin_I	-7.7	-7.7	-7.7	-7.7	-7.7	Val137	
7	Hatomarubigin_C	-7.7	-7.7	-7.7	-7.7	-7.7	Val137	
9	Hatomarubigin_E	-7.7	-7.7	-7.7	-7.7	-7.7	Val137	
63	Tetrangulol	-7.7	-7.7	-7.7	-7.7	-7.7		
64	Ochromocinone	-7.7	-7.6	-7.7	-7.7	-7.7		
6	Hatomarubigin_B	-7.7	-7.6	-7.7	-7.7	-7.7		
32	Gaudimycin_B	-7.5	-7.7	-7.7	-7.6	-7.7		
104	Elmycin_A	-7.5	-7.7	-7.5	-7.6	-7.7	Val137	
135	Yoronomycin	-7.6	-8.1	-7.5	-7.7	-7.6	Val137	

APPENDIX II
Docking score and interacted residue of protein (3W7F with MG)
with the Angucycline Group Antibiotics (ligands), and the arrangements are according to dock scores (kcal/mol)

#	Compound Name Docked to Protein 3W7F with MG	0	56	58	8	146	145	29	148	27	28	Residues that interacts with the original ligand & Residues that interacts with the ligands			Residues that NOT interacts with the original ligand
												Run1	Run2	Run3	
	3W7F_FromPDB Part A														
	Ligand of Protein														
	Moromycin_A														
	Saquayamycin_B														
	Hatomarubigin_D														
	Saquayamycin_B														
	Saquayamycin_A														
	Saprolmycin_E														
	Saquayamycin_D														
	Saprolmycin_C														
	Saprolmycin_D														
		32	23	17	21	18	21	20	16	19	17	20			
	Ala134	Ala134	Ala134	Ala134	Ala134	Ala134	Ala134	Ala134	Ala134	Ala134	Ala134	Ala134	Ala134		
	Ala157	Ala157	Ala157	Ala157		Ala157	Ala157	Ala157	Ala157	Ala157	Ala157	Ala157	Ala157		
	Arg171	Arg171			Arg171								Arg171		
	Arg265	Arg265													
	Arg45	Arg45	Arg45	Arg45	Arg45	Arg45	Arg45		Arg45	Arg45	Arg45	Arg45	Arg45		
	Asn168	Asn168	Asn168	Asn168	Asn168	Asn168	Asn168	Asn168	Asn168	Asn168	Asn168	Asn168	Asn168		
	Asp172				Asp172										
	Asp48			Asp48	Asp48	Asp48	Asp48	Asp48	Asp48	Asp48	Asp48	Asp48	Asp48		
	Cys44				Cys44					Cys44		Cys44	Cys44		
	Gln165	Gln165	Gln165	Gln165	Gln165				Gln165	Gln165	Gln165	Gln165	Gln165		
	Gly138	Gly138	Gly138	Gly138	Gly138	Gly138	Gly138	Gly138	Gly138	Gly138	Gly138	Gly138	Gly138		
	Gly161	Gly161	Gly161	Gly161	Gly161	Gly161	Gly161	Gly161	Gly161	Gly161	Gly161	Gly161	Gly161		
	His18	His18			His18										
	Leu141	Leu141	Leu141	Leu141	Leu141	Leu141	Leu141	Leu141	Leu141	Leu141	Leu141	Leu141	Leu141		
	Leu145	Leu145	Leu145	Leu145	Leu145	Leu145	Leu145	Leu145	Leu145	Leu145	Leu145	Leu145	Leu145		
	Leu160	Leu160	Leu160	Leu160	Leu160	Leu160	Leu160	Leu160	Leu160	Leu160	Leu160	Leu160	Leu160		
	Leu164	Leu164	Leu164	Leu164	Leu164	Leu164	Leu164	Leu164	Leu164	Leu164	Leu164	Leu164	Leu164		
	Lys20	Lys20													
	Met15														
	Phe22	Phe22	Phe22	Phe22	Phe22	Phe22	Phe22	Phe22	Phe22	Phe22	Phe22	Phe22	Phe22		
	Phe233	Phe233		Phe233		Phe233	Phe233	Phe233	Phe233	Phe233	Phe233	Phe233	Phe233		
	Ser19	Ser19				Ser19	Ser19	Ser19							
	Ser21	Ser21													
	Tyr248	Tyr248	Tyr248	Tyr248		Tyr248	Tyr248	Tyr248	Tyr248	Tyr248	Tyr248	Tyr248	Tyr248		
	Tyr41	Tyr41	Tyr41	Tyr41	Tyr41	Tyr41	Tyr41		Tyr41	Tyr41	Tyr41	Tyr41	Tyr41		
	Unk303			Unk303	Unk303	Unk303	Unk303	Unk303	Unk303	Unk303	Unk303	Unk303	Unk303		
	Unk304	Unk304	Unk304	Unk304	Unk304	Unk304	Unk304	Unk304	Unk304	Unk304	Unk304	Unk304	Unk304		
	Unk305	Unk305	Unk305	Unk305	Unk305	Unk305	Unk305	Unk305	Unk305	Unk305	Unk305	Unk305	Unk305		
	Val133		Val133	Val133	Val133		Val133	Val133	Val133	Val133	Val133	Val133	Val133		
	Val137		Val137	Val137	Val137	Val137	Val137	Val137	Val137	Val137	Val137	Val137	Val137		
	Unk301	Unk302	Arg181	Arg181	Arg181	Arg181	Arg181	Arg181	Arg181	Arg181	Arg181	Arg181	Arg181		
	Unk302	Asn179	Asn179	Asn179	Asn179	Asn179	Asn179	Asn179	Asn179	Asn179	Asn179	Asn179	Asn179		
	HOH401	Asp176	Asp52	Asp114	Asp114	Asp114	Asp114	Asp114	Asp114	Asp114	Asp114	Asp114	Asp114		
	HOH402	Asp52	Ile241	Ile241	Asp52	Asp52	Asp52	Asp52	Asp52	Asp52	Asp52	Asp52	Asp52		
	HOH403	Ile241	Phe26	Phe26	Ile241	Asp52	Asp52	Asp52	Ile241	Phe26	Phe26	Phe26	Phe26		
	HOH404	Phe26	Phe26	Phe26	Ile241	Ile241	Ile241	Ile241	Ile241	Ile241	Ile241	Ile241	Ile241		
	HOH405	Tyr129	Tyr183	Tyr129	Tyr129	Tyr129	Tyr129	Tyr129	Tyr129	Tyr129	Tyr129	Tyr129	Tyr129		
	HOH406	Tyr129													
	HOH407	Tyr183													
	HOH408														

APPENDIX II

**Docking score and interacted residue of protein (3W7F with MG)
with the Angucycline Group Antibiotics (ligands), and the arrangements are according to dock scores (kcal/mol) (continue)**

#	Compound Name Docked to Protein 3W7F with MG	Run1	Run2	Run3	AV	Dock	Residues that interacts with the original ligand & Residues that interacts with the ligands	Residues that NOT interacts with the original ligand
2	Marangucycline_B	-13.2	-13.3	-13.5	-13.4	-13.5	Ala134 Ala157 Arg171 Arg265 Arg45 Asn168 Asp172 Asp48 Cys44 Gln165 Gly138 Gly161 Leu141 Leu145 Leu60 Leu64 Met15 Phe22 Phe233 Ser19 Tyr248	Asp176 Gln175 Ile241 Phe26
26	Saprolmycin_B	-13.3	-12.4	-13.0	-13.3	-13.3	Ala134 Ala157 Arg171 Arg265 Arg45 Asn168 Asp172 Asp48 Cys44 Gln165 Gly138 Gly161 His18 Leu141 Leu145 Leu60 Leu64 Lys20 Met15 Phe22 Phe233 Ser19 Tyr248	Asp176 Gln175 Ile241 Phe26 Tyr129 Tyr183 Val37
143	PI_085	-12.4	-12.7	-12.7	-12.7	-13	Ala134 Ala157 Arg171 Arg265 Arg45 Asn168 Asp172 Asp48 Cys44 Gln165 Gly138 Gly161 His18 Leu141 Leu145 Leu60 Leu64 Lys20 Met15 Phe22 Phe233 Ser19 Tyr248	Asp176 Gln175 Ile241 Phe26 Tyr129 Tyr183 Val37
134	PI_1894B (Vineomycin A) (OS-4742A1)	-12.7	-12.5	-12.5	-12.5	-12.5	Ala134 Ala157 Arg171 Arg265 Arg45 Asn168 Asp172 Asp48 Cys44 Gln165 Gly138 Gly161 His18 Leu141 Leu145 Leu60 Leu64 Lys20 Met15 Phe22 Phe233 Ser19 Tyr248	Asp176 Gln175 Ile241 Phe26 Tyr129 Tyr183 Val37
38	Grincamycin	-12.5	-12.5	-12.5	-12.5	-12.5	Ala134 Ala157 Arg171 Arg265 Arg45 Asn168 Asp172 Asp48 Cys44 Gln165 Gly138 Gly161 His18 Leu141 Leu145 Leu60 Leu64 Lys20 Met15 Phe22 Phe233 Ser19 Tyr248	Asp176 Gln175 Ile241 Phe26 Tyr129 Tyr183 Val37
42	JBIR_91	-12.5	-12.5	-12.5	-12.5	-12.5	Ala134 Ala157 Arg171 Arg265 Arg45 Asn168 Asp172 Asp48 Cys44 Gln165 Gly138 Gly161 His18 Leu141 Leu145 Leu60 Leu64 Lys20 Met15 Phe22 Phe233 Ser19 Tyr248	Asp176 Gln175 Ile241 Phe26 Tyr129 Tyr183 Val37
57	Moromycin_B	-12.5	-12.5	-12.5	-12.5	-12.5	Ala134 Ala157 Arg171 Arg265 Arg45 Asn168 Asp172 Asp48 Cys44 Gln165 Gly138 Gly161 His18 Leu141 Leu145 Leu60 Leu64 Lys20 Met15 Phe22 Phe233 Ser19 Tyr248	Asp176 Gln175 Ile241 Phe26 Tyr129 Tyr183 Val37
47	N05WA963A	-12.4	-12.4	-12.5	-12.5	-12.5	Ala134 Ala157 Arg171 Arg265 Arg45 Asn168 Asp172 Asp48 Cys44 Gln165 Gly138 Gly161 His18 Leu141 Leu145 Leu60 Leu64 Lys20 Met15 Phe22 Phe233 Ser19 Tyr248	Asp176 Gln175 Ile241 Phe26 Tyr129 Tyr183 Val37
151	Saquayamycin_K4	-12.4	-12.4	-12.4	-12.4	-12.4	Ala134 Ala157 Arg171 Arg265 Arg45 Asn168 Asp172 Asp48 Cys44 Gln165 Gly138 Gly161 His18 Leu141 Leu145 Leu60 Leu64 Lys20 Met15 Phe22 Phe233 Ser19 Tyr248	Asp176 Gln175 Ile241 Phe26 Tyr129 Tyr183 Val37
25	Saprolmycin_A	-12.3	-12.3	-12.3	-12.3	-12.3	Ala134 Ala157 Arg171 Arg265 Arg45 Asn168 Asp172 Asp48 Cys44 Gln165 Gly138 Gly161 His18 Leu141 Leu145 Leu60 Leu64 Lys20 Met15 Phe22 Phe233 Ser19 Tyr248	Asp176 Gln175 Ile241 Phe26 Tyr129 Tyr183 Val37
71	Urdamycin_F	-12.2	-12.2	-12.2	-12.2	-12.3	Ala134 Ala157 Arg171 Arg265 Arg45 Asn168 Asp172 Asp48 Cys44 Gln165 Gly138 Gly161 His18 Leu141 Leu145 Leu60 Leu64 Lys20 Met15 Phe22 Phe233 Ser19 Tyr248	Asp176 Gln175 Ile241 Phe26 Tyr129 Tyr183 Val37

APPENDIX II
Docking score and interacted residue of protein (3W7F with MG)
with the Angucycline Group Antibiotics (ligands), and the arrangements are according to dock scores (kcal/mol) (continue)

#	Compound Name Docked to Protein 3W7F with MG	Run1	Run2	Run3	AV	Dock	Residues that interacts with the original ligand & Residues that interacts with the ligands	Residues that NOT interacts with the original ligand
144	PI_087	-12.2	-12.3	-12.2	-12.2	-12.3	Ala134, Ala134, Ala134, Ala134, Ala157, Ala157, Arg171, Arg171, Arg45, Arg45, Arg45, Asn168, Asn168, Asp48, Asp48, Gln165, Gln165, Gly138, Gly138, Gly161, Gly161, Leu141, Leu141, Leu145, Leu145, Leu160, Leu160, Leu164, Leu164, Phe22, Phe22, Phe233, Phe233, Tyr248, Tyr248, Tyr248, Tyr248, Tyr41, Tyr41, Tyr41, Tyr41, Unk303, Unk303, Unk304, Unk304, Unk305, Unk305, Val133, Val133, Val137, Val137	Arg181, Asn179, Asp114, Asp176, Asp52, Gln115, Gly55, Ile51, Ile57, Tyr129, Tyr183
92	BA_12100B	-11.4	-10.5	-12.3	-11.4	-12.3	Ala134, Ala134, Ala134, Ala157, Ala157, Arg171, Arg171, Arg45, Arg45, Arg45, Asn168, Asn168, Asp48, Asp48, Gln165, Gln165, Gly138, Gly138, Gly161, Gly161, Leu141, Leu141, Leu145, Leu145, Leu160, Leu160, Leu164, Leu164, Phe22, Phe22, Phe233, Phe233, Tyr248, Tyr248, Tyr248, Tyr248, Tyr41, Tyr41, Tyr41, Tyr41, Unk303, Unk303, Unk304, Unk304, Unk305, Unk305, Val133, Val133, Val137, Val137	Arg181, Asn179, Asp114, Asp176, Asp52, Gln115, Gly55, Ile51, Ile57, Tyr129, Tyr183
49	N05WA963D	-12.2	-12.2	-12.2	-12.2	-12.2	Ala134, Ala134, Ala134, Ala157, Ala157, Arg171, Arg171, Arg45, Arg45, Arg45, Asn168, Asn168, Asp48, Asp48, Gln165, Gln165, Gly138, Gly138, Gly161, Gly161, Leu141, Leu141, Leu145, Leu145, Leu160, Leu160, Leu164, Leu164, Phe22, Phe22, Phe233, Phe233, Tyr248, Tyr248, Tyr248, Tyr248, Tyr41, Tyr41, Tyr41, Tyr41, Unk303, Unk303, Unk304, Unk304, Unk305, Unk305, Val133, Val133, Val137, Val137	Arg181, Asn179, Asp114, Asp176, Asp52, Gln115, Gly55, Ile51, Ile57, Tyr129, Tyr183
141	Kerriamycin_C	-12.2	-12.2	-12.2	-12.2	-12.2	Ala134, Ala134, Ala134, Ala157, Ala157, Arg171, Arg171, Arg45, Arg45, Arg45, Asn168, Asn168, Asp48, Asp48, Gln165, Gln165, Gly138, Gly138, Gly161, Gly161, Leu141, Leu141, Leu145, Leu145, Leu160, Leu160, Leu164, Leu164, Phe22, Phe22, Phe233, Phe233, Tyr248, Tyr248, Tyr248, Tyr248, Tyr41, Tyr41, Tyr41, Tyr41, Unk303, Unk303, Unk304, Unk304, Unk305, Unk305, Val133, Val133, Val137, Val137	Arg181, Asn179, Asp114, Asp176, Asp52, Gln115, Gly55, Ile51, Ile57, Tyr129, Tyr183
69	Urdamycin_C	-10.2	-12.2	-12.1	-11.5	-12.2	Ala134, Ala134, Ala134, Ala157, Ala157, Arg171, Arg171, Arg45, Arg45, Arg45, Asn168, Asn168, Asp48, Asp48, Gln165, Gln165, Gly138, Gly138, Gly161, Gly161, Leu141, Leu141, Leu145, Leu145, Leu160, Leu160, Leu164, Leu164, Phe22, Phe22, Phe233, Phe233, Tyr248, Tyr248, Tyr248, Tyr248, Tyr41, Tyr41, Tyr41, Tyr41, Unk303, Unk303, Unk304, Unk304, Unk305, Unk305, Val133, Val133, Val137, Val137	Arg181, Asn179, Asp114, Asp176, Asp52, Gln115, Gly55, Ile51, Ile57, Tyr129, Tyr183
72	Urdamycin_G	-12.1	-12.1	-12.1	-12.1	-12.1	Ala134, Ala134, Ala134, Ala157, Ala157, Arg171, Arg171, Arg45, Arg45, Arg45, Asn168, Asn168, Asp48, Asp48, Gln165, Gln165, Gly138, Gly138, Gly161, Gly161, Leu141, Leu141, Leu145, Leu145, Leu160, Leu160, Leu164, Leu164, Phe22, Phe22, Phe233, Phe233, Tyr248, Tyr248, Tyr248, Tyr248, Tyr41, Tyr41, Tyr41, Tyr41, Unk303, Unk303, Unk304, Unk304, Unk305, Unk305, Val133, Val133, Val137, Val137	Arg181, Asn179, Asp114, Asp176, Asp52, Gln115, Gly55, Ile51, Ile57, Tyr129, Tyr183
65	PI_080 (=PI-6621)	-12.0	-11.8	-12.1	-12.0	-12.1	Ala134, Ala134, Ala134, Ala157, Ala157, Arg171, Arg171, Arg45, Arg45, Arg45, Asn168, Asn168, Asp48, Asp48, Gln165, Gln165, Gly138, Gly138, Gly161, Gly161, Leu141, Leu141, Leu145, Leu145, Leu160, Leu160, Leu164, Leu164, Phe22, Phe22, Phe233, Phe233, Tyr248, Tyr248, Tyr248, Tyr248, Tyr41, Tyr41, Tyr41, Tyr41, Unk303, Unk303, Unk304, Unk304, Unk305, Unk305, Val133, Val133, Val137, Val137	Arg181, Asn179, Asp114, Asp176, Asp52, Gln115, Gly55, Ile51, Ile57, Tyr129, Tyr183
97	BA_12100Z2	-11.4	-11.0	-12.1	-11.5	-12.1	Ala134, Ala134, Ala134, Ala157, Ala157, Arg171, Arg171, Arg45, Arg45, Arg45, Asn168, Asn168, Asp48, Asp48, Gln165, Gln165, Gly138, Gly138, Gly161, Gly161, Leu141, Leu141, Leu145, Leu145, Leu160, Leu160, Leu164, Leu164, Phe22, Phe22, Phe233, Phe233, Tyr248, Tyr248, Tyr248, Tyr248, Tyr41, Tyr41, Tyr41, Tyr41, Unk303, Unk303, Unk304, Unk304, Unk305, Unk305, Val133, Val133, Val137, Val137	Arg181, Asn179, Asp114, Asp176, Asp52, Gln115, Gly55, Ile51, Ile57, Tyr129, Tyr183
84	A_7884	-12.0	-12.0	-12.0	-12.0	-12	Ala134, Ala134, Ala134, Ala157, Ala157, Arg171, Arg171, Arg45, Arg45, Arg45, Asn168, Asn168, Asp48, Asp48, Gln165, Gln165, Gly138, Gly138, Gly161, Gly161, Leu141, Leu141, Leu145, Leu145, Leu160, Leu160, Leu164, Leu164, Phe22, Phe22, Phe233, Phe233, Tyr248, Tyr248, Tyr248, Tyr248, Tyr41, Tyr41, Tyr41, Tyr41, Unk303, Unk303, Unk304, Unk304, Unk305, Unk305, Val133, Val133, Val137, Val137	Arg181, Asn179, Asp114, Asp176, Asp52, Gln115, Gly55, Ile51, Ile57, Tyr129, Tyr183
37	Landomycin_Z	-12.0	-12.0	-12.0	-12.0	-12	Ala134, Ala134, Ala134, Ala157, Ala157, Arg171, Arg171, Arg45, Arg45, Arg45, Asn168, Asn168, Asp48, Asp48, Gln165, Gln165, Gly138, Gly138, Gly161, Gly161, Leu141, Leu141, Leu145, Leu145, Leu160, Leu160, Leu164, Leu164, Phe22, Phe22, Phe233, Phe233, Tyr248, Tyr248, Tyr248, Tyr248, Tyr41, Tyr41, Tyr41, Tyr41, Unk303, Unk303, Unk304, Unk304, Unk305, Unk305, Val133, Val133, Val137, Val137	Arg181, Asn179, Asp114, Asp176, Asp52, Gln115, Gly55, Ile51, Ile57, Tyr129, Tyr183
124	Landomycin_C	-11.6	-12.0	-11.7	-11.8	-12	Ala134, Ala134, Ala134, Ala157, Ala157, Arg171, Arg171, Arg45, Arg45, Arg45, Asn168, Asn168, Asp48, Asp48, Gln165, Gln165, Gly138, Gly138, Gly161, Gly161, Leu141, Leu141, Leu145, Leu145, Leu160, Leu160, Leu164, Leu164, Phe22, Phe22, Phe233, Phe233, Tyr248, Tyr248, Tyr248, Tyr248, Tyr41, Tyr41, Tyr41, Tyr41, Unk303, Unk303, Unk304, Unk304, Unk305, Unk305, Val133, Val133, Val137, Val137	Arg181, Asn179, Asp114, Asp176, Asp52, Gln115, Gly55, Ile51, Ile57, Tyr129, Tyr183

APPENDIX II
Docking score and interacted residue of protein (3W7F with MG)
with the Angucycline Group Antibiotics (ligands), and the arrangements are according to dock scores (kcal/mol) (continue)

#	Compound Name Docked to Protein 3W7F with MG	Run1	Run2	Run3	AV	Dock	Residues that interacts with the original ligand & Residues that interacts with the ligands	Residues that NOT interacts with the original ligand
73	Sakyomicin_A	-9.3	-9.3	-9.3	-9.3	-9.3	Ala134 Ala134 Ala134 Ala171 Arg171 Arg171 Arg265 Arg45 Arg45 Arg45 Asn168 Asn168 Asn168 Asp48 Cys44 Gln165 Gln165 Gln165 Gly138 Gly138 Gly138 Gly161 Gly161 Gly161 His18 His18 Leu141 Leu141 Leu141	Leu107 Phe26
51	Rabelomycin	-9.3	-9.3	-9.3	-9.3	-9.3	Leu164 Leu164 Leu164	
62	Tetrangomycin	-9.3	-9.3	-9.3	-9.3	-9.3	Phe22 Phe22 Phe22 Ser19 Ser19 Ser19	
55	8_O_methyl_7_deoxo_7_hydroxytetrangomycin	-9.3	-9.3	-9.3	-9.3	-9.3	ALA134 ALA134 ALA134	ALA24 ILE241 PHE26
53	8_O_methyltetrangomycin	-9.3	-9.3	-9.3	-9.3	-9.3	Ala134 Ala134 Asn168 Asn168 Asp48 Cys44 Gln165 Gln165 Gly138 Gly138 Gly161 Gly161	Val37 Phe26
154	Retymicin	-9.3	-9.3	-9.3	-9.3	-9.3	Arg45 Arg45 Asn168 Asn168 Asp48 Cys44 Gln165 Gln165 Gly138 Gly138 Gly161 Gly161 Leu141 Leu141 Leu141	Phe26 Phe26 Tyr129 Val37
50	JBIR_88	-9.3	-9.3	-9.3	-9.3	-9.3	Leu141 Leu141 Leu141 Leu141 Leu141 Leu141 Leu141	no 2D Phe26 Tyr129 Val37
68	8_O_methylrabelomycin	-9.3	-9.3	-9.3	-9.3	-9.3	Ala134 Ala134 Arg45 Arg45 Asp48 Asp48 Cys44 Cys44 Gln165 Gln165 Gly138 Gly138 Gly161 Gly161	
114	Fridamycin_C	-9.3	-9.3	-9.3	-9.3	-9.3	Ala134 Ala134 Ala157	
4	Actinosporin_B	-8.5	-9.3	-8.4	-8.7	-9.3	Arg45 Arg45 Asn168 Asn168 Asp48 Asp48	Phe26
136	Urdamycin_I	-9.2	-9.2	-9.2	-9.2	-9.2	Met15 Phe22 Phe22 Ser19 Ser19 Tyr248 Tyr248 Tyr248 Tyr41 Tyr41 Tyr41 Tyr41 Unk304 Unk304 Unk305 Unk305	Phe26 Phe26

APPENDIX II
Docking score and interacted residue of protein (3W7F with MG)
with the Angucycline Group Antibiotics (ligands), and the arrangements are according to dock scores (kcal/mol) (continue)

#	Compound Name Docked to Protein 3W7F with MG	Run1	Run2	Run3	AV	Dock	Residues that interacts with the original ligand & Residues that interacts with the ligands		Residues that NOT interacts with the original ligand
12	Warkmycin	-8.8	-8.2	-8.7	-8.7	-8.6	Arg171, Arg171, Arg265, Arg45, Arg45, Asn168, Asp172, Asp48, Cys44, Gln165, His18, Leu141, Lys20, Phe22, Ser19, Tyr248, Tyr41, Unk303, Unk304, Unk305, Val133, Val137		
100	Benzanthrין_A	-8.2	-8.3	-8.7	-8.7	-8.6	Ala134, Arg171, Arg265, Arg45, Arg45, Asn168, Asp172, Asp48, Cys44, Gln165, His18, Leu141, Lys20, Phe22, Ser19, Tyr248, Tyr41, Unk303, Unk304, Unk305, Val133, Val137		
101	Benzanthrין_B	-8.7	-8.7	-8.7	-8.7	-8.6	Ala134, Arg171, Arg265, Arg45, Arg45, Asn168, Asp172, Asp48, Cys44, Gln165, His18, Leu141, Lys20, Phe22, Ser19, Tyr248, Tyr41, Unk303, Unk304, Unk305, Val133, Val137		
20	UWM6	-8.7	-8.7	-8.7	-8.6	-8.7	Arg171, Arg171, Arg265, Arg45, Arg45, Asn168, Asp172, Asp48, Cys44, Gln165, His18, Leu141, Lys20, Phe22, Ser19, Tyr248, Tyr41, Unk303, Unk304, Unk305, Val133, Val137		
127	TAN_1085	-8.6	-8.6	-8.7	-8.6	-8.7	Arg171, Arg171, Arg265, Arg45, Arg45, Asn168, Asp172, Asp48, Cys44, Gln165, His18, Leu141, Lys20, Phe22, Ser19, Tyr248, Tyr41, Unk303, Unk304, Unk305, Val133, Val137		
15	Langkocycline_A2 (Not Docked)	0.0	0.0	0.0	0.0	0.0	Arg171, Arg171, Arg265, Arg45, Arg45, Asn168, Asp172, Asp48, Cys44, Gln165, His18, Leu141, Lys20, Phe22, Ser19, Tyr248, Tyr41, Unk303, Unk304, Unk305, Val133, Val137		
16	Langkocycline_A3 (Not Docked)	0.0	0.0	0.0	0.0	0.0	Arg171, Arg171, Arg265, Arg45, Arg45, Asn168, Asp172, Asp48, Cys44, Gln165, His18, Leu141, Lys20, Phe22, Ser19, Tyr248, Tyr41, Unk303, Unk304, Unk305, Val133, Val137		
17	Langkocycline_B1 (Not Docked)	0.0	0.0	0.0	0.0	0.0	Arg171, Arg171, Arg265, Arg45, Arg45, Asn168, Asp172, Asp48, Cys44, Gln165, His18, Leu141, Lys20, Phe22, Ser19, Tyr248, Tyr41, Unk303, Unk304, Unk305, Val133, Val137		
18	Langkocycline_B2 (Not Docked)	0.0	0.0	0.0	0.0	0.0	Arg171, Arg171, Arg265, Arg45, Arg45, Asn168, Asp172, Asp48, Cys44, Gln165, His18, Leu141, Lys20, Phe22, Ser19, Tyr248, Tyr41, Unk303, Unk304, Unk305, Val133, Val137		
43	JBIR_92 (Not Docked)	0.0	0.0	0.0	0.0	0.0	Arg171, Arg171, Arg265, Arg45, Arg45, Asn168, Asp172, Asp48, Cys44, Gln165, His18, Leu141, Lys20, Phe22, Ser19, Tyr248, Tyr41, Unk303, Unk304, Unk305, Val133, Val137		
44	JBIR_93 (Not Docked)	0.0	0.0	0.0	0.0	0.0	Arg171, Arg171, Arg265, Arg45, Arg45, Asn168, Asp172, Asp48, Cys44, Gln165, His18, Leu141, Lys20, Phe22, Ser19, Tyr248, Tyr41, Unk303, Unk304, Unk305, Val133, Val137		

APPENDIX II
Docking score and interacted residue of protein (3W7F with MG)
with the Angucycline Group Antibiotics (ligands), and the arrangements are according to dock scores (kcal/mol) (continue)

Residues that NOT interacts with the original ligand										Residues that interacts with the original ligand & Residues that interacts with the ligands										Dock	AV	Run3	Run2	Run1	Compound Name Docked to Protein 3W7F with MG	#	
																				0	0	0.0	0.0	0.0	0.0	Urdamycin_D (Not Docked)	66
																				0	0	0.0	0.0	0.0	0.0	Urdamycin_H (Not Docked)	67
																				0	0	0.0	0.0	0.0	0.0	8_O_methylurdamycin_A (Not Docked)	79
																				0	0	0.0	0.0	0.0	0.0	Urdamycin_K (Not Docked)	138
																				0	0	0.0	0.0	0.0	0.0	Saquayamycin_Z (Not Docked)	153

APPENDIX III
Docking score and interacted residue of protein (3W7F without MG)
with the Angucycline Group Antibiotics (ligands), and the arrangements are according to dock scores (kcal/mol)

#	Compound Name Docked to Protein 3W7F without MG	Ligand of Protein	Saquayamycin_K4	Moromycin_A	Saquayamycin_B	Hatomarubigin_D	Saquayamycin_B	Saquayamycin_D	Marangucycline_B	Saprolymycin_C	Grincamycin_F
0	3W7F with its ligand From PDB Part A										
151											
56											
58											
8											
146											
148											
2											
27											
39											
Run1		-8.2	-14.9	-14.4	-14.2	-13.9	-13.6	-13.5	-13.4	-13.3	-13.2
Run2		-8.3	-13.1	-14.4	-14.1	-13.9	-13.6	-13.5	-13.4	-13.4	-13.2
Run3		-7.9	-15.0	-14.4	-14.4	-13.9	-13.6	-13.6	-13.4	-13.4	-13.3
AV.		-8.1	-14.3	-14.4	-14.2	-13.9	-13.6	-13.5	-13.4	-13.4	-13.2
Dock		-8.3	-15.0	-14.4	-14.4	-13.9	-13.6	-13.6	-13.4	-13.4	-13.3
		29	21	24	18	21	15	17	20	20	15
	Ala134	Ala134	Ala134	Ala134	Ala134	Ala134	Ala134	Ala134	Ala134	Ala134	Ala134
	Ala157	Ala157	Ala157	Ala157	Ala157	Ala157	Ala157	Ala157	Ala157	Ala157	Ala157
	Arg171	Arg171	Arg171	Arg171	Arg171	Arg171	Arg171	Arg171	Arg171	Arg171	Arg171
	Arg265	Arg265	Arg265			Arg45					Arg265
	Arg45	Arg45	Arg45	Arg45	Arg45	Arg45	Arg45	Arg45	Arg45	Arg45	Arg45
	Asn168	Asn168	Asn168	Asn168	Asn168	Asn168	Asn168	Asn168	Asn168	Asn168	Asn168
	Asp172	Asp172	Asp172	Asp172	Asp172	Asp172	Asp172	Asp172	Asp172	Asp172	Asp172
	Asp48	Asp48	Asp48	Asp48	Asp48	Asp48	Asp48	Asp48	Asp48	Asp48	Asp48
	Cys44				Cys44					Cys44	Cys44
	Gln165			Gln165	Gln165	Gln165	Gln165	Gln165	Gln165	Gln165	Gln165
	Gly138		Gly138	Gly138	Gly138		Gly138	Gly138	Gly138	Gly138	Gly138
	Gly161		Gly161	Gly161	Gly161	Gly161	Gly161		Gly161	Gly161	Gly161
	His18	His18	His18								His18
	Ile241	Ile241	Ile241	Ile241				Ile241	Ile241		Ile241
	Leu141	Leu141	Leu141	Leu141	Leu141	Leu141	Leu141	Leu141	Leu141	Leu141	Leu141
	Leu145	Leu145	Leu145	Leu145	Leu145	Leu145	Leu145	Leu145	Leu145	Leu145	Leu145
	Leu160	Leu160	Leu160	Leu160	Leu160	Leu160	Leu160	Leu160	Leu160	Leu160	Leu160
	Leu164	Leu164	Leu164	Leu164	Leu164	Leu164	Leu164	Leu164	Leu164	Leu164	Leu164
	Lys20	Lys20	Lys20								Lys20
	Met15										
	Phe22	Phe22	Phe22	Phe22	Phe22	Phe22	Phe22	Phe22	Phe22	Phe22	Phe22
	Phe233	Phe233	Phe233	Phe233				Phe233	Phe22	Phe22	Phe22
	Ser19	Ser19	Ser19								Ser19
	Ser21	Ser21									Ser21
	Tyr248	Tyr248	Tyr248	Tyr248	Tyr248		Tyr248	Tyr248	Tyr248		Tyr248
	Tyr41	Tyr41	Tyr41	Tyr41	Tyr41	Tyr41	Tyr41	Tyr41	Tyr41	Tyr41	Tyr41
	Val133	Val133	Val133	Val133	Val133	Val133	Val133	Val133	Val133	Val133	Val133
	Val137	Val137	Val137	Val137	Val137	Val137		Val137	Val137	Val137	Val137
	Unk301	Arg181	Arg181	Arg181	Arg181	Arg181	Arg181	Arg181	Arg181	Arg181	Arg181
	Unk302	Asn179	Asn179	Asn179	Asn179	Asn179	Asn179	Asn179	Asn179	Asn179	Asn179
	HOH401	Asp176	Asp176	Asp176	Asp176	Asp176	Asp176	Asp176	Asp176	Asp176	Asp176
	HOH402	Asp52	Asp52	Asp52	Asp52	Asp52	Asp52	Asp52	Asp52	Asp52	Asp52
	HOH403	Glu175	Glu175	Tyr129	Asp52	Asp52	Asp52	Asp52	Tyr129	Asp52	Asp52
	HOH404			Tyr183	Tyr129	Gln115	Tyr129	Tyr129	Tyr183	Gln115	Gln115
	HOH405				Tyr183	Ile51	Tyr183				Glu175
	HOH406						Tyr129				Tyr129
	HOH407										Tyr183
	HOH408										

APPENDIX III

**Docking score and interacted residue of protein (3W7F without MG)
with the Angucycline Group Antibiotics (ligands), and the arrangements are according to dock scores (kcal/mol) (continue)**

#	Compound Name Docked to Protein 3W7F without MG	Run1	Run2	Run3	AV.	Dock	Residues that interacts with the original ligand & Residues that interacts with the ligands		Residues that NOT interacts with the original ligand	
49	N05WA963D	-12.3	-12.4	-12.3	-12.3	-12.4				
65	PI_080 (=PI-6621)	-12.4	-11.8	-11.1	-11.8	-12.4				
37	Landomycin_Z	-12.3	-11.9	-12.3	-12.2	-12.3				
155	Vineomycin_A1	-12.3	-11.0	-12.3	-11.9	-12.3				
124	Landomycin_C	-12.2	-12.2	-12.2	-12.2	-12.2				
57	Moromycin_B	-12.2	-12.2	-12.2	-12.2	-12.2				
38	Grincamycin	-12.2	-12.2	-12.1	-12.2	-12.2				
59	Sch_47554	-12.1	-12.1	-12.1	-12.1	-12.1				
35	Landomycin_A	-12.1	-12.0	-12.0	-12.0	-12.1				
147	Saquayamycin_C	-12.1	-11.5	-11.5	-11.7	-12.1				
25	Saprolmycin_A	-12.0	-12.0	-12.0	-12.0	-12.0				
142	PI_083	-12.0	-12.0	-12.0	-12.0	-12.0				
		13	21	18	17	19	10	24	14	18
		Ala134	Ala134	Ala134	Ala134	Ala134				
		Ala157	Ala157	Ala157	Ala157	Ala157				
		Arg171	Arg171	Arg171	Arg171	Arg171				
		Arg265	Arg265	Arg265	Arg265	Arg265				
		Arg45	Arg45	Arg45	Arg45	Arg45				
		Asn168	Asn168	Asn168	Asn168	Asn168				
		Asp172	Asp172	Asp172	Asp172	Asp172				
		Asp48	Asp48	Asp48	Asp48	Asp48				
		Gln165	Gln165	Gln165	Gln165	Gln165				
		Gly138	Gly138	Gly138	Gly138	Gly138				
		Gly161	Gly161	Gly161	Gly161	Gly161				
		His18	His18	His18	His18	His18				
		Ile241	Ile241	Ile241	Ile241	Ile241				
		Leu141	Leu141	Leu141	Leu141	Leu141				
		Leu145	Leu145	Leu145	Leu145	Leu145				
		Leu160	Leu160	Leu160	Leu160	Leu160				
		Leu164	Leu164	Leu164	Leu164	Leu164				
		Lys20	Lys20	Lys20	Lys20	Lys20				
		Phe22	Phe22	Phe22	Phe22	Phe22				
		Phe26	Phe26	Phe26	Phe26	Phe26				
		Ser19	Ser19	Ser19	Ser19	Ser19				
		Ser21	Ser21	Ser21	Ser21	Ser21				
		Tyr248	Tyr248	Tyr248	Tyr248	Tyr248				
		Tyr41	Tyr41	Tyr41	Tyr41	Tyr41				
		Val133	Val133	Val133	Val133	Val133				
		Val137	Val137	Val137	Val137	Val137				
		Ala244	Arg181	Arg181	Arg181	Arg181				
		Arg181	Asn179	Asn179	Asn179	Asn179				
		Asn179	Asp114	Asp114	Asp114	Asp114				
		Asp114	Asp176	Asp176	Asp176	Asp176				
		Asp176	Asp49	Asp52	Asp52	Asp52				
		Asp52	Asp52	Glu175	Glu175	Glu175				
		Tyr129	Tyr129	Ile40	Ile40	Ile40				
		Tyr183	Tyr183	Val53	Val53	Val53				
		Val53	Tyr183	Leu145	Leu145	Leu145				
		Val37	Val37	Tyr183	Tyr183	Tyr183				

APPENDIX III

**Docking score and interacted residue of protein (3W7F without MG)
with the Angucycline Group Antibiotics (ligands), and the arrangements are according to dock scores (kcal/mol) (continue)**

#	Compound Name Docked to Protein 3W7F without MG	Run1	Run2	Run3	AV.	Dock	Residues that interacts with the original ligand & Residues that interacts with the ligands																				Residues that NOT interacts with the original ligand
23	L_digitoxosyl_dehydrabelomycin	-10.5	-10.4	-10.0	-10.3	-10.3	-10.3	-10.1	-10.2	-10.1	-10.2	-10.1	-10.2	-10.1	-10.2	-10.2	-10.2	-9.9	-10.2	-10.0	-10.1						
128	Landomycin_F																										
77	Sakyomicin_E																										
75	Sakyomicin_C																										
73	Sakyomicin_A																										
125	Landomycin_D																										
40	JBIR_90																										
140	Kerriamycin_B																										
87	C104_Aglycon																										
41	JBIR_116																										
60	Sch_47555																										
19	Fradimycin_C																										
		Ala134	Ala134	Ala134	Ala134	Ala134	Ala134	Ala134	Ala134	Ala134	Ala134	Ala134	Ala134	Ala134	Ala134	Ala134	Ala134	Ala134	Ala134	Ala134	Ala134	Ala134					
		Arg171	Arg171	Arg171	Arg171	Arg171	Arg171	Arg171	Arg171	Arg171	Arg171	Arg171	Arg171	Arg171	Arg171	Arg171	Arg171	Arg171	Arg171	Arg171	Arg171	Arg171					
		Arg45	Arg45	Arg45	Arg45	Arg45	Arg45	Arg45	Arg45	Arg45	Arg45	Arg45	Arg45	Arg45	Arg45	Arg45	Arg45	Arg45	Arg45	Arg45	Arg45	Arg45					
		Asn168	Asn168	Asn168	Asn168	Asn168	Asn168	Asn168	Asn168	Asn168	Asn168	Asn168	Asn168	Asn168	Asn168	Asn168	Asn168	Asn168	Asn168	Asn168	Asn168	Asn168					
		Asp172	Asp172	Asp172	Asp172	Asp172	Asp172	Asp172	Asp172	Asp172	Asp172	Asp172	Asp172	Asp172	Asp172	Asp172	Asp172	Asp172	Asp172	Asp172	Asp172	Asp172					
		Asp48	Asp48	Asp48	Asp48	Asp48	Asp48	Asp48	Asp48	Asp48	Asp48	Asp48	Asp48	Asp48	Asp48	Asp48	Asp48	Asp48	Asp48	Asp48	Asp48	Asp48					
		Cys44	Cys44	Cys44	Cys44	Cys44	Cys44	Cys44	Cys44	Cys44	Cys44	Cys44	Cys44	Cys44	Cys44	Cys44	Cys44	Cys44	Cys44	Cys44	Cys44	Cys44					
		Gln165	Gln165	Gln165	Gln165	Gln165	Gln165	Gln165	Gln165	Gln165	Gln165	Gln165	Gln165	Gln165	Gln165	Gln165	Gln165	Gln165	Gln165	Gln165	Gln165	Gln165					
		Gly138	Gly138	Gly138	Gly138	Gly138	Gly138	Gly138	Gly138	Gly138	Gly138	Gly138	Gly138	Gly138	Gly138	Gly138	Gly138	Gly138	Gly138	Gly138	Gly138	Gly138					
		Gly161	Gly161	Gly161	Gly161	Gly161	Gly161	Gly161	Gly161	Gly161	Gly161	Gly161	Gly161	Gly161	Gly161	Gly161	Gly161	Gly161	Gly161	Gly161	Gly161	Gly161					
		His18	His18	His18	His18	His18	His18	His18	His18	His18	His18	His18	His18	His18	His18	His18	His18	His18	His18	His18	His18	His18					
		Leu141	Leu141	Leu141	Leu141	Leu141	Leu141	Leu141	Leu141	Leu141	Leu141	Leu141	Leu141	Leu141	Leu141	Leu141	Leu141	Leu141	Leu141	Leu141	Leu141	Leu141					
		Leu164	Leu164	Leu164	Leu164	Leu164	Leu164	Leu164	Leu164	Leu164	Leu164	Leu164	Leu164	Leu164	Leu164	Leu164	Leu164	Leu164	Leu164	Leu164	Leu164	Leu164					
		Lys20	Lys20	Lys20	Lys20	Lys20	Lys20	Lys20	Lys20	Lys20	Lys20	Lys20	Lys20	Lys20	Lys20	Lys20	Lys20	Lys20	Lys20	Lys20	Lys20	Lys20					
		Met15	Met15	Met15	Met15	Met15	Met15	Met15	Met15	Met15	Met15	Met15	Met15	Met15	Met15	Met15	Met15	Met15	Met15	Met15	Met15	Met15					
		Phe22	Phe22	Phe22	Phe22	Phe22	Phe22	Phe22	Phe22	Phe22	Phe22	Phe22	Phe22	Phe22	Phe22	Phe22	Phe22	Phe22	Phe22	Phe22	Phe22	Phe22					
		Phe26	Phe26	Phe26	Phe26	Phe26	Phe26	Phe26	Phe26	Phe26	Phe26	Phe26	Phe26	Phe26	Phe26	Phe26	Phe26	Phe26	Phe26	Phe26	Phe26	Phe26					
		Ser19	Ser19	Ser19	Ser19	Ser19	Ser19	Ser19	Ser19	Ser19	Ser19	Ser19	Ser19	Ser19	Ser19	Ser19	Ser19	Ser19	Ser19	Ser19	Ser19	Ser19					
		Tyr248	Tyr248	Tyr248	Tyr248	Tyr248	Tyr248	Tyr248	Tyr248	Tyr248	Tyr248	Tyr248	Tyr248	Tyr248	Tyr248	Tyr248	Tyr248	Tyr248	Tyr248	Tyr248	Tyr248	Tyr248					
		Tyr41	Tyr41	Tyr41	Tyr41	Tyr41	Tyr41	Tyr41	Tyr41	Tyr41	Tyr41	Tyr41	Tyr41	Tyr41	Tyr41	Tyr41	Tyr41	Tyr41	Tyr41	Tyr41	Tyr41	Tyr41					
		Val133	Val133	Val133	Val133	Val133	Val133	Val133	Val133	Val133	Val133	Val133	Val133	Val133	Val133	Val133	Val133	Val133	Val133	Val133	Val133	Val133					
		Val137	Val137	Val137	Val137	Val137	Val137	Val137	Val137	Val137	Val137	Val137	Val137	Val137	Val137	Val137	Val137	Val137	Val137	Val137	Val137	Val137					
		Asp176	Leu107	Ile40	Tyr129																						
		Ile169	Tyr129	Leu107																							
		Tyr129	Val111	Val37																							
		Tyr183																									

APPENDIX III
Docking score and interacted residue of protein (3W7F without MG)
with the Angucycline Group Antibiotics (ligands), and the arrangements are according to dock scores (kcal/mol) (continue)

#	Compound Name Docked to Protein 3W7F without MG	Run1	Run2	Run3	AV.	Dock	Residues that interacts with the original ligand & Residues that interacts with the ligands	Residues that NOT interacts with the original ligand
32	Gaudimycin_B	-9.6	-9.6	-9.6	-9.6	15	Ala134, Ala134, Ala157, Arg171, Arg45, Asn168, Asp48, Cys44, Gln165, Gly138, Gly161, His18, Ile241, Leu141, Leu145, Leu160, Leu164, Met15, Phe22, Phe26, Ser19, Tyr248, Tyr41, Tyr41, Val133, Val137	Ala244, Ile40, Leu107, Val37
115	Fujianmycin_A	-9.6	-9.6	-9.6	-9.6	12	Ala134, Ala134, Ala157, Arg171, Arg45, Asn168, Asp48, Cys44, Gln165, Gly138, Gly161, His18, Ile241, Leu141, Leu145, Leu160, Leu164, Met15, Phe22, Phe26, Ser19, Tyr248, Tyr41, Tyr41, Val133, Val137	Ala244, Ile40, Leu107, Val37
61	BE_23254	-9.5	-9.5	-9.5	-9.5	12	Ala134, Ala134, Ala157, Arg171, Arg45, Asn168, Asp48, Cys44, Gln165, Gly138, Gly161, His18, Ile241, Leu141, Leu145, Leu160, Leu164, Met15, Phe22, Phe26, Ser19, Tyr248, Tyr41, Tyr41, Val133, Val137	Ala244, Ile40, Leu107, Val37
104	Elmycin_A	-9.5	-9.5	-9.5	-9.5	10	Ala134, Ala134, Ala157, Arg171, Arg45, Asn168, Asp48, Cys44, Gln165, Gly138, Gly161, His18, Ile241, Leu141, Leu145, Leu160, Leu164, Met15, Phe22, Phe26, Ser19, Tyr248, Tyr41, Tyr41, Val133, Val137	Ala244, Ile40, Leu107, Val37
81	Tetrangulol methyl ether	-9.6	-9.6	-9.6	-9.6	11	Ala134, Ala134, Ala157, Arg171, Arg45, Asn168, Asp48, Cys44, Gln165, Gly138, Gly161, His18, Ile241, Leu141, Leu145, Leu160, Leu164, Met15, Phe22, Phe26, Ser19, Tyr248, Tyr41, Tyr41, Val133, Val137	Ala244, Ile40, Leu107, Val37
52	Mayamycin	-9.5	-9.5	-9.5	-9.5	11	Ala134, Ala134, Ala157, Arg171, Arg45, Asn168, Asp48, Cys44, Gln165, Gly138, Gly161, His18, Ile241, Leu141, Leu145, Leu160, Leu164, Met15, Phe22, Phe26, Ser19, Tyr248, Tyr41, Tyr41, Val133, Val137	Ala244, Ile40, Leu107, Val37
11	Rubiginone_B2	-9.5	-9.5	-9.5	-9.5	14	Ala134, Ala134, Ala157, Arg171, Arg45, Asn168, Asp48, Cys44, Gln165, Gly138, Gly161, His18, Ile241, Leu141, Leu145, Leu160, Leu164, Met15, Phe22, Phe26, Ser19, Tyr248, Tyr41, Tyr41, Val133, Val137	Ala244, Ile40, Leu107, Val37
108	Elmycin_E	-9.5	-9.5	-9.5	-9.5	14	Ala134, Ala134, Ala157, Arg171, Arg45, Asn168, Asp48, Cys44, Gln165, Gly138, Gly161, His18, Ile241, Leu141, Leu145, Leu160, Leu164, Met15, Phe22, Phe26, Ser19, Tyr248, Tyr41, Tyr41, Val133, Val137	Ala244, Ile40, Leu107, Val37
119	X_14881_C	-9.5	-9.5	-9.5	-9.5	14	Ala134, Ala134, Ala157, Arg171, Arg45, Asn168, Asp48, Cys44, Gln165, Gly138, Gly161, His18, Ile241, Leu141, Leu145, Leu160, Leu164, Met15, Phe22, Phe26, Ser19, Tyr248, Tyr41, Tyr41, Val133, Val137	Ala244, Ile40, Leu107, Val37
9	Hatomarubigin_E	-9.5	-9.5	-9.5	-9.5	13	Ala134, Ala134, Ala157, Arg171, Arg45, Asn168, Asp48, Cys44, Gln165, Gly138, Gly161, His18, Ile241, Leu141, Leu145, Leu160, Leu164, Met15, Phe22, Phe26, Ser19, Tyr248, Tyr41, Tyr41, Val133, Val137	Ala244, Ile40, Leu107, Val37
110	X_14881_D	-9.5	-9.5	-9.5	-9.5	13	Ala134, Ala134, Ala157, Arg171, Arg45, Asn168, Asp48, Cys44, Gln165, Gly138, Gly161, His18, Ile241, Leu141, Leu145, Leu160, Leu164, Met15, Phe22, Phe26, Ser19, Tyr248, Tyr41, Tyr41, Val133, Val137	Ala244, Ile40, Leu107, Val37
133	Homo_UWM6	-9.5	-9.5	-9.5	-9.5	13	Ala134, Ala134, Ala157, Arg171, Arg45, Asn168, Asp48, Cys44, Gln165, Gly138, Gly161, His18, Ile241, Leu141, Leu145, Leu160, Leu164, Met15, Phe22, Phe26, Ser19, Tyr248, Tyr41, Tyr41, Val133, Val137	Ala244, Ile40, Leu107, Val37

APPENDIX III

**Docking score and interacted residue of protein (3W7F without MG)
with the Angucycline Group Antibiotics (ligands), and the arrangements are according to dock scores (kcal/mol) (continue)**

#	Compound Name Docked to Protein 3W7F without MG	Residues that interacts with the original ligand & Residues that interacts with the ligands														Residues that NOT interacts with the original ligand																		
		Run1	Run2	Run3	AV.	Dock	Ala134	Ala157	Arg171	Arg265	Arg45	Asn168	Asn168	Asp172	Cys44	Gln165	Gln138	Gly161	His18	Leu141	Leu145	Leu160	Leu164	Lys20	Met15	Phe22	Phe26	Ser19	Tyr248	Tyr41	Val133	Val137	Val137	Val137
157	Azicemicin_B	-9.0	-9.0	-9.0	-9.0	-9.0	Ala134	Ala157	Arg171	Arg265	Arg45	Asn168	Asn168	Asp172		Gln165	Gln138	Gly161	His18	Leu141	Leu145	Leu160	Leu164	Lys20	Met15	Phe22	Phe26	Ser19	Tyr248	Tyr41	Val133	Val137	Val137	Val137
136	Urdamycin_I	-9.0	-9.0	-9.0	-9.0	-9.0	Ala134	Ala157	Arg171	Arg265	Arg45	Asn168	Asn168	Asp172		Gln165	Gln138	Gly161	His18	Leu141	Leu145	Leu160	Leu164	Lys20	Met15	Phe22	Phe26	Ser19	Tyr248	Tyr41	Val133	Val137	Val137	Val137
121	X_14881_A	-8.8	-8.8	-9.0	-8.9	-9.0	Ala134	Ala157	Arg171	Arg265	Arg45	Asn168	Asn168	Asp172	Cys44	Gln165	Gln138	Gly161	His18	Leu141	Leu145	Leu160	Leu164	Lys20	Met15	Phe22	Phe26	Ser19	Tyr248	Tyr41	Val133	Val137	Val137	Val137
80	Seitomycin	-8.8	-8.8	-8.8	-8.8	-8.8	Ala134	Ala157	Arg171	Arg265	Arg45	Asn168	Asn168	Asp172		Gln165	Gln138	Gly161	His18	Leu141	Leu145	Leu160	Leu164	Lys20	Met15	Phe22	Phe26	Ser19	Tyr248	Tyr41	Val133	Val137	Val137	Val137
4	Actinosporin_B	-8.7	-8.7	-8.5	-8.6	-8.7	Ala134	Ala157	Arg171	Arg265	Arg45	Asn168	Asn168	Asp172		Gln165	Gln138	Gly161	His18	Leu141	Leu145	Leu160	Leu164	Lys20	Met15	Phe22	Phe26	Ser19	Tyr248	Tyr41	Val133	Val137	Val137	Val137
15	Langkocycline_A2 (Not Docked)	0.0	0.0	0.0	0.0	0.0	Ala134	Ala157	Arg171	Arg265	Arg45	Asn168	Asn168	Asp172		Gln165	Gln138	Gly161	His18	Leu141	Leu145	Leu160	Leu164	Lys20	Met15	Phe22	Phe26	Ser19	Tyr248	Tyr41	Val133	Val137	Val137	Val137
16	Langkocycline_A3 (Not Docked)	0.0	0.0	0.0	0.0	0.0	Ala134	Ala157	Arg171	Arg265	Arg45	Asn168	Asn168	Asp172		Gln165	Gln138	Gly161	His18	Leu141	Leu145	Leu160	Leu164	Lys20	Met15	Phe22	Phe26	Ser19	Tyr248	Tyr41	Val133	Val137	Val137	Val137
17	Langkocycline_B1 (Not Docked)	0.0	0.0	0.0	0.0	0.0	Ala134	Ala157	Arg171	Arg265	Arg45	Asn168	Asn168	Asp172		Gln165	Gln138	Gly161	His18	Leu141	Leu145	Leu160	Leu164	Lys20	Met15	Phe22	Phe26	Ser19	Tyr248	Tyr41	Val133	Val137	Val137	Val137
18	Langkocycline_B2 (Not Docked)	0.0	0.0	0.0	0.0	0.0	Ala134	Ala157	Arg171	Arg265	Arg45	Asn168	Asn168	Asp172		Gln165	Gln138	Gly161	His18	Leu141	Leu145	Leu160	Leu164	Lys20	Met15	Phe22	Phe26	Ser19	Tyr248	Tyr41	Val133	Val137	Val137	Val137
43	JBIR_92 (Not Docked)	0.0	0.0	0.0	0.0	0.0	Ala134	Ala157	Arg171	Arg265	Arg45	Asn168	Asn168	Asp172		Gln165	Gln138	Gly161	His18	Leu141	Leu145	Leu160	Leu164	Lys20	Met15	Phe22	Phe26	Ser19	Tyr248	Tyr41	Val133	Val137	Val137	Val137
44	JBIR_93 (Not Docked)	0.0	0.0	0.0	0.0	0.0	Ala134	Ala157	Arg171	Arg265	Arg45	Asn168	Asn168	Asp172		Gln165	Gln138	Gly161	His18	Leu141	Leu145	Leu160	Leu164	Lys20	Met15	Phe22	Phe26	Ser19	Tyr248	Tyr41	Val133	Val137	Val137	Val137
66	Urdamycin_D (Not Docked)	0.0	0.0	0.0	0.0	0.0	Ala134	Ala157	Arg171	Arg265	Arg45	Asn168	Asn168	Asp172		Gln165	Gln138	Gly161	His18	Leu141	Leu145	Leu160	Leu164	Lys20	Met15	Phe22	Phe26	Ser19	Tyr248	Tyr41	Val133	Val137	Val137	Val137

APPENDIX III
Docking score and interacted residue of protein (3W7F without MG)
with the Angucycline Group Antibiotics (ligands), and the arrangements are according to dock scores (kcal/mol) (continue)

Residues that NOT interacts with the original ligand										Residues that interacts with the original ligand & Residues that interacts with the ligands										Dock	AV.	Run3	Run2	Run1	Compound Name Docked to Protein 3W7F without MG	#	
																					0	0.0	0.0	0.0	0.0	Urdamycin_H (Not Docked)	67
																					0	0.0	0.0	0.0	0.0	8_O_methylurdamycin_A (Not Docked)	79
																					0	0.0	0.0	0.0	0.0	Urdamycin_K (Not Docked)	138
																					0	0.0	0.0	0.0	0.0	Saquayamycin_Z (Not Docked)	153

APPENDIX IV

The 2D Interactions Between Docked Angucycline Group Antibiotics (Ligands) with Proteins

Code	Name	3W7F With MG	3W7F Without MG	3ACW
		Electrostatic, Van der Waals,		Covalent Bond, Water, Metal
	Protein & Ligand From PDB			
0	Ligand of protein (Docked Ligand)			
1	Marangucycline_A			
2	Marangucycline_B			
3	Actinosporin_A			

APPENDIX IV

The 2D Interactions Between Docked Angucycline Group Antibiotics (Ligands) with Proteins (continue)

Code	Name	3W7F With MG	3W7F Without MG	3ACW
			Electrostatic, Van der Waals,	Covalent Bond, Water, Metal
8	Hatamarubigin_D			
9	Hatamarubigin_E			
10	Hatamarubigin_F			
11	Rubiginone_B2			

APPENDIX IV

The 2D Interactions Between Docked Angucycline Group Antibiotics (Ligands) with Proteins (continue)

Code	Name	3W7F With MG	3W7F Without MG	3ACW
			Electrostatic, Van der Waals,	Covalent Bond, Water, Metal
12	Warkmycin			
13	Waldiomycin			
14	Langkocycline_A1			
15	Langkocycline_A2	Not Docked	Not Docked	Not Docked
16	Langkocycline_A3	Not Docked	Not Docked	Not Docked
17	Langkocycline_B1	Not Docked	Not Docked	Not Docked
18	Langkocycline_B2	Not Docked	Not Docked	Not Docked

APPENDIX IV

The 2D Interactions Between Docked Angucycline Group Antibiotics (Ligands) with Proteins (continue)

Code	Name	3W7F With MG	3W7F Without MG	3ACW
			Electrostatic, Van der Waals,	Covalent Bond, Water, Metal
19	Fradimycin_C			
20	UWM6			
21	Jadomycin_B			

APPENDIX IV

The 2D Interactions Between Docked Angucycline Group Antibiotics (Ligands) with Proteins (continue)

Code	Name	3W7F With MG	3W7F Without MG	3ACW
			Electrostatic, Van der Waals,	Covalent Bond, Water, Metal
22	Jadomycin_A			
23	L_digitoxosyl_dehydrabelomycin			
24	Dehydrabelomycin			
25	Saprolmycin_A			

APPENDIX IV

The 2D Interactions Between Docked Angucycline Group Antibiotics (Ligands) with Proteins (continue)

Code	Name	3W7F With MG	3W7F Without MG	3ACW
			Electrostatic, Van der Waals,	Covalent Bond, Water, Metal
26	Saprolmycin_B			
27	Saprolmycin_C			
28	Saprolmycin_D			
29	Saprolmycin_E			
30	Saquayamycin_A 30_11.6			

APPENDIX IV

The 2D Interactions Between Docked Angucycline Group Antibiotics (Ligands) with Proteins (continue)

Code	Name	3W7F With MG	3W7F Without MG	3ACW
			Electrostatic, Van der Waals,	Covalent Bond, Water, Metal
36	Landomycin_E			
37	Landomycin_Z			
38	Grincamycin			
39	Grincamycin_F			
40	JBIR_90			

APPENDIX IV

The 2D Interactions Between Docked Angucycline Group Antibiotics (Ligands) with Proteins (continue)

Code	Name	3W7F With MG	3W7F Without MG	3ACW
			Electrostatic, Van der Waals,	Covalent Bond, Water, Metal
41	JBIR_116			
42	JBIR_91			
43	JBIR_92	Not Docked	Not Docked	Not Docked
44	JBIR_93	Not Docked	Not Docked	Not Docked
45	Urdamycin_A			

APPENDIX IV

The 2D Interactions Between Docked Angucycline Group Antibiotics (Ligands) with Proteins (continue)

Code	Name	3W7F With MG	3W7F Without MG	3ACW
			Electrostatic, Van der Waals,	Covalent Bond, Water, Metal
46	Urdamycin_B			
47	N05WA963A			
48	N05WA963B			
49	N05WA963D			
50	JBIR_88		Not Docked	Not Docked

APPENDIX IV

The 2D Interactions Between Docked Angucycline Group Antibiotics (Ligands) with Proteins (continue)

Code	Name	3W7F With MG	3W7F Without MG	3ACW
			Electrostatic, Van der Waals,	Covalent Bond, Water, Metal
54	8_O_methyltetragulol			
55	8_O_methyl_7_deoxy_7_hydroxytetragomycin			
56	Moromycin_A			
57	Moromycin_B			

APPENDIX IV

The 2D Interactions Between Docked Angucycline Group Antibiotics (Ligands) with Proteins (continue)

Code	Name	3W7F With MG	3W7F Without MG	3ACW
			Electrostatic, Van der Waals,	Covalent Bond, Water, Metal
58	Saqayamycin_B			
59	Sch_47554			
60	Sch_47555			
61	BE_23254			

APPENDIX IV

The 2D Interactions Between Docked Angucycline Group Antibiotics (Ligands) with Proteins (continue)

Code	Name	3W7F With MG	3W7F Without MG	3ACW
			Electrostatic, Van der Waals,	Covalent Bond, Water, Metal
62	Tetrangomycin			
63	Tetrangulol			
64	Ochromicinone			
65	PI_080 (=PI-6621)			
66	Urdamycin_D	Not Docked	Not Docked	Not Docked
67	Urdamycin_H	Not Docked	Not Docked	Not Docked

APPENDIX IV

The 2D Interactions Between Docked Angucycline Group Antibiotics (Ligands) with Proteins (continue)

Code	Name	3W7F With MG	3W7F Without MG	3ACW
			Electrostatic, Van der Waals,	Covalent Bond, Water, Metal
68	8-O-methylrabelomycin			
69	Urdamycin_C			Not Docked
70	Urdamycin_E			Not Docked
71	Urdamycin_F			

APPENDIX IV

The 2D Interactions Between Docked Angucycline Group Antibiotics (Ligands) with Proteins (continue)

Code	Name	3W7F With MG	3W7F Without MG	3ACW
			Electrostatic, Van der Waals,	Covalent Bond, Water, Metal
72	Urdamycin_G			
73	Sakyomicin_A			
74	Sakyomicin_B			
75	Sakyomicin_C			

APPENDIX IV

The 2D Interactions Between Docked Angucycline Group Antibiotics (Ligands) with Proteins (continue)

Code	Name	3W7F With MG	3W7F Without MG	3ACW
			Electrostatic, Van der Waals,	Covalent Bond, Water, Metal
76	Sakyomicin_D			
77	Sakyomicin_E			
78	BE_7585A			
79	8_O_methylurdamycin_A	Not Docked	Not Docked	Not Docked

APPENDIX IV

The 2D Interactions Between Docked Angucycline Group Antibiotics (Ligands) with Proteins (continue)

Code	Name	3W7F With MG	3W7F Without MG	3ACW
			Electrostatic, Van der Waals,	Covalent Bond, Water, Metal
84	A_7884			
85	Aquamycin			
86	SM_196A			
87	C104_Aglycon			

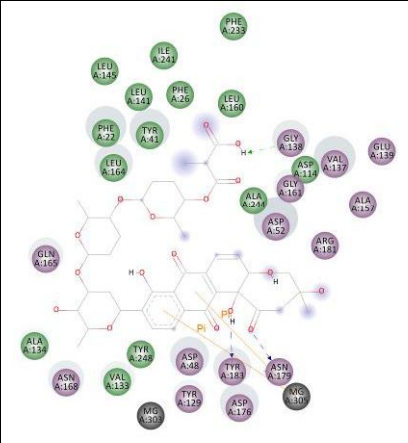
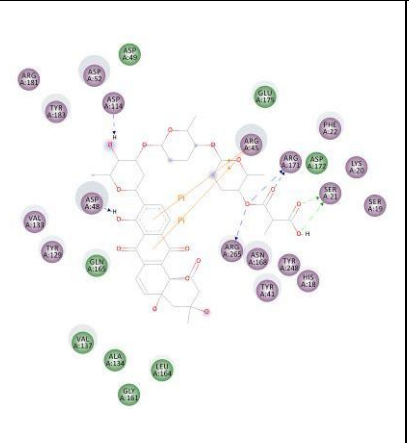
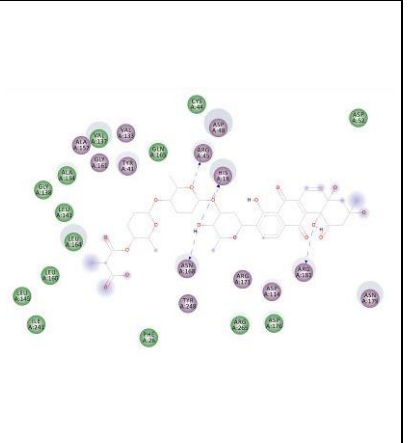
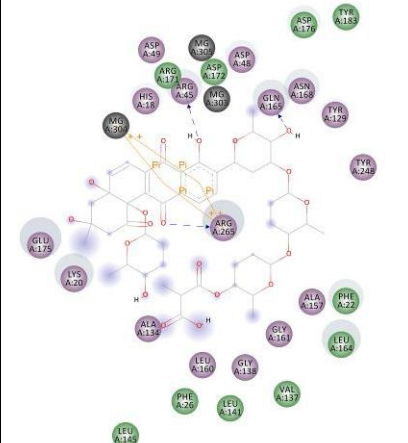
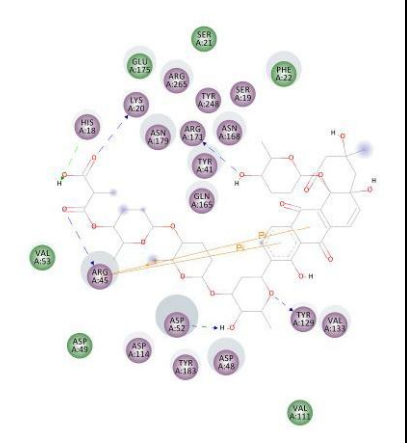
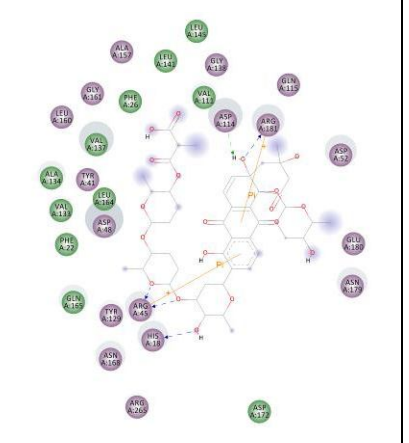
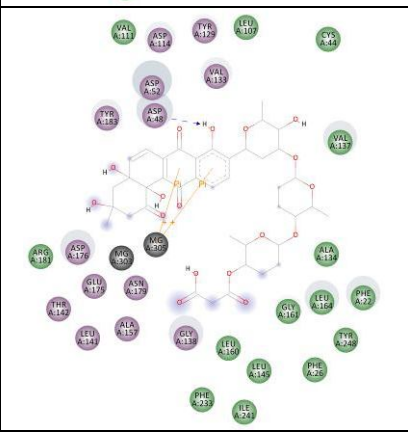
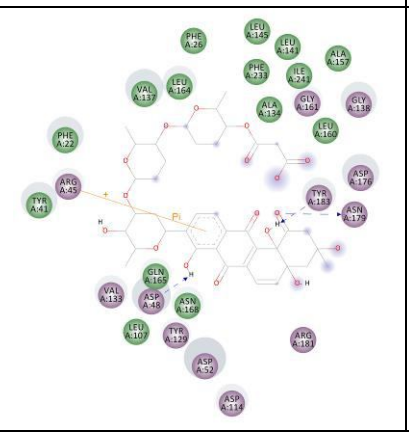
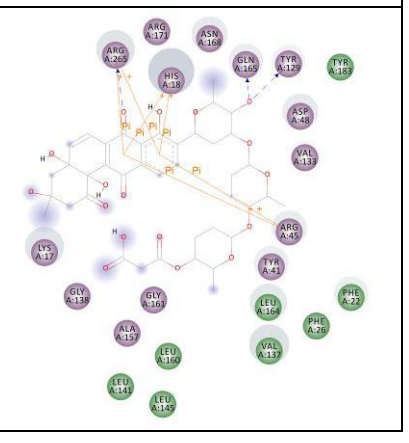
APPENDIX IV

The 2D Interactions Between Docked Angucycline Group Antibiotics (Ligands) with Proteins (continue)

Code	Name	3W7F With MG	3W7F Without MG	3ACW
			Electrostatic, Van der Waals,	Covalent Bond, Water, Metal
88	C104			
89	3_Deoxyrabelomycin			
90	6_Deoxy 8 O methyl rabelomycin			
91	Aggreticin (OM-4842)			

APPENDIX IV

The 2D Interactions Between Docked Angucycline Group Antibiotics (Ligands) with Proteins (continue)

Code	Name	3W7F With MG	3W7F Without MG	3ACW
			Electrostatic, Van der Waals,	Covalent Bond, Water, Metal
92	BA_12100B			
93	BA_12100C			
94	BA_12100D			

APPENDIX IV

The 2D Interactions Between Docked Angucycline Group Antibiotics (Ligands) with Proteins (continue)

Code	Name	3W7F With MG	3W7F Without MG	3ACW
			Electrostatic, Van der Waals,	Covalent Bond, Water, Metal
95	BA_12100E			
96	BA_12100Z1			
97	BA_12100Z2			
98	BA_12100Z3			

APPENDIX IV

The 2D Interactions Between Docked Angucycline Group Antibiotics (Ligands) with Proteins (continue)

Code	Name	3W7F With MG	3W7F Without MG	3ACW
			Electrostatic, Van der Waals,	Covalent Bond, Water, Metal
99	BA_12100MY_1			
100	Benzanthrins_A			
101	Benzanthrins_B			
102	Capoamycin			

APPENDIX IV

The 2D Interactions Between Docked Angucycline Group Antibiotics (Ligands) with Proteins (continue)

Code	Name	3W7F With MG	3W7F Without MG	3ACW
			Electrostatic, Van der Waals,	Covalent Bond, Water, Metal
103	De_O_acylcapomycin			
104	Elmycin_A			
105	Elmycin_B			
106	Elmycin_C			

APPENDIX IV

The 2D Interactions Between Docked Angucycline Group Antibiotics (Ligands) with Proteins (continue)

Code	Name	3W7F With MG	3W7F Without MG	3ACW
			Electrostatic, Van der Waals,	Covalent Bond, Water, Metal
107	Elmynin_D			
108	Elmynin_E			
109	X_14881_B			
110	X_14881_D			

APPENDIX IV

The 2D Interactions Between Docked Angucycline Group Antibiotics (Ligands) with Proteins (continue)

Code	Name	3W7F With MG	3W7F Without MG	3ACW
			Electrostatic, Van der Waals,	Covalent Bond, Water, Metal
111	Emycin_A			
112	SF_2315_A			
113	SS_228Y			
114	Fridamycin_C			

APPENDIX IV

The 2D Interactions Between Docked Angucycline Group Antibiotics (Ligands) with Proteins (continue)

Code	Name	3W7F With MG	3W7F Without MG	3ACW
			Electrostatic, Van der Waals,	Covalent Bond, Water, Metal
123	Landomycin_B			
124	Landomycin_C			
125	Landomycin_D			
126	PD116740			

APPENDIX IV

The 2D Interactions Between Docked Angucycline Group Antibiotics (Ligands) with Proteins (continue)

Code	Name	3W7F With MG	3W7F Without MG	3ACW
			Electrostatic, Van der Waals,	Covalent Bond, Water, Metal
127	TAN_1085			
128	Landomycin_F			
129	Homorabelomycin			
130	Prejadomycin			

APPENDIX IV

The 2D Interactions Between Docked Angucycline Group Antibiotics (Ligands) with Proteins (continue)

Code	Name	3W7F With MG	3W7F Without MG	3ACW
			Electrostatic, Van der Waals,	Covalent Bond, Water, Metal
131	Homoprejudomycin			
132	11_HydroxyTetrangomycin			
133	Homo_UWM6			
134	PI_1894B (Vincomicin A) (OS-4742A1)			

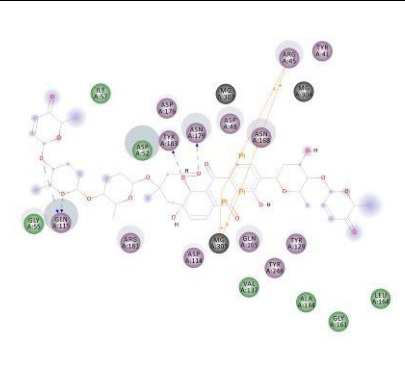
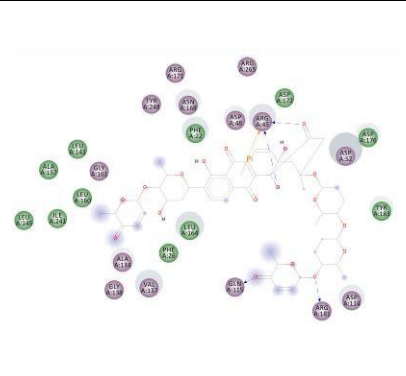
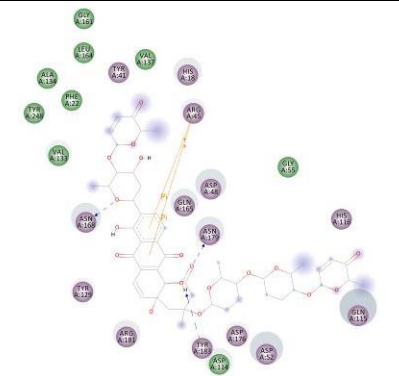
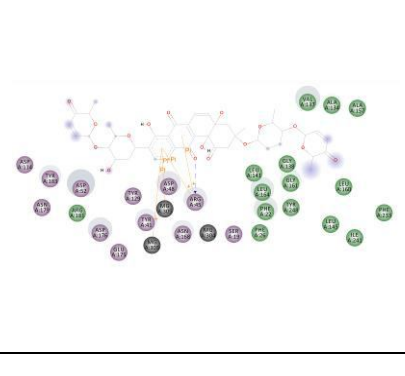
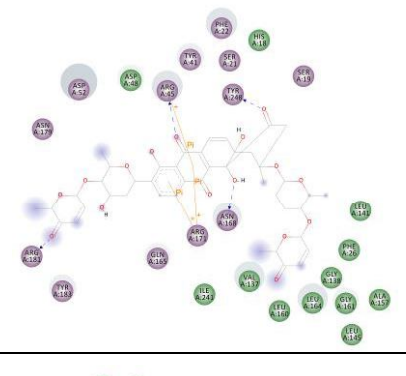
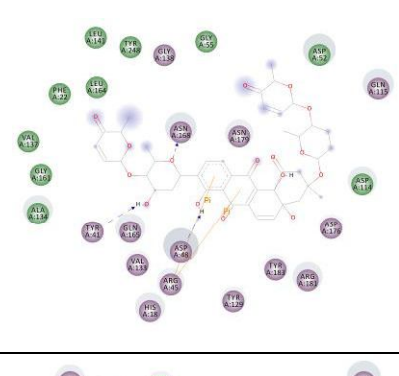
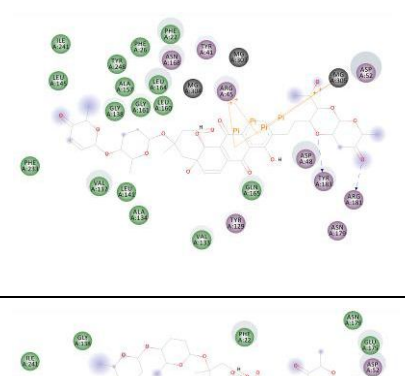
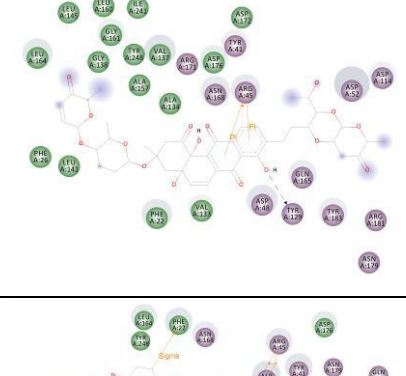
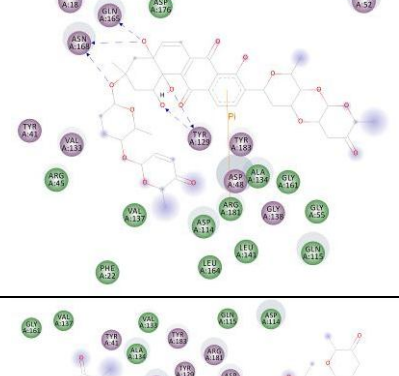
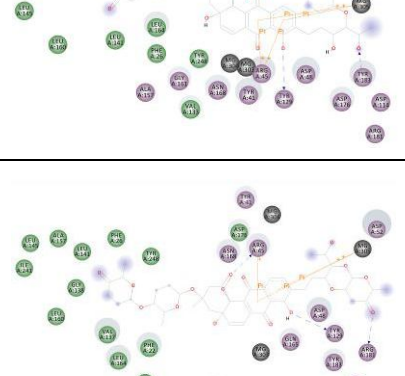
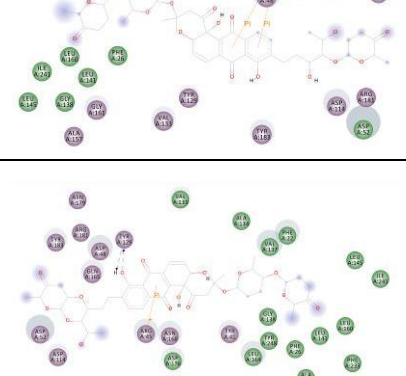
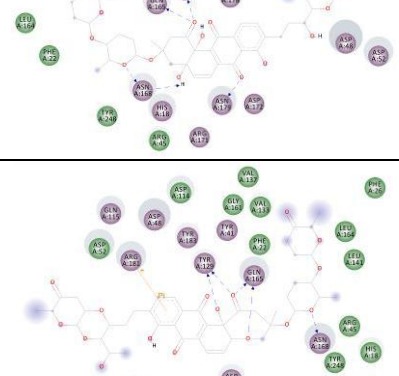
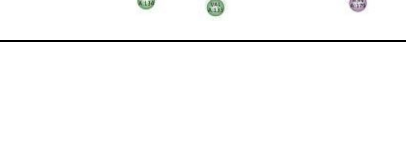


APPENDIX IV

The 2D Interactions Between Docked Angucycline Group Antibiotics (Ligands) with Proteins (continue)

Code	Name	3W7F With MG	3W7F Without MG	3ACW
			Electrostatic, Van der Waals,	Covalent Bond, Water, Metal
140	Kerriamycin_B			
141	Kerriamycin_C			
142	PI_083			
143	PI_085			

APPENDIX IV

The 2D Interactions Between Docked Angucycline Group Antibiotics (Ligands) with Proteins (continue)

Code	Name	3W7F With MG	3W7F Without MG	3ACW
			Electrostatic, Van der Waals,	Covalent Bond, Water, Metal
144	PI_087			
145	Saquayamycin_A			
146	Saquayamycin_B 146_I3.2			
147	Saquayamycin_C			
148	Saquayamycin_D			

APPENDIX IV

The 2D Interactions Between Docked Angucycline Group Antibiotics (Ligands) with Proteins (continue)

Code	Name	3W7F With MG	3W7F Without MG	3ACW
			Electrostatic, Van der Waals,	Covalent Bond, Water, Metal
149	Saquayamycin_K1			
150	Saquayamycin_K2			
151	Saquayamycin_K4			
152	Saquayamycin_K7			
153	Saquayamycin_Z	Not Docked	Not Docked	Not Docked

APPENDIX IV

The 2D Interactions Between Docked Angucycline Group Antibiotics (Ligands) with Proteins (continue)

Code	Name	3W7F With MG	3W7F Without MG	3ACW
			Electrostatic, Van der Waals,	Covalent Bond, Water, Metal
154	Retymicin			
155	Vincomycin_A1			
156	Azicemicin_A			
157	Azicemicin_B			

Curriculum Vitae

Personal Information:

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Graduation and Degrees:

2001 – 2002 (B.Sc. in Biology) College of Science, Dept. of Biology, Salahaddin University, Erbil-Iraq.
1996 – 1997 (Diploma in Physiotherapy) Erbil Technical Institute, Medical Part, Dept. of Physiotherapy.
1994 – 1995 High School Certificate, Baccalaureate Exam. for Secondary Schools (Scientific Section).

Certifications:

Jul. 2013 Certificate of International Computer Driving License (ICDL Course) Started from 03/06/2013 to 08/07/2013, Salahaddin University-Erbil, Iraq.
Sep. to Dec.2012 Microsoft Office Specialist (M.O.S.) Courses: MS-Word 2010, MS-Excel 2010, MS-Access 2010, MS-Power Point 2010, MS-Outlook 2010, and Comp TIA Network+, certificate from Information Technology Academy (ITA) KRG, Iraq.

Summary of the relevant works:

- 1- College of Medicine, Hawler Medical University, Erbil-IRAQ.
.From 11/2006 till now, Senior Biologist in Dept. of Anatomy & Histology\ Working in lab of Medical Biology as a Demonstrator.
- 2- Presidency of Hawler Medical University, Erbil-IRAQ.
.From 11/2005 to 10/2006, Biologist.
- 3- College of Nursing, University of Salahaddin, Erbil-IRAQ.
.From 2/2004 to 11/2005, Biologist \ Working in labs of Biology, Physiology, Biochemistry as a Demonstrator.
- 4- Erbil Technical Institute\ Medical Part \ Dept. of Physiotherapy, Erbil-IRAQ.
.From 7/2003 to 1/2004, Assistant Biologist\ Working in labs of Physiotherapy, Biology, and Physiology.

Language skills:

Mother Tongue Kurdish, fluency in English, Arabic, & Turkish languages.

Languages	Reading	Writing	Speaking
English	Excellent	Excellent	Good
Arabic	Excellent	Excellent	Excellent
Turkish	Good	Fair	Good



T.C.
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YÜKSEK LİSANS / DOKTORA TEZ ÇALIŞMASI İNTİHAL RAPORU
FORMU

ÖĞRENCİ BİLGİLERİ

ADI VE SOYADI	Hazem Abbas Tofiq AL-BUSTANY
ÖĞRENCİ NO	13801303
EĞİTİM - ÖĞRETİM YILI	2015-2016
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ANABİLİM DALI	Biyoloji
PROGRAM	<input checked="" type="checkbox"/> Yüksek Lisans <input type="checkbox"/> Doktora <input type="checkbox"/> Tezsiz Yüksek Lisans (Dönem Projesi)
TEZ KONUSU	SCREENING OF ANGYCYCLINE ANTIBIOTICS AS POTENTIAL DRUG CANDIDATES AGAINST MRSA BY DOCKING ANALYSIS

İNTİHAL RAPORU BİLGİLERİ

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SAYFA SAYISI	171
BENZERLİK ORANI	%15
RAPORLAMA TARİHİ	26/02/2016

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- Kabul/Onay sayfaları hariç,
- Kaynakça hariç
- Alıntılar hariç/dâhil
- Diğer

Dicle Üniversitesi Fen Bilimleri Enstitüsü Lisansüstü Programlarda Tez Çalışması İntihal Raporu Uygulama Esasları'nı inceledim ve bu Uygulama Esasları'nda belirtilen azami benzerlik oranlarına göre tez çalışmamın herhangi bir intihal içermediğini; aksinin tespit edilmesi durumunda doğabilecek her türlü hukuki sorumluluğu kabul ettiğimi ve vermiş olduğum bilgilerin doğru olduğunu beyan ederim.

Gereğini saygılarımla arz ederim.

Hazem Abbas Tofiq AL-BUSTANY

12/04/2016

Prof. Dr. Ebru İNCE
Tez Danışmanı

12/04/2016

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